

DISCUSSION GÉNÉRALE

Contexte sociétal

Les premières causes de mortalité dans le monde sont les maladies cardiovasculaires ischémiques (maladies coronariennes et accidents vasculaires cérébraux principalement). Ces pathologies se développent massivement dans le cadre de troubles métaboliques associés à l'obésité, au syndrome métabolique et au diabète de type 2 (Fornoni et al., 2005). La prévalence de ces pathologies a fortement augmenté lors de ces dernières décennies, avec en cause une alimentation trop riche en lipides et en sucres ajoutés, et également une trop forte séentarité des populations.

La modification de l'alimentation chez les populations des pays développés relève de différents facteurs sociaux-économiques qui dépassent notre propos. Cependant l'arrivée des fast food ainsi que l'augmentation de la consommation de plats industriels, peu coûteux et riches en acides gras saturés, participent à l'évolution de la prévalence de l'obésité et du syndrome métabolique (Chen et al., 2012 ; Mejova et al., 2015).

Une sensibilisation des populations au danger de ce type d'alimentation, pourrait limiter leur consommation et ainsi réduire les risques cardiovasculaires associés. Cette sensibilisation, commence par une meilleure compréhension des mécanismes d'action délétères, induits par les troubles métaboliques, liés à ces déséquilibres alimentaires, notamment sur la fonction vasculaire.

C'est pourquoi, au cours de nos différents travaux, nous avons tenté de comprendre comment la fonction vasculaire est impactée par différents désordres métaboliques au cours de stade précoces, favorisant ainsi une éventuelle intervention préventive avant l'installation de dommages irréversibles.

Cible biologique

L'intérêt scientifique est généralement porté sur la compréhension et l'explication de phénomènes responsables d'états pathologiques établis. Il semble pourtant nécessaire de tenter de comprendre l'apparition de désordres physiologiques chez des personnes asymptomatiques, afin de limiter le décours vers un état pathologie avancé.

Dans ce contexte, une partie de nos travaux s'est intéressé^{30372e} aux effets de la consommation de boissons sucrées sur la fonction vasculaire chez les personnes saines. La consommation de produits contenant des sucres ajoutés, est courante dans l'alimentation moderne. En effet, nous pouvons en retrouver dans toutes les préparations industrielles, et

particulièrement dans les boissons de type sodas, boissons énergétiques, eaux aromatisées, et même les jus de fruits. Ces boissons créent un apport considérable de sucres ajoutés à notre organisme, bien au-delà des besoins nutritionnels quotidiens. C'est pourquoi, nous observons une forte corrélation entre l'augmentation de la consommation de ces boissons et l'augmentation de la prévalence de l'obésité (Malik et al., 2006).

Différentes études ont révélé les dangers de ces sucres, notamment leur effet addictif, par une étude comparant la dépendance de rongeurs à la cocaïne et au sucre (Lenoir et al., 2007). Si les animaux ont dès le départ le choix entre les deux composants, ils vont préférer le sucre à la cocaïne. De plus, lorsqu'ils présentent du sucre à des rongeurs dépendants de la cocaïne depuis plusieurs semaines, les animaux vont orienter leur préférence, une fois encore vers le sucre, malgré la forte dépendance que crée le stupéfiant. Ainsi, en plus du plaisir lié au goût sucré sur les papilles, la consommation de sucre provoquerait une addiction. Cependant, d'autres études sont nécessaires afin de mettre en lumière les connexions nerveuses potentiellement impliquées, et évaluer si cette addiction est liée à la notion de plaisir, ou si la molécule de glucose est capable d'enclencher des mécanismes cellulaires au niveau du cortex cérébral, qui modifieraient la sensibilité des individus à ce composé. Une autre étude intéressante a montré que la consommation excessive de sucre chez des rongeurs femelles en gestation, prédisposait leur fœtus à développer un syndrome métabolique (Saad et al., 2016). De plus, il semblerait que l'association de produits sucrés et riches en lipides, potentialise leur consommation réciproque (Ishomoto et al., 2013).

Ces boissons sucrées contiennent du saccharose, constitué de 45% de glucose et 55% de fructose. Le fructose, un sucre simple contenu principalement dans les fruits, participe à l'élévation de la production de triglycérides, et donc à l'accumulation de masse grasse lorsqu'il est consommé en forte quantité (Jurgens et al., 2005 ; Bantle et al., 2000). Tandis que le glucose agit de manière néfaste sur la fonction vasculaire, et participe d'avantage à l'installation de dysfonctions endothéliales (Ceriello et al., 1996 ; Lorenzi et al., 1986). Ainsi, au cours de notre étude portant sur l'impact vasculaire d'une hyperglycémie aigüe, nous nous sommes particulièrement intéressés au rôle du glucose sur la fonction vasculaire.

L'altération de la fonction vasculaire s'apparente principalement à une dysfonction endothéliale, causée par un déséquilibre de production entre les agents vasoconstricteurs et vasodilatateurs. En effet, l'endothélium, qui pourrait bien être le plus important organe endocrine de l'organisme, libère différents agents responsables de la régulation de

l'homéostasie vasculaire, dont le NO qui joue un rôle prépondérant dans la relaxation endothélium-dépendante des vaisseaux. Ainsi, l'apparition de dysfonction endothéliale est généralement causée par une diminution de la biodisponibilité de NO, liée à une augmentation du stress oxydant (Rochette et al., 2013).

En plus d'être une source calorique importante, ces bombes sucrées modifient de manière transitoire la balance redox des cellules cibles de cette hyperglycémie. En effet comme nous l'avons décrit au cours ce travail de doctorat, et notamment dans une letter (Meziat et al., 2016) en réponse à deux commentaires réalisés sur notre méta-analyse (Loader et al., 2015), une forte concentration de glucose engendre une production excessive d'POR qui entraînent une diminution de la biodisponibilité du NO. Ce phénomène est lié à une diminution du substrat de la eNOS, la L-arginine, essentielle à la production du gazotransmetteur. En effet, il a été démontré que l'hyperglycémie, *via* une forte production d'POR, stimule l'expression d'ADMA et d'arginase, responsables de la dégradation de la L-arginine. Les POR oxydent également le cofacteur essentiel de la eNOS, le BH₄ en BH₂, qui ne peut alors plus jouer son rôle essentiel dans le couplage entre le transfert d'électrons et la synthèse de NO. La eNOS devient alors une source potentielle d'POR et participe à l'installation d'un stress oxydant. De plus, le NO est une molécule très réactive qui peut se coupler à l'anion superoxyde pour produire du peroxynitrite, un agent oxydant très puissant responsable de nombreux dégâts cellulaires irréversibles.

Ainsi, une hyperglycémie aigue causée, par exemple, par la consommation d'une boisson sucrée entraîne une altération de la fonction de relaxation liée à l'endothélium, en altérant la voie du NO. Cependant, ces altérations de la capacité de relaxation sont transitoires (Mah et al., 2012), et les mécanismes déterminant le rétablissement de la fonction endothéliale, et notamment du NO ne sont pas bien décrits. Nous pouvons émettre l'hypothèse que dans un premier temps, l'insuline permet de rétablir la glycémie, permettant ainsi de limiter la stimulation de production d'POR au niveau de la paroi vasculaire. Et dans un second temps, les mécanismes antioxydants restaurent les désordres cellulaires causés par le stress oxydant, permettant ainsi de rétablir la voie de la eNOS et donc la capacité de relaxation endothéliale des vaisseaux.

Une capacité adaptative

Un élément intéressant qui ressort de l'ensemble de ces études, est la capacité de l'endothélium à s'adapter à différents types de stress, à court ou à long terme. En effet, les

dysfonctions endothéliales causées par un stress hyperglycémique aigu, sont transitoires, et donc rapidement rétablit grâce à des mécanismes adaptatifs. En l'occurrence, la capacité de captation du glucose plasmatique par les tissus insulino-sensibles permettant de réguler rapidement la glycémie, mais également les défenses antioxydantes de l'organisme qui permettent de rétablir le déséquilibre redox cellulaire. L'endothélium produit de nombreuses enzymes antioxydantes (SOD, Catalase, glutathione peroxydase), permettant de préserver, ou du moins de restaurer, la biodisponibilité du NO. Le retour rapide de la fonction de relaxation endothéliale permet d'augmenter le flux sanguin et donc l'apport de glucose aux muscles et aux tissus adipeux, favorisant ainsi la baisse de la glycémie.

La réponse adaptative à ce type de stress définit en partie l'état de santé de la fonction vasculaire. En effet, chez des personnes insulino-résistantes ou diabétiques, ces mécanismes adaptatifs sont délétères, les tissus de stockage sont moins sensibles à la stimulation de l'insuline, et les défenses antioxydantes sont dépassées par une trop forte production d'EUR, engendrant des altérations vasculaires permanentes qui participent à l'installation de l'athérosclérose. Cette pathologie vasculaire est caractérisée par une accumulation de processus pro-oxydants et pro-inflammatoires, qui favorisent l'adhésion leucocytaire et la prolifération des cellules musculaires lisses. La formation d'AGE favorise également ce processus athérosclérotique, dont la dangerosité dépend de la rupture de la plaque d'athérome, qui peut venir obstruer des artères, et ainsi aboutir à une ischémie.

C'est pourquoi, aujourd'hui, l'évaluation de la tolérance au glucose est réalisée par un test d'hyperglycémie postprandiale, qui reflète d'avantage la capacité de l'organisme à s'adapter à un stress aigu, grâce notamment, à l'action de l'endothélium. En effet, *via* l'action de l'insuline, l'endothélium participe à la régulation de la glycémie. La stimulation de la voie du NO responsable d'un relâchement artériel, entraîne une augmentation du flux sanguin, qui permet d'augmenter la surface d'échange entre les cellules endothéliales et les molécules circulantes, favorisant ainsi la captation du glucose.

La compréhension des mécanismes biologiques demande de prendre en compte le maximum d'éléments pouvant interagir sur le phénomène étudié. Ainsi, l'étude de la fonction vasculaire ne peut se faire sans la totalité des tissus qui composent et qui participent à la régulation de l'homéostasie vasculaire.

Le modèle de rat syndrome métabolique (SMet) caractérisé au cours de l'étude 2, en accord avec certaines observations faites sur des individus obèses ou SMet (Hugget et al.,

2004 ; Agapitov et al., 2008), ne présente pas d'hypertension artérielle malgré une activité importante du SNS, et une concentration élevée de catécholamines plasmatique. De manière intéressante, notre étude révèle que ce phénomène adaptatif serait lié à une suractivation de la eNOS par un mécanisme adrénnergique, limitant ainsi l'augmentation du tonus vasculaire, et donc de la pression artérielle systémique. Ainsi, lors de phase précoce du développement du SMet, l'endothélium est capable de mettre en place des mécanismes compensatoires qui permettent de limiter les altérations liés aux désordres métaboliques induit par la pathologie. Le SNS agit également au niveau du tissu adipeux. Il régule la capacité lipolytique des adipocytes grâce à des stimulations adrénnergiques, *via* des récepteurs α -adrénnergiques anti-lipolytiques, et des récepteurs β -adrénnergiques lipolytiques. Le SNS agit également au niveau du phénotype des adipocytes, en réduisant notamment leur taille (Lee et al., 2013) et en stimulant le phénomène de « browning » au niveau des adipocytes beiges (Ye et al., 2013), via une activation des récepteurs β -adrénnergiques. Cependant, ces phénomènes devront être plus clairement décrits par des études ultérieures.

Depuis quelques années, il apparaît très clairement que le tissu adipeux périvasculaire, jusqu'alors ignoré, semble participer à la fonction biologique des vaisseaux. Il semble ainsi nécessaire de l'inclure dans l'évaluation des capacités d'adaptation vasculaire, au cours d'une phase d'installation de troubles métaboliques chroniques.

De manière intéressante, nos travaux révèlent qu'en présence de son PVAT, le muscle lisse de l'aorte de rats SMet est plus sensible au NO en comparaison aux rats Ctrl, alors que sans la présence du PVAT, la relaxation de l'artère NO-dépendante tend à être délétère. Plusieurs publications ont mis en évidence l'effet anti-contractile du PVAT sur des artères saines, par des mécanismes impliquant l'ouverture de canaux potassiques par des composés tels que le sulfure d'hydrogène (Wojcicka et al., 2010) ou bien le PAME (palmitic acid methyl ester) (Lee et al., 2011). D'autres travaux mettent en évidence que le PVAT pourrait stimuler la guanylate cyclase soluble par le peroxyde d'hydrogène (Gao et al., 2007). La majorité des équipes qui se sont intéressées à l'influence du PVAT sur la fonction vasculaire en condition pathologique ont utilisé des modèles d'animaux ou bien des tissus humains dans des états avancés d'obésité et/ou de diabète de type 2 (Virdis et al., 2014 ; Meijer et al., 2013 ; Ma et al., 2010 ; Galvez et al., 2006). Cependant, Gil Ortega et al. montrent sur des animaux soumis à un régime *high fat* court (8 semaines), une augmentation de la production de NO au

sein du PVAT-HF (Gil-Ortega et al., 2010). Ce phénomène semblerait expliqué par le rôle de la leptine.

Ainsi, lors de la phase précoce de l'installation de pathologie métabolique telle que l'obésité, le phénotype et le profil sécrétoire du PVAT semble se modifier afin de préserver la fonction de relaxation vasculaire. Nous pouvons imaginer que lors de cette phase d'adaptation, la masse du PVAT étant plus importante, les adipocytes ont potentiellement une capacité de production et de sécrétion plus élevée. Ainsi, une quantité plus importante d'agents tels que le PAME, l' H_2S , l' H_2O_2 ou la leptine, peuvent être libérés et stimuler davantage la relaxation artérielle, permettant ainsi de préserver le tonus artériel.

De plus, Soltis et Cassis, semblent montrer que le PVAT absorbe les catécholamines, limitant ainsi leur effet pro-contractile. Ceci pourrait expliquer l'absence d'hypertension artérielle chez certains sujets atteints d'obésité ou de syndrome métabolique, dont le PVAT plus épais, pourrait bloquer l'action des catécholamines (Soltis et al., 1991).

Pour mieux évaluer les effets propres des agents vasoactifs produits nous avons utilisé le secretome du PVAT de nos différents animaux. De manière très intéressante, lorsque l'on met en contact un PVAT de rat SMet a priori non délétère, avec une aorte saine, la capacité de relaxation de l'artère est altérée, et ce *via* un mécanisme a priori, stress oxydant dépendant. Ces résultats démontrent la capacité de l'artère, à s'adapter face à un stress chronique métabolique, et au-delà d'une adaptation endothéliale, on s'aperçoit que la première cible semble être le PVAT. Cette hypothèse est appuyée par la « brownisation » du tissu après les 15 semaines de régime HFS, qui semble témoigner d'une augmentation du métabolisme du tissu *via* une capacité thermogénique plus importante, permettant de transformer une plus grande quantité de substrat en énergie thermique. Malgré cette transformation phénotypique, les sécrétions du PVAT aortique de rat HFS sont délétères pour une artère saine, suggérant que la communication interne entre le PVAT et les tissus artériels est assez complexe, et que les mécanismes d'adaptation ne sont pas seulement liés à la transformation du PVAT. D'ailleurs, lorsque l'on utilise le sécrétome du PVAT Ctrl sur une artère Ctrl, la relaxation est diminuée de moitié par rapport à la capacité de relaxation de l'artère avec le PVAT laissé intact autour. L'isolation du sécrétome est essentielle à la compréhension des mécanismes impliqués dans l'effet vasoactif du PVAT sur la fonction vasculaire. Cependant la séparation des deux tissus peut altérer la communication. En effet, au niveau des petits vaisseaux, le PVAT s'imbrique dans le tissu artériel, mais la structure est plus complexe pour les gros vaisseaux, où l'aventice forme une barrière entre le tissu artériel et le PVAT. Pour

communiquer avec le tissu sous-jacent, le PVAT utilise le *vasa vasorum*, un complexe artériel composé de petites artéries et de capillaires (Brown et al., 2014 ; Ahmed et al., 2004). Ainsi, la séparation du PVAT de son tissu artériel au niveau aortique par exemple, annihile les liens entre les composantes vasculaires, et peut limiter l'action de certaines substances vasoactives possédant une demi-vie limitée. L'utilisation de lits artériels perfusés, tel que le lit artériel mésentérique, pourrait être une bonne stratégie permettant d'évaluer la stimulation de l'endothélium ou du PVAT de manière plus spécifique, en préservant les liens entre les différents tissus.

La proximité du PVAT avec le tissu artériel, et sa capacité pro-oxydative et pro-inflammatoire, sont d'un grand intérêt pour la compréhension des mécanismes à l'origine de dysfonction vasculaires, et offrent un large panel d'explorations futures.

L'exercice : une stratégie intéressante pour lutter contre les désordres métaboliques quotidiens.

L'exercice physique est aujourd'hui reconnu pour ses capacités à prévenir et corriger l'apparition de certains états pathologiques (Pedersen et al., 2015). L'OMS a d'ailleurs mis au point en 2010, les « *Recommandations mondiales en matière d'activité physique pour la santé* » afin de préconiser une prévention primaire des maladies non-transmissibles par l'exercice physique sur l'ensemble des populations. On estime que l'inactivité serait responsable de 6 à 10% des décès liés à des pathologies cardiovasculaires (Lee et al., 2012). En effet, la sédentarité est un facteur de risque indépendant du développement d'altérations métaboliques ou cardiovasculaires (Hammer et al., 2012). Ainsi, l'exercice physique est une stratégie de prise en charge globale, très intéressante pour traiter/limiter les désordres métaboliques qui caractérisent le syndrome métabolique (Martin-Cordero et al., 2011 ; Golbidi et al., 2012). De plus, il est bien démontré aujourd'hui que la pratique régulière d'exercice physique entraîne des adaptations vasculaires bénéfiques qui limitent la rigidité artérielle et les dysfonctions endothéliales liées au vieillissement (Santos-Parker et al., 2014). Ces adaptations sont majoritairement associées à l'augmentation de la protection contre le stress oxydant et les phénomènes pro-inflammatoires, qui participent à la réduction de l'épaisseur de la paroi artérielle (Santos-Parker et al., 2014 ; Ashor et al., 2014). Ces améliorations de la fonction vasculaire sont également associées à une augmentation de la

capacité de relaxation des artères, grâce à une stimulation de la voie du NO (Maiorana et al., 2003).

En accord avec la littérature, nos travaux démontrent que l'exercice permet de protéger la fonction vasculaire en amont ou bien pendant l'installation de désordre métabolique. En effet, nous avons montré qu'un protocole d'entraînement modéré permet de limiter les effets néfastes d'un stress hyperglycémique aigu sur la fonction endothéliale. Nous ne pouvons pas parler ici de correction de l'effet délétère de l'hyperglycémie aiguë, puisque ce stress altère significativement la relaxation endothélium-dépendante de nos animaux entraînés en comparaison à la réponse en normoglycémie. Cependant, l'exercice physique potentialise la réponse vasculaire, et la capacité de relaxation endothélium-dépendante est identique chez un animal sédentaire en condition de normoglycémie et un animal entraîné en hyperglycémie. Bien que les capacités de potentialisation des défenses antioxydantes de l'exercice physique sur la fonction vasculaire, ainsi que sur la voie du NO soient des mécanismes potentiellement explicatifs de ce phénomène, d'autres explorations doivent être menées pour le confirmer.

Une stratégie de prévention secondaire a été utilisée sur des animaux développant un syndrome métabolique. Chez ces animaux, l'exercice a permis d'améliorer la fonction vasculaire mais dans sa globalité, c'est-à-dire en prenant en compte l'implication vasoactive du PVAT. D'ailleurs, l'exercice dans ce modèle, semble impacter plus particulièrement le phénotype et le métabolisme du PVAT, plutôt que le tissu artériel (endothélium et muscle lisse vasculaire). Les effets de l'exercice physique sur le tissu adipeux sont aujourd'hui bien décrits, et montrent des effets bénéfiques principalement sur le tissu adipeux sous-cutané abdominal. Une étude menée sur des rats sains, n'a montré aucun impact de l'exercice sur la capacité vasoactive du PVAT, malgré une diminution de la masse de celui-ci dans le groupe de rat entraîné (Araujo et al., 2013). A notre connaissance, deux études seulement, menées par la même équipe, ont évalué ce phénomène en condition pathologique, chez des porcs atteints de dyslipidémie, mais ils ne montrent pas d'amélioration de la fonction coronarienne par l'exercice physique *via* le PVAT. Pourtant, la capacité de l'exercice physique à améliorer la fonction vasculaire, ainsi que le métabolisme et le phénotype du tissu adipeux, en font une stratégie préventive/thérapeutique de choix. Il semblerait intéressant d'étudier le tissu adipeux périvasculaire humain, prélevé chez des personnes obèses, après un protocole de réhabilitation par l'exercice physique. Ce nouvel acteur offre de nouvelles perspectives dans la compréhension de la fonction vasculaire, et pourrait constituer un lien intéressant entre l'obésité ou le syndrome métabolique, et le développement de complications vasculaires.

CONCLUSION GENERALE ET PERSPECTIVES

Ces travaux de doctorat ont permis de mettre en valeurs le rôle clé de la fonction endothéliale et du couple eNOS/Stress oxydant, dans l'adaptation de la réponse vasculaire à différents troubles de l'homéostasie, qu'ils soient métaboliques aigu (boisson sucrée) ou chronique (régime de type « Western »). En effet, dans nos travaux, nous avons pu mettre en avant qu'un simple stress hyperglycémique aigu peut avoir des conséquences fonctionnelles sur la vasomotricité endothélium-dépendante. Néanmoins, en nous appuyant sur la littérature abondante, rapportant des altérations vasculaires endothéliales persistantes chez les sujets souffrant de désordres métaboliques tels que le diabète de type 2 ou le syndrome métabolique, nous interprétons cette dysfonction endothéliale transitoire associée à l'hyperglycémie, comme un élément précurseur de l'installation d'une pathologie vasculaire chronique. Ceci, néanmoins, reste à démontrer. En effet, cette diminution de la réponse de l'endothélium au cours d'un stress aigu est observée sur quelques territoires seulement et pourrait finalement être un élément adaptatif permettant une meilleure distribution sanguine à d'autres territoires corporels, tel que la circulation splanchnique. Il semble donc hâtif de conclure de manière définitive, sur le lien entre cette diminution de la fonction endothéliale au cours d'un stress hyperglycémique, et l'installation d'une pathologie vasculaire persistante. De nouveaux travaux, visant à multiplier ce type de stress et d'en observer les conséquences progressives sur la fonction vasculaire, semblent essentiels afin de pouvoir tirer des conclusions plus fines.

Il ne semble donc pas évident que la fonction vasculaire soit dangereusement impactée par ce type de stress. En effet, comme nous avons pu le montrer dans les études n°2 et n°3, celle-ci possède une forte capacité d'adaptation, lui permettant, même face à un stress récurrent (régime High Fat High Sucrose) de mettre en place un certain nombre de mécanismes régulateurs, aboutissant à une normalisation de ses capacités de réponses physiologiques. Ainsi, dans l'étude n°2, une amélioration de l'activation de la eNOS à un stress adrénergique semble pouvoir réguler la pression artérielle en dépit de l'hyperactivation sympathique. Dans l'étude n°3, une bio-communication qui reste à élucider entre le PVAT et la musculature lisse du vaisseau, semble contribuer à une amélioration de la sensibilité au NO et ainsi à un maintien des capacités vasomotrices artérielles, en dépit de l'apparition d'une légère dysfonction endothéliale. Une des limites de ce travail de doctorat, est clairement que notre modèle de rat nourrit avec un régime HFS n'a développé qu'un état peu avancé de la pathologie métabolique. Néanmoins, ce qui peut être considéré comme une limite, nous a aussi permis d'étudier des phénomènes physiologiques ne pouvant être observés que dans la

phase précoce de ce type de maladie, et ainsi faire apparaître un certain nombre de mécanismes de compensation, que nous n'aurions certainement pas pu observer à un stade plus avancé. Néanmoins, si ceci semble évident au regard de la littérature scientifique, ces hypothèses doivent être confirmées expérimentalement.

Un élément essentiel de ce travail a aussi été de confirmer le rôle relativement protecteur de l'exercice physique. Comme déjà largement décrit dans la littérature, celui-ci semble pouvoir agir sur le bon fonctionnement du système cardiovasculaire, notamment *via* son action sur l'endothélium artériel et la biodisponibilité du NO. Dans ce travail, nous avons apporté un nouvel élément, en montrant que l'exercice est non seulement capable d'impacter la voie de la eNOS au niveau de l'endothélium vasculaire, mais aussi au niveau du PVAT. Par ailleurs, il semble, à partir de notre travail, que le PVAT puisse aussi constituer une cible privilégiée des effets bénéfiques de l'exercice sur l'homéostasie vasculaire. Cependant, comment le PVAT communique avec l'endothélium artériel et avec le reste du vaisseau, demeure néanmoins une zone d'ombre.

Ainsi, pour conclure, bien que la découverte du rôle de l'endothélium artériel dans la régulation de la fonction vasculaire ne soit pas très récente (Zalthen et al., 1978), la complexité de son rôle et de son impact sur la santé cardiovasculaire, le maintiennent en première ligne dans la compréhension, la modulation et le traitement potentiel de nombreuses maladies cardiovasculaires. La découverte plus récente et moins documentée de son interaction potentielle avec le PVAT, ajoute encore un peu de complexité au schéma de départ. Cependant, ce PVAT contribue à mieux comprendre le fonctionnement de l'endothélium vasculaire, et apparaît comme une potentielle nouvelle cible thérapeutique.

BIBLIOGRAPHIE

A

- Adams V, Linke A, Kränkel N, et al (2005) Impact of Regular Physical Activity on the NAD(P)H Oxidase and Angiotensin Receptor System in Patients With Coronary Artery Disease. *Circulation* 111:555–562. doi: 10.1161/01.CIR.0000154560.88933.7E
- Agapitov AV, Correia ML de G, Sinkey CA, Haynes WG (2008) Dissociation Between Sympathetic Nerve Traffic and Sympathetically Mediated Vascular Tone in Normotensive Human Obesity. *Hypertension* 52:687–695. doi: 10.1161/HYPERTENSIONAHA.107.109603
- Ago T, Kuroda J, Kamouchi M, et al (2011) Pathophysiological Roles of NADPH Oxidase/Nox Family Proteins in the Vascular System. *Circulation Journal* 75:1791–1800. doi: 10.1253/circj.CJ-11-0388
- Ahmed SR, Johansson BL, Karlsson MG, et al (2004) Human saphenous vein and coronary bypass surgery: ultrastructural aspects of conventional and “no-touch” vein graft preparations. *Histol Histopathol* 19:421–433.
- Alberti KGMM, Eckel RH, Grundy SM, et al (2009) Harmonizing the Metabolic Syndrome. *Circulation* 120:1640–1645. doi: 10.1161/CIRCULATIONAHA.109.192644
- Alderton WK, Cooper CE, Knowles RG (2001) Nitric oxide synthases: structure, function and inhibition. *Biochem J* 357:593–615.
- Alvarez GE, Beske SD, Ballard TP, Davy KP (2002) Sympathetic neural activation in visceral obesity. *Circulation* 106:2533–2536.
- Antonopoulos AS, Margaritis M, Coutinho P, et al (2015) Adiponectin as a Link Between Type 2 Diabetes and Vascular NADPH Oxidase Activity in the Human Arterial Wall: The Regulatory Role of Perivascular Adipose Tissue. *Diabetes* 64:2207–2219. doi: 10.2337/db14-1011
- Apovian CM, Bigornia S, Mott M, et al (2008) Adipose macrophage infiltration is associated with insulin resistance and vascular endothelial dysfunction in obese subjects. *Arterioscler Thromb Vasc Biol* 28:1654–1659. doi: 10.1161/ATVBAHA.108.170316
- Araujo HN, Valgas da Silva CP, Sponton ACS, et al (2015) Perivascular adipose tissue and vascular responses in healthy trained rats. *Life Sci.* doi: 10.1016/j.lfs.2014.12.032
- Arita Y, Kihara S, Ouchi N, et al (2012) Paradoxical decrease of an adipose-specific protein, adiponectin, in obesity. *Biochem Biophys Res Commun* 425:560–564. doi: 10.1016/j.bbrc.2012.08.024
- Arvola P, Wu X, Kähönen M, et al (1999) Exercise enhances vasorelaxation in experimental obesity associated hypertension. *Cardiovasc Res* 43:992–1002.

B

- Ballinger SW, Patterson C, Knight-Lozano CA, et al (2002) Mitochondrial integrity and

- function in atherogenesis. *Circulation* 106:544–549.
- Barrière E, Tazi KA, Pessione F, et al (2001) Role of small-conductance Ca²⁺-dependent K⁺ channels in in vitro nitric oxide-mediated aortic hyporeactivity to alpha-adrenergic vasoconstriction in rats with cirrhosis. *J Hepatol* 35:350–357.
- Basta G, Schmidt AM, De Caterina R (2004) Advanced glycation end products and vascular inflammation: implications for accelerated atherosclerosis in diabetes. *Cardiovasc Res* 63:582–592. doi: 10.1016/j.cardiores.2004.05.001
- Battault S, Singh F, Gayrard S, et al (2016) Endothelial function does not improve with high-intensity continuous exercise training in SHR: implications of eNOS uncoupling. *Hypertens Res* 39:70–78. doi: 10.1038/hr.2015.114
- Bedford MT, Clarke SG (2009) Protein arginine methylation in mammals: who, what, and why. *Mol Cell* 33:1–13. doi: 10.1016/j.molcel.2008.12.013
- Bełtowski J (2006) Role of leptin in blood pressure regulation and arterial hypertension. *J Hypertens* 24:789–801. doi: 10.1097/01.hjh.0000222743.06584.66
- Benjamin N, Dutton JA, Ritter JM (1991) Human vascular smooth muscle cells inhibit platelet aggregation when incubated with glyceryl trinitrate: evidence for generation of nitric oxide. *Br J Pharmacol* 102:847–850.
- Berg A, Frey I, Baumstark MW, et al (1994) Physical activity and lipoprotein lipid disorders. *Sports Med* 17:6–21.
- Berg AH, Scherer PE (2005) Adipose Tissue, Inflammation, and Cardiovascular Disease. *Circulation Research* 96:939–949. doi: 10.1161/01.RES.0000163635.62927.34
- Berggren JR, Hulver MW, Houmard JA (2005) Fat as an endocrine organ: influence of exercise. *J Appl Physiol* 99:757–764. doi: 10.1152/japplphysiol.00134.2005
- Berry CE, Hare JM (2004) Xanthine oxidoreductase and cardiovascular disease: molecular mechanisms and pathophysiological implications. *J Physiol (Lond)* 555:589–606. doi: 10.1113/jphysiol.2003.055913
- Berul CI, Maguire CT, Gehrmann J, Reddy S (2000) Progressive atrioventricular conduction block in a mouse myotonic dystrophy model. *J Interv Card Electrophysiol* 4:351–358.
- Beske SD, Alvarez GE, Ballard TP, Davy KP (2002) Reduced cardiovagal baroreflex gain in visceral obesity: implications for the metabolic syndrome. *Am J Physiol Heart Circ Physiol* 282:H630–635. doi: 10.1152/ajpheart.00642.2001
- Bivalacqua TJ, Armstrong JS, Biggerstaff J, et al (2003) Gene transfer of extracellular SOD to the penis reduces O[·] and improves erectile function in aged rats. *American Journal of Physiology - Heart and Circulatory Physiology* 284:H1408–H1421. doi: 10.1152/ajpheart.00770.2002
- Blankenberg S, Rupprecht HJ, Bickel C, et al (2003) Glutathione peroxidase 1 activity and cardiovascular events in patients with coronary artery disease. *N Engl J Med* 349:1605–1613. doi: 10.1056/NEJMoa030535
- Bolotina VM, Najibi S, Palacino JJ, et al (1994) Nitric oxide directly activates calcium-

- dependent potassium channels in vascular smooth muscle. *Nature* 368:850–853. doi: 10.1038/368850a0
- Bonetti PO, Lerman LO, Lerman A (2003) Endothelial dysfunction: a marker of atherosclerotic risk. *Arterioscler Thromb Vasc Biol* 23:168–175.
- Boo YC, Jo H (2003) Flow-dependent regulation of endothelial nitric oxide synthase: role of protein kinases. *American Journal of Physiology - Cell Physiology* 285:C499–C508. doi: 10.1152/ajpcell.00122.2003
- Boo YC, Sorescu G, Boyd N, et al (2002) Shear stress stimulates phosphorylation of endothelial nitric-oxide synthase at Ser1179 by Akt-independent mechanisms: role of protein kinase A. *J Biol Chem* 277:3388–3396. doi: 10.1074/jbc.M108789200
- Bossy-Wetzel E, Lipton SA (2003) Nitric oxide signaling regulates mitochondrial number and function. *Cell Death Differ* 10:757–760. doi: 10.1038/sj.cdd.4401244
- Boström P, Wu J, Jedrychowski MP, et al (2012) A PGC1- α -dependent myokine that drives brown-fat-like development of white fat and thermogenesis. *Nature* 481:463–468. doi: 10.1038/nature10777
- Boulbou MS, Koukoulis GN, Makri ED, et al (2005) Circulating adhesion molecules levels in type 2 diabetes mellitus and hypertension. *Int J Cardiol* 98:39–44. doi: 10.1016/j.ijcard.2003.07.037
- Boulé NG, Haddad E, Kenny GP, et al (2001) Effects of exercise on glycemic control and body mass in type 2 diabetes mellitus: a meta-analysis of controlled clinical trials. *JAMA* 286:1218–1227.
- Boulé NG, Kenny GP, Haddad E, et al (2003) Meta-analysis of the effect of structured exercise training on cardiorespiratory fitness in Type 2 diabetes mellitus. *Diabetologia* 46:1071–1081. doi: 10.1007/s00125-003-1160-2
- Boveris A, Cadenas E (1975) Mitochondrial production of superoxide anions and its relationship to the antimycin insensitive respiration. *FEBS Lett* 54:311–314.
- Bradley RL, Jeon JY, Liu F-F, Maratos-Flier E (2008) Voluntary exercise improves insulin sensitivity and adipose tissue inflammation in diet-induced obese mice. *Am J Physiol Endocrinol Metab* 295:E586–594. doi: 10.1152/ajpendo.00309.2007
- Brandes RP, Kreuzer J (2005) Vascular NADPH oxidases: molecular mechanisms of activation. *Cardiovasc Res* 65:16–27. doi: 10.1016/j.cardiores.2004.08.007
- Brandes RP, Schröder K (2008) Differential vascular functions of Nox family NADPH oxidases. *Curr Opin Lipidol* 19:513–518. doi: 10.1097/MOL.0b013e32830c91e3
- Britton KA, Pedley A, Massaro JM, et al (2012) Prevalence, distribution, and risk factor correlates of high thoracic periaortic fat in the Framingham Heart Study. *J Am Heart Assoc* 1:e004200. doi: 10.1161/JAHA.112.004200
- Brown CM, Dulloo AG, Montani J-P (2008) Sugary drinks in the pathogenesis of obesity and cardiovascular diseases. *Int J Obes (Lond)* 32 Suppl 6:S28-34. doi: 10.1038/ijo.2008.204

- Brown NK, Zhou Z, Zhang J, et al (2014) Perivascular Adipose Tissue in Vascular Function and Disease. *Arteriosclerosis, Thrombosis, and Vascular Biology* 34:1621–1630. doi: 10.1161/ATVBAHA.114.303029
- Brownlee M (2001) Biochemistry and molecular cell biology of diabetic complications. *Nature* 414:813–820. doi: 10.1038/414813a
- Bruun JM, Helge JW, Richelsen B, Stallknecht B (2006) Diet and exercise reduce low-grade inflammation and macrophage infiltration in adipose tissue but not in skeletal muscle in severely obese subjects. *Am J Physiol Endocrinol Metab* 290:E961-967. doi: 10.1152/ajpendo.00506.2005
- Bunker AK, Laughlin MH (2010) Influence of exercise and perivascular adipose tissue on coronary artery vasomotor function in a familial hypercholesterolemic porcine atherosclerosis model. *J Appl Physiol* 108:490–497. doi: 10.1152/japplphysiol.00999.2009
- Busetto L, Digito M, Dalla Montá P, et al (1993) Omental and epigastric adipose tissue lipolytic activity in human obesity. Effect of abdominal fat distribution and relationship with hyperinsulinemia. *Horm Metab Res* 25:365–371. doi: 10.1055/s-2007-1002121



- Cadenas E, Davies KJ (2000) Mitochondrial free radical generation, oxidative stress, and aging. *Free Radic Biol Med* 29:222–230.
- Cai H, Harrison DG (2000) Endothelial dysfunction in cardiovascular diseases: the role of oxidant stress. *Circ Res* 87:840–844.
- Cao L, Choi EY, Liu X, et al (2011) White to brown fat phenotypic switch induced by genetic and environmental activation of a hypothalamic-adipocyte axis. *Cell Metab* 14:324–338. doi: 10.1016/j.cmet.2011.06.020
- Cardoso CG, Gomides RS, Queiroz ACC, et al (2010) Acute and chronic effects of aerobic and resistance exercise on ambulatory blood pressure. *Clinics (Sao Paulo)* 65:317–325. doi: 10.1590/S1807-59322010000300013
- Cawthon WP, Sethi JK (2008) TNF-alpha and adipocyte biology. *FEBS Lett* 582:117–131. doi: 10.1016/j.febslet.2007.11.051
- Ceriello A, Taboga C, Tonutti L, et al (2002) Evidence for an independent and cumulative effect of postprandial hypertriglyceridemia and hyperglycemia on endothelial dysfunction and oxidative stress generation: effects of short- and long-term simvastatin treatment. *Circulation* 106:1211–1218.
- Chang L, Villacorta L, Li R, et al (2012) Loss of Perivascular Adipose Tissue on Peroxisome Proliferator-Activated Receptor- γ Deletion in Smooth Muscle Cells Impairs

- Intravascular Thermoregulation and Enhances AtherosclerosisClinical Perspective. Circulation 126:1067–1078. doi: 10.1161/CIRCULATIONAHA.112.104489
- Channon K (2004) Tetrahydrobiopterin: Regulator of Endothelial Nitric Oxide Synthase in Vascular Disease. Trends in Cardiovascular Medicine 14:323–327. doi: 10.1016/j.tcm.2004.10.003
- Charkoudian N, Joyner MJ, Barnes SA, et al (2006) Relationship between muscle sympathetic nerve activity and systemic hemodynamics during nitric oxide synthase inhibition in humans. Am J Physiol Heart Circ Physiol 291:H1378-1383. doi: 10.1152/ajpheart.00234.2006
- Chatterjee TK, Stoll LL, Denning GM, et al (2009) Proinflammatory phenotype of perivascular adipocytes: influence of high-fat feeding. Circ Res 104:541–549. doi: 10.1161/CIRCRESAHA.108.182998
- Chen C-A, Wang T-Y, Varadharaj S, et al (2010) S-glutathionylation uncouples eNOS and regulates its cellular and vascular function. Nature 468:1115–1118. doi: 10.1038/nature09599
- Chen D-D, Chen L-Y, Xie J-B, et al (2014) Tetrahydrobiopterin regulation of eNOS redox function. Curr Pharm Des 20:3554–3562.
- Chen H, Montagnani M, Funahashi T, et al (2003) Adiponectin Stimulates Production of Nitric Oxide in Vascular Endothelial Cells. J Biol Chem 278:45021–45026. doi: 10.1074/jbc.M307878200
- Chen Hi H–, Chiang I-P, Jen CJ (1996) Exercise Training Increases Acetylcholine-Stimulated Endothelium-Derived Nitric Oxide Release in Spontaneously Hypertensive Rats. J Biomed Sci 3:454–460.
- Cheng F, Torzewski M, Degreif A, et al (2013) Impact of Glutathione Peroxidase-1 Deficiency on Macrophage Foam Cell Formation and Proliferation: Implications for Atherogenesis. PLOS ONE 8:e72063. doi: 10.1371/journal.pone.0072063
- Chin-Dusting JPF, Willems L, Kaye DM (2007) l-Arginine transporters in cardiovascular disease: A novel therapeutic target. Pharmacology & Therapeutics 116:428–436. doi: 10.1016/j.pharmthera.2007.08.001
- Chobanian AV, Bakris GL, Black HR, et al (2003) The Seventh Report of the Joint National Committee on Prevention, Detection, Evaluation, and Treatment of High Blood Pressure: the JNC 7 report. JAMA 289:2560–2572. doi: 10.1001/jama.289.19.2560
- Chouchani ET, Methner C, Nadtochiy SM, et al (2013) Cardioprotection by S-nitrosation of a cysteine switch on mitochondrial complex I. Nat Med 19:753–759. doi: 10.1038/nm.3212
- Chow W-S, Cheung BMY, Tso AWK, et al (2007) Hypoadiponectinemia as a predictor for the development of hypertension: a 5-year prospective study. Hypertension 49:1455–1461. doi: 10.1161/HYPERTENSIONAHA.107.086835
- Christiansen T, Paulsen SK, Bruun JM, et al (2010) Exercise training versus diet-induced

- weight-loss on metabolic risk factors and inflammatory markers in obese subjects: a 12-week randomized intervention study. *American Journal of Physiology - Endocrinology and Metabolism* 298:E824–E831. doi: 10.1152/ajpendo.00574.2009
- Christ-Roberts CY, Pratipanawatr T, Pratipanawatr W, et al (2004) Exercise training increases glycogen synthase activity and GLUT4 expression but not insulin signaling in overweight nondiabetic and type 2 diabetic subjects. *Metab Clin Exp* 53:1233–1242.
- Chughtai HL, Morgan TM, Rocco M, et al (2010) Renal sinus fat and poor blood pressure control in middle-aged and elderly individuals at risk for cardiovascular events. *Hypertension* 56:901–906. doi: 10.1161/HYPERTENSIONAHA.110.157370
- Clementi E, Nisoli E (2005) Nitric oxide and mitochondrial biogenesis: a key to long-term regulation of cellular metabolism. *Comp Biochem Physiol, Part A Mol Integr Physiol* 142:102–110. doi: 10.1016/j.cbpb.2005.04.022
- Cornelissen VA, Fagard RH (2005) Effect of resistance training on resting blood pressure: a meta-analysis of randomized controlled trials. *J Hypertens* 23:251–259.
- Cornwell TL, Pryzwansky KB, Wyatt TA, Lincoln TM (1991) Regulation of sarcoplasmic reticulum protein phosphorylation by localized cyclic GMP-dependent protein kinase in vascular smooth muscle cells. *Mol Pharmacol* 40:923–931.
- Corson MA, James NL, Latta SE, et al (1996) Phosphorylation of endothelial nitric oxide synthase in response to fluid shear stress. *Circ Res* 79:984–991.
- Crabtree MJ, Channon KM (2011) Synthesis and recycling of tetrahydrobiopterin in endothelial function and vascular disease. *Nitric Oxide* 25:81–88. doi: 10.1016/j.niox.2011.04.004
- Crabtree MJ, Smith CL, Lam G, et al (2008) Ratio of 5,6,7,8-tetrahydrobiopterin to 7,8-dihydrobiopterin in endothelial cells determines glucose-elicited changes in NO vs. superoxide production by eNOS. *Am J Physiol Heart Circ Physiol* 294:H1530–1540. doi: 10.1152/ajpheart.00823.2007



da Silva CA, Ribeiro JP, Canto JCAU, et al (2012) High-intensity aerobic training improves endothelium-dependent vasodilation in patients with metabolic syndrome and type 2 diabetes mellitus. *Diabetes Res Clin Pract* 95:237–245. doi: 10.1016/j.diabres.2011.09.034

Davis ME, Cai H, Drummond GR, Harrison DG (2001) Shear stress regulates endothelial nitric oxide synthase expression through c-Src by divergent signaling pathways. *Circ Res* 89:1073–1080.

Davis ME, Cai H, McCann L, et al (2003) Role of c-Src in regulation of endothelial nitric oxide synthase expression during exercise training. *American Journal of Physiology -*

- Heart and Circulatory Physiology 284:H1449–H1453. doi: 10.1152/ajpheart.00918.2002
- Dawson J, Walters M (2006) Uric acid and xanthine oxidase: future therapeutic targets in the prevention of cardiovascular disease? Br J Clin Pharmacol 62:633–644.
- De Glisezinski I, Crampes F, Harant I, et al (1998) Endurance training changes in lipolytic responsiveness of obese adipose tissue. Am J Physiol 275:E951-956.
- Deedwania PC, Gupta R (2006) Management issues in the metabolic syndrome. J Assoc Physicians India 54:797–810.
- Demircan S, Yazici M, Diraman E, et al (2008) The effect of glucose-insulin-potassium treatment on myocardial oxidative stress in patients with acute coronary syndromes undergoing percutaneous coronary intervention. Coron Artery Dis 19:99–104. doi: 10.1097/MCA.0b013e3282f27c34
- Dimmeler S, Fleming I, Fisslthaler B, et al (1999) Activation of nitric oxide synthase in endothelial cells by Akt-dependent phosphorylation. Nature 399:601–605. doi: 10.1038/21224
- Dora KA, Doyle MP, Duling BR (1997) Elevation of intracellular calcium in smooth muscle causes endothelial cell generation of NO in arterioles. Proc Natl Acad Sci USA 94:6529–6534.
- Dreifaldt M, Souza D, Bodin L, et al (2013) The Vasa Vasorum and Associated Endothelial Nitric Oxide Synthase is More Important for Saphenous Vein Than Arterial Bypass Grafts. ANGIOLOGY 64:293–299. doi: 10.1177/0003319712443729
- Drexler H, Lu W (1992) Endothelial dysfunction of hindquarter resistance vessels in experimental heart failure. Am J Physiol 262:H1640-1645.
- Drummond GR, Cai H, Davis ME, et al (2000) Transcriptional and posttranscriptional regulation of endothelial nitric oxide synthase expression by hydrogen peroxide. Circ Res 86:347–354.
- Dubrovska G, Verlohren S, Luft FC, Gollasch M (2004) Mechanisms of ADRF release from rat aortic adventitial adipose tissue. Am J Physiol Heart Circ Physiol 286:H1107-1113. doi: 10.1152/ajpheart.00656.2003
- Duerrscheidt N, Stielow C, Muller G, et al (2006) NO-mediated regulation of NAD(P)H oxidase by laminar shear stress in human endothelial cells. J Physiol (Lond) 576:557–567. doi: 10.1113/jphysiol.2006.111070
- Elizalde M, Rydén M, van Harmelen V, et al (2000) Expression of nitric oxide synthases in subcutaneous adipose tissue of nonobese and obese humans. J Lipid Res 41:1244–1251.

Enerbäck S (2009) The origins of brown adipose tissue. *N Engl J Med* 360:2021–2023. doi: 10.1056/NEJMcibr0809610

Ervin RB (2009) Prevalence of metabolic syndrome among adults 20 years of age and over, by sex, age, race and ethnicity, and body mass index: United States, 2003–2006. *Natl Health Stat Report* 1–7.

Esler M, Lambert E, Schlaich M (2010) Point: Chronic activation of the sympathetic nervous system is the dominant contributor to systemic hypertension. *J Appl Physiol* 109:1996–1998; discussion 2016. doi: 10.1152/japplphysiol.00182.2010

F

Fagherazzi G, Vilier A, Saes Sartorelli D, et al (2013) Consumption of artificially and sugar-sweetened beverages and incident type 2 diabetes in the Etude Epidemiologique aupres des femmes de la Mutuelle Generale de l'Education Nationale-European Prospective Investigation into Cancer and Nutrition cohort. *Am J Clin Nutr* 97:517–523. doi: 10.3945/ajcn.112.050997

Farah C, Kleindienst A, Bolea G, et al (2013) Exercise-induced cardioprotection: a role for eNOS uncoupling and NO metabolites. *Basic Res Cardiol* 108:389. doi: 10.1007/s00395-013-0389-2

Fehm HL, Kern W, Peters A (2006) The selfish brain: competition for energy resources. *Prog Brain Res* 153:129–140. doi: 10.1016/S0079-6123(06)53007-9

Feldman BJ, Streeper RS, Farese RV, Yamamoto KR (2006) Myostatin modulates adipogenesis to generate adipocytes with favorable metabolic effects. *PNAS* 103:15675–15680. doi: 10.1073/pnas.0607501103

Féletalou M, Vanhoutte PM (2006) Endothelium-derived hyperpolarizing factor: where are we now? *Arterioscler Thromb Vasc Biol* 26:1215–1225. doi: 10.1161/01.ATV.0000217611.81085.c5

Felix JVC, Michelini LC (2007) Training-induced pressure fall in spontaneously hypertensive rats is associated with reduced angiotensinogen mRNA expression within the nucleus tractus solitarii. *Hypertension* 50:780–785. doi: 10.1161/HYPERTENSIONAHA.107.094474

Ferrannini E (2005) Insulin and blood pressure: connected on a circumference? *Hypertension* 45:347–348. doi: 10.1161/01.HYP.0000155464.44905.6c

Ferrannini E, Natali A, Capaldo B, et al (1997) Insulin resistance, hyperinsulinemia, and blood pressure: role of age and obesity. European Group for the Study of Insulin Resistance (EGIR). *Hypertension* 30:1144–1149.

Fésüs G, Dubrovska G, Gorzelnik K, et al (2007) Adiponectin is a novel humoral vasodilator. *Cardiovascular Research* 75:719–727. doi:

10.1016/j.cardiores.2007.05.025

- Feyter HMD, Praet SF, Broek NM van den, et al (2007) Exercise Training Improves Glycemic Control in Long-Standing Insulin-Treated Type 2 Diabetic Patients. *Diabetes Care* 30:2511–2513. doi: 10.2337/dc07-0183
- Fitzgibbons TP, Czech MP (2014) Epicardial and perivascular adipose tissues and their influence on cardiovascular disease: basic mechanisms and clinical associations. *J Am Heart Assoc* 3:e000582. doi: 10.1161/JAHA.113.000582
- Fitzgibbons TP, Kogan S, Aouadi M, et al (2011) Similarity of mouse perivascular and brown adipose tissues and their resistance to diet-induced inflammation. *Am J Physiol Heart Circ Physiol* 301:H1425–1437. doi: 10.1152/ajpheart.00376.2011
- Flammer AJ, Anderson T, Celermajer DS, et al (2012) The Assessment of Endothelial Function. *Circulation* 126:753–767. doi: 10.1161/CIRCULATIONAHA.112.093245
- Fleming I (2009) Molecular mechanisms underlying the activation of eNOS. *Pflugers Arch - Eur J Physiol* 459:793–806. doi: 10.1007/s00424-009-0767-7
- Floras JS, Ponikowski P (2015) The sympathetic/parasympathetic imbalance in heart failure with reduced ejection fraction. *Eur Heart J* 36:1974–1982b. doi: 10.1093/eurheartj/ehv087
- Fornoni A, Raij L (2005) Metabolic syndrome and endothelial dysfunction. *Curr Hypertens Rep* 7:88–95.
- Förstermann U, Münz T (2006) Endothelial nitric oxide synthase in vascular disease: from marvel to menace. *Circulation* 113:1708–1714. doi: 10.1161/CIRCULATIONAHA.105.602532
- Förstermann U, Sessa WC (2012) Nitric oxide synthases: regulation and function. *Eur Heart J* 33:829–837, 837a–837d. doi: 10.1093/eurheartj/ehr304
- Frisbee JC (2006) Impaired hemorrhage tolerance in the obese Zucker rat model of metabolic syndrome. *J Appl Physiol* 100:465–473. doi: 10.1152/japplphysiol.01062.2005
- Fu WJ, Haynes TE, Kohli R, et al (2005) Dietary L-arginine supplementation reduces fat mass in Zucker diabetic fatty rats. *J Nutr* 135:714–721.
- Fukai T, Siegfried MR, Ushio-Fukai M, et al (2000) Regulation of the vascular extracellular superoxide dismutase by nitric oxide and exercise training. *J Clin Invest* 105:1631–1639. doi: 10.1172/JCI9551
- Fukai T, Ushio-Fukai M (2011) Superoxide Dismutases: Role in Redox Signaling, Vascular Function, and Diseases. *Antioxidants & Redox Signaling* 15:1583–1606. doi: 10.1089/ars.2011.3999
- Fulton D, Gratton J-P, McCabe TJ, et al (1999) Regulation of endothelium-derived nitric oxide production by the protein kinase Akt. *Nature* 399:597–601. doi: 10.1038/21218
- Furchtgott RF, Zawadzki JV (1980) The obligatory role of endothelial cells in the relaxation of arterial smooth muscle by acetylcholine. *Nature* 288:373–376.



- Gadegbeku CA, Dhandayuthapani A, Sadler ZE, Egan BM (2002) Raising lipids acutely reduces baroreflex sensitivity. *Am J Hypertens* 15:479–485.
- Gaede PH, Jepsen PV, Larsen JNB, et al (2003) [The Steno-2 study. Intensive multifactorial intervention reduces the occurrence of cardiovascular disease in patients with type 2 diabetes]. *Ugeskr Laeg* 165:2658–2661.
- Gao Y-J, Lu C, Su L-Y, et al (2007) Modulation of vascular function by perivascular adipose tissue: the role of endothelium and hydrogen peroxide. *British Journal of Pharmacology* 151:323–331. doi: 10.1038/sj.bjp.0707228
- Gao Y-J, Takemori K, Su L-Y, et al (2006) Perivascular adipose tissue promotes vasoconstriction: the role of superoxide anion. *Cardiovascular Research* 71:363–373. doi: 10.1016/j.cardiores.2006.03.013
- Gaudiot N, Ribière C, Jaubert AM, Giudicelli Y (2000) Endogenous nitric oxide is implicated in the regulation of lipolysis through antioxidant-related effect. *Am J Physiol, Cell Physiol* 279:C1603-1610.
- Gehrman J, Hammer PE, Maguire CT, et al (2000) Phenotypic screening for heart rate variability in the mouse. *Am J Physiol Heart Circ Physiol* 279:H733-740.
- Ghosh AK, Vaughan DE (2012) PAI-1 in tissue fibrosis. *J Cell Physiol* 227:493–507. doi: 10.1002/jcp.22783
- Gil-Ortega M, Somoza B, Huang Y, et al (2015) Regional differences in perivascular adipose tissue impacting vascular homeostasis. *Trends in Endocrinology & Metabolism* 26:367–375. doi: 10.1016/j.tem.2015.04.003
- Gil-Ortega M, Stucchi P, Guzmán-Ruiz R, et al (2010) Adaptative nitric oxide overproduction in perivascular adipose tissue during early diet-induced obesity. *Endocrinology* 151:3299–3306. doi: 10.1210/en.2009-1464
- Glisezinski I de, Moro C, Pillard F, et al (2003) Aerobic training improves exercise-induced lipolysis in SCAT and lipid utilization in overweight men. *American Journal of Physiology - Endocrinology and Metabolism* 285:E984–E990. doi: 10.1152/ajpendo.00152.2003
- Golbidi S, Mesdaghinia A, Laher I (2012) Exercise in the Metabolic Syndrome. *Oxidative Medicine and Cellular Longevity* 2012:e349710. doi: 10.1155/2012/349710
- Goldhammer E, Ben-Sira D, Zaid G, et al (2007) Paraoxonase activity following exercise-based cardiac rehabilitation program. *J Cardiopulm Rehabil Prev* 27:151–154. doi: 10.1097/HCR.0000270691.09258.b1
- Goldstein DS, McCarty R, Polinsky RJ, Kopin IJ (1983) Relationship between plasma

- norepinephrine and sympathetic neural activity. *Hypertension* 5:552–559.
- Gollisch KSC, Brandauer J, Jessen N, et al (2009) Effects of exercise training on subcutaneous and visceral adipose tissue in normal- and high-fat diet-fed rats. *Am J Physiol Endocrinol Metab* 297:E495-504. doi: 10.1152/ajpendo.90424.2008
- Gow AJ, Stamler JS (1998) Reactions between nitric oxide and haemoglobin under physiological conditions. *Nature* 391:169–173. doi: 10.1038/34402
- Graham DA, Rush JWE (2004) Exercise training improves aortic endothelium-dependent vasorelaxation and determinants of nitric oxide bioavailability in spontaneously hypertensive rats. *J Appl Physiol* 96:2088–2096. doi: 10.1152/japplphysiol.01252.2003
- Grasser EK, Dulloo A, Montani J-P (2014a) Cardiovascular responses to the ingestion of sugary drinks using a randomised cross-over study design: Does glucose attenuate the blood pressure-elevating effect of fructose? *Br J Nutr* 112:183–192. doi: 10.1017/S0007114514000622
- Grasser EK, Yepuri G, Dulloo AG, Montani J-P (2014b) Cardio- and cerebrovascular responses to the energy drink Red Bull in young adults: a randomized cross-over study. *Eur J Nutr* 53:1561–1571. doi: 10.1007/s00394-014-0661-8
- Grassi G (2006) Sympathetic overdrive and cardiovascular risk in the metabolic syndrome. *Hypertens Res* 29:839–847. doi: 10.1291/hypres.29.839
- Grassi G, Seravalle G (2006) Autonomic imbalance and metabolic syndrome: unravelling interactions, mechanisms and outcomes. *J Hypertens* 24:47–49.
- Grassi G, Seravalle G, Cattaneo BM, et al (1995) Sympathetic Activation and Loss of Reflex Sympathetic Control in Mild Congestive Heart Failure. *Circulation* 92:3206–3211. doi: 10.1161/01.CIR.92.11.3206
- Grassi G, Seravalle G, Trevano FQ, et al (2007) Neurogenic abnormalities in masked hypertension. *Hypertension* 50:537–542. doi: 10.1161/HYPERTENSIONAHA.107.092528
- Green DJ, O'Driscoll G, Joyner MJ, Cable NT (2008) Exercise and cardiovascular risk reduction: Time to update the rationale for exercise? *Journal of Applied Physiology* 105:766–768. doi: 10.1152/japplphysiol.01028.2007
- Green DJ, Spence A, Halliwill JR, et al (2011) Exercise and vascular adaptation in asymptomatic humans. *Exp Physiol* 96:57–70. doi: 10.1113/expphysiol.2009.048694
- Greenstein AS, Khavandi K, Withers SB, et al (2009) Local Inflammation and Hypoxia Abolish the Protective Anticontractile Properties of Perivascular Fat in Obese Patients. *Circulation* 119:1661–1670. doi: 10.1161/CIRCULATIONAHA.108.821181
- Grundy SM, Brewer HB, Cleeman JI, et al (2004) Definition of metabolic syndrome: Report of the National Heart, Lung, and Blood Institute/American Heart Association conference on scientific issues related to definition. *Circulation* 109:433–438. doi: 10.1161/01.CIR.0000111245.75752.C6

- Gu P, Xu A (2013) Interplay between adipose tissue and blood vessels in obesity and vascular dysfunction. *Rev Endocr Metab Disord* 14:49–58. doi: 10.1007/s11154-012-9230-8
- Gündüz F, Koçer G, Ulker S, et al (2011) Exercise training enhances flow-mediated dilation in spontaneously hypertensive rats. *Physiol Res* 60:589–597.
- Guzik TJ, Sadowski J, Guzik B, et al (2006) Coronary Artery Superoxide Production and Nox Isoform Expression in Human Coronary Artery Disease. *Arteriosclerosis, Thrombosis, and Vascular Biology* 26:333–339. doi: 10.1161/01.ATV.0000196651.64776.51

H

- Haack KKV, Zucker IH (2015) Central mechanisms for exercise training-induced reduction in sympatho-excitation in chronic heart failure. *Auton Neurosci* 188:44–50. doi: 10.1016/j.autneu.2014.10.015
- Haastrup AT, Stepniakowski KT, Goodfriend TL, Egan BM (1998) Intralipid enhances alpha₁-adrenergic receptor mediated pressor sensitivity. *Hypertension* 32:693–698.
- Hadi HAR, Carr CS, Al Suwaidi J (2005) Endothelial dysfunction: cardiovascular risk factors, therapy, and outcome. *Vasc Health Risk Manag* 1:183–198.
- Hajer GR, Haeften TW van, Visseren FLJ (2008) Adipose tissue dysfunction in obesity, diabetes, and vascular diseases. *European Heart Journal* 29:2959–2971. doi: 10.1093/eurheartj/ehn387
- Halaas JL, Gajiwala KS, Maffei M, et al (1995) Weight-reducing effects of the plasma protein encoded by the obese gene. *Science* 269:543–546.
- Hambrecht R, Adams V, Erbs S, et al (2003) Regular physical activity improves endothelial function in patients with coronary artery disease by increasing phosphorylation of endothelial nitric oxide synthase. *Circulation* 107:3152–3158. doi: 10.1161/01.CIR.0000074229.93804.5C
- Hamer M, Stamatakis E (2012) Low-Dose Physical Activity Attenuates Cardiovascular Disease Mortality in Men and Women With Clustered Metabolic Risk Factors. *Circ Cardiovasc Qual Outcomes* 5:494–499. doi: 10.1161/CIRCOUTCOMES.112.965434
- Hamilton SJ, Watts GF (2013) Endothelial dysfunction in diabetes: pathogenesis, significance, and treatment. *Rev Diabet Stud* 10:133–156. doi: 10.1900/RDS.2013.10.133
- Harrison D, Griendling KK, Landmesser U, et al (2003) Role of oxidative stress in atherosclerosis. *Am J Cardiol* 91:7A–11A.
- Hattori Y, Suzuki M, Hattori S, Kasai K (2003) Globular adiponectin upregulates nitric oxide production in vascular endothelial cells. *Diabetologia* 46:1543–1549. doi: 10.1007/s00125-003-1224-3
- Haynes WG, Morgan DA, Walsh SA, et al (1997) Receptor-mediated regional sympathetic

- nerve activation by leptin. *J Clin Invest* 100:270–278.
- Haynes WG, Noon JP, Walker BR, Webb DJ (1993) Inhibition of nitric oxide synthesis increases blood pressure in healthy humans. *J Hypertens* 11:1375–1380.
- Heise T, Magnusson K, Heinemann L, Sawicki PT (1998) Insulin resistance and the effect of insulin on blood pressure in essential hypertension. *Hypertension* 32:243–248.
- Heitzer T, Schlinzig T, Krohn K, et al (2001) Endothelial dysfunction, oxidative stress, and risk of cardiovascular events in patients with coronary artery disease. *Circulation* 104:2673–2678.
- Henricson J, Tesselaar E, Persson K, et al (2007) Assessment of microvascular function by study of the dose-response effects of iontophoretically applied drugs (acetylcholine and sodium nitroprusside)--methods and comparison with in vitro studies. *Microvasc Res* 73:143–149. doi: 10.1016/j.mvr.2006.10.004
- Higashi Y, Noma K, Yoshizumi M, Kihara Y (2009) Endothelial function and oxidative stress in cardiovascular diseases. *Circ J* 73:411–418.
- Higashi Y, Sasaki S, Kurisu S, et al (1999a) Regular aerobic exercise augments endothelium-dependent vascular relaxation in normotensive as well as hypertensive subjects: role of endothelium-derived nitric oxide. *Circulation* 100:1194–1202.
- Higashi Y, Sasaki S, Sasaki N, et al (1999b) Daily aerobic exercise improves reactive hyperemia in patients with essential hypertension. *Hypertension* 33:591–597.
- Hink U, Tsilimingas N, Wendt M, Münz T (2003) Mechanisms underlying endothelial dysfunction in diabetes mellitus: therapeutic implications. *Treat Endocrinol* 2:293–304.
- Hirata M, Suzuki M, Ishii R, et al (2011) Genetic Defect in Phospholipase C δ 1 Protects Mice From Obesity by Regulating Thermogenesis and Adipogenesis. *Diabetes* 60:1926–1937. doi: 10.2337/db10-1500
- Hogarth AJ, Mackintosh AF, Mary DASG (2006) The Sympathetic Drive After Acute Myocardial Infarction in Hypertensive Patients*. *Am J Hypertens* 19:1070–1076. doi: 10.1016/j.amjhyper.2006.03.015
- Hotamisligil GS, Arner P, Caro JF, et al (1995) Increased adipose tissue expression of tumor necrosis factor-alpha in human obesity and insulin resistance. *J Clin Invest* 95:2409–2415. doi: 10.1172/JCI117936
- Hotamisligil GS, Budavari A, Murray D, Spiegelman BM (1994) Reduced tyrosine kinase activity of the insulin receptor in obesity-diabetes. Central role of tumor necrosis factor-alpha. *J Clin Invest* 94:1543–1549.
- Hu FB, Malik VS (2010) Sugar-sweetened beverages and risk of obesity and type 2 diabetes: epidemiologic evidence. *Physiol Behav* 100:47–54. doi: 10.1016/j.physbeh.2010.01.036
- Huang C, Huang J, Tian Y, et al (2014) Sugar sweetened beverages consumption and risk of coronary heart disease: a meta-analysis of prospective studies. *Atherosclerosis*

234:11–16. doi: 10.1016/j.atherosclerosis.2014.01.037

Hug C, Wang J, Ahmad NS, et al (2004) T-cadherin is a receptor for hexameric and high-molecular-weight forms of Acrp30/adiponectin. PNAS 101:10308–10313. doi: 10.1073/pnas.0403382101

Huggett RJ, Burns J, Mackintosh AF, Mary DASG (2004) Sympathetic neural activation in nondiabetic metabolic syndrome and its further augmentation by hypertension. Hypertension 44:847–852. doi: 10.1161/01.HYP.0000147893.08533.d8

Hulver MW, Zheng D, Tanner CJ, et al (2002) Adiponectin is not altered with exercise training despite enhanced insulin action. Am J Physiol Endocrinol Metab 283:E861–865. doi: 10.1152/ajpendo.00150.2002

J

Iacobellis G, Ribaudo MC, Leto G, et al (2002) Influence of excess fat on cardiac morphology and function: study in uncomplicated obesity. Obes Res 10:767–773. doi: 10.1038/oby.2002.104

Ignarro LJ, Buga GM, Wood KS, et al (1987) Endothelium-derived relaxing factor produced and released from artery and vein is nitric oxide. Proc Natl Acad Sci USA 84:9265–9269.

Ignarro LJ, Cirino G, Casini A, Napoli C (1999) Nitric oxide as a signaling molecule in the vascular system: an overview. J Cardiovasc Pharmacol 34:879–886.

IJzerman RG, de Jongh RT, Beijk M a. M, et al (2003) Individuals at increased coronary heart disease risk are characterized by an impaired microvascular function in skin. Eur J Clin Invest 33:536–542.

Ishibashi J, Seale P (2010) Medicine. Beige can be slimming. Science 328:1113–1114. doi: 10.1126/science.1190816

Ishii T, Yamakita T, Yamagami K, et al (2001) Effect of exercise training on serum leptin levels in type 2 diabetic patients. Metab Clin Exp 50:1136–1140. doi: 10.1053/meta.2001.26745

J

Jankovic A, Korac A, Buzadzic B, et al (2016) Targeting the NO/superoxide ratio in adipose tissue: relevance to obesity and diabetes management. British Journal of Pharmacology n/a-n/a. doi: 10.1111/bph.13498

Jerez S, Scacchi F, Sierra L, et al (2012) Vascular hyporeactivity to angiotensin II and noradrenaline in a rabbit model of obesity. J Cardiovasc Pharmacol 59:49–57. doi:

10.1097/FJC.0b013e318235156a

Jin Z-G, Wong C, Wu J, Berk BC (2005) Flow shear stress stimulates Gab1 tyrosine phosphorylation to mediate protein kinase B and endothelial nitric-oxide synthase activation in endothelial cells. *J Biol Chem* 280:12305–12309. doi: 10.1074/jbc.M500294200

Jobgen WS, Fried SK, Fu WJ, et al (2006) Regulatory role for the arginine–nitric oxide pathway in metabolism of energy substrates. *The Journal of Nutritional Biochemistry* 17:571–588. doi: 10.1016/j.jnutbio.2005.12.001

Jones CJ, DeFily DV, Patterson JL, Chilian WM (1993) Endothelium-dependent relaxation competes with alpha 1- and alpha 2-adrenergic constriction in the canine epicardial coronary microcirculation. *Circulation* 87:1264–1274.

Joyner MJ, Green DJ (2009) Exercise protects the cardiovascular system: effects beyond traditional risk factors. *J Physiol (Lond)* 587:5551–5558. doi: 10.1113/jphysiol.2009.179432

Joyner MJ, Nauss LA, Warner MA, Warner DO (1992) Sympathetic modulation of blood flow and O₂ uptake in rhythmically contracting human forearm muscles. *Am J Physiol* 263:H1078-1083.

K

Kadowaki T, Yamauchi T, Kubota N, et al (2006) Adiponectin and adiponectin receptors in insulin resistance, diabetes, and the metabolic syndrome. *J Clin Invest* 116:1784–1792. doi: 10.1172/JCI29126

Kaur J (2014) A comprehensive review on metabolic syndrome. *Cardiol Res Pract* 2014:943162. doi: 10.1155/2014/943162

Kelly DP, Scarpulla RC (2004) Transcriptional regulatory circuits controlling mitochondrial biogenesis and function. *Genes Dev* 18:357–368. doi: 10.1101/gad.1177604

Ketonen J, Shi J, Martonen E, Mervaala E (2010) Periadventitial adipose tissue promotes endothelial dysfunction via oxidative stress in diet-induced obese C57Bl/6 mice. *Circ J* 74:1479–1487.

Khan BV, Harrison DG, Olbrych MT, et al (1996) Nitric oxide regulates vascular cell adhesion molecule 1 gene expression and redox-sensitive transcriptional events in human vascular endothelial cells. *Proc Natl Acad Sci USA* 93:9114–9119.

Kishi T (2012) Heart failure as an autonomic nervous system dysfunction. *J Cardiol* 59:117–122. doi: 10.1016/j.jcc.2011.12.006

Kobayashi S, Maesato K, Moriya H, et al (2005) Insulin resistance in patients with chronic kidney disease. *American Journal of Kidney Diseases* 45:275–280. doi: 10.1053/j.ajkd.2004.09.034

- Kobzik L, Reid MB, Bredt DS, Stamler JS (1994) Nitric oxide in skeletal muscle. *Nature* 372:546–548. doi: 10.1038/372546a0
- Koppenol WH (1998) The basic chemistry of nitrogen monoxide and peroxynitrite. *Free Radic Biol Med* 25:385–391.
- Kosteli A, Sugaru E, Haemmerle G, et al (2010) Weight loss and lipolysis promote a dynamic immune response in murine adipose tissue. *J Clin Invest* 120:3466–3479. doi: 10.1172/JCI42845
- Kumagai K, Reid IA (1994) Angiotensin II exerts differential actions on renal nerve activity and heart rate. *Hypertension* 24:451–456.
- Kunsch C, Medford RM (1999) Oxidative stress as a regulator of gene expression in the vasculature. *Circ Res* 85:753–766.

Q

- Lafontan M, Berlan M (2003) Do regional differences in adipocyte biology provide new pathophysiological insights? *Trends in Pharmacological Sciences* 24:276–283. doi: 10.1016/S0165-6147(03)00132-9
- Lafontan M, Langin D (2009) Lipolysis and lipid mobilization in human adipose tissue. *Progress in Lipid Research* 48:275–297. doi: 10.1016/j.plipres.2009.05.001
- Lambert GW, Straznicky NE, Lambert EA, et al (2010) Sympathetic nervous activation in obesity and the metabolic syndrome--causes, consequences and therapeutic implications. *Pharmacol Ther* 126:159–172. doi: 10.1016/j.pharmthera.2010.02.002
- Lamboley M, Pittet P, Koenigsberger M, et al (2005) Evidence for signaling via gap junctions from smooth muscle to endothelial cells in rat mesenteric arteries: possible implication of a second messenger. *Cell Calcium* 37:311–320. doi: 10.1016/j.ceca.2004.11.004
- Lancet Diabetes Endocrinology (2015) Sugar intake: lowering the bar. *Lancet Diabetes Endocrinol* 3:305. doi: 10.1016/S2213-8587(15)00102-3
- Landmesser U, Spiekermann S, Preuss C, et al (2007) Angiotensin II induces endothelial xanthine oxidase activation: role for endothelial dysfunction in patients with coronary disease. *Arterioscler Thromb Vasc Biol* 27:943–948. doi: 10.1161/01.ATV.0000258415.32883.bf
- Lastra G, Manrique C, Sowers JR (2006) Obesity, Cardiometabolic Syndrome, and Chronic Kidney Disease: The Weight of the Evidence. *Advances in Chronic Kidney Disease* 13:365–373. doi: 10.1053/j.ackd.2006.07.011
- Lauer N, Suvorava T, Rüther U, et al (2005) Critical involvement of hydrogen peroxide in exercise-induced up-regulation of endothelial NO synthase. *Cardiovascular Research* 65:254–262. doi: 10.1016/j.cardiores.2004.09.010
- Lavrencic A, Salobir BG, Keber I (2000) Physical training improves flow-mediated dilation

- in patients with the polymetabolic syndrome. *Arterioscler Thromb Vasc Biol* 20:551–555.
- Lee KY, Yamamoto Y, Boucher J, et al (2013) Shox2 is a molecular determinant of depot-specific adipocyte function. *Proc Natl Acad Sci USA* 110:11409–11414. doi: 10.1073/pnas.1310331110
- Lee MH-H, Chen S-J, Tsao C-M, Wu C-C (2014) Perivascular Adipose Tissue Inhibits Endothelial Function of Rat Aortas via Caveolin-1. *PLOS ONE* 9:e99947. doi: 10.1371/journal.pone.0099947
- Lee Y-C, Chang H-H, Chiang C-L, et al (2011) Role of Perivascular Adipose Tissue–Derived Methyl Palmitate in Vascular Tone Regulation and Pathogenesis of HypertensionClinical Perspective. *Circulation* 124:1160–1171. doi: 10.1161/CIRCULATIONAHA.111.027375
- Lehman SJ, Massaro JM, Schlett CL, et al (2010) Peri-aortic fat, cardiovascular disease risk factors, and aortic calcification: the Framingham Heart Study. *Atherosclerosis* 210:656–661. doi: 10.1016/j.atherosclerosis.2010.01.007
- Li C, Wang Z, Wang C, et al (2015) Perivascular Adipose Tissue-Derived Adiponectin Inhibits Collar-Induced Carotid Atherosclerosis by Promoting Macrophage Autophagy. *PLOS ONE* 10:e0124031. doi: 10.1371/journal.pone.0124031
- Li FYL, Cheng KKY, Lam KSL, et al (2011) Cross-talk between adipose tissue and vasculature: role of adiponectin. *Acta Physiol (Oxf)* 203:167–180. doi: 10.1111/j.1748-1716.2010.02216.x
- Li H, Jamal J, Plaza C, et al (2014) Structures of human constitutive nitric oxide synthases. *Acta Crystallogr D Biol Crystallogr* 70:2667–2674. doi: 10.1107/S1399004714017064
- Li P-L, Zhang Y (2013) Cross talk between ceramide and redox signaling: implications for endothelial dysfunction and renal disease. *Handb Exp Pharmacol* 171–197. doi: 10.1007/978-3-7091-1511-4_9
- Li Y, Huang TT, Carlson EJ, et al (1995) Dilated cardiomyopathy and neonatal lethality in mutant mice lacking manganese superoxide dismutase. *Nat Genet* 11:376–381. doi: 10.1038/ng1295-376
- Li Y-L, Ding Y, Agnew C, Schultz HD (2008) Exercise training improves peripheral chemoreflex function in heart failure rabbits. *Journal of Applied Physiology* 105:782–790. doi: 10.1152/japplphysiol.90533.2008
- Libby P, DiCarli M, Weissleder R (2010a) The vascular biology of atherosclerosis and imaging targets. *J Nucl Med* 51 Suppl 1:33S–37S. doi: 10.2967/jnumed.109.069633
- Libby P, Okamoto Y, Rocha VZ, Folco E (2010b) Inflammation in atherosclerosis: transition from theory to practice. *Circ J* 74:213–220.
- Liggett SB, Cresci S, Kelly RJ, et al (2008) A GRK5 polymorphism that inhibits beta-adrenergic receptor signaling is protective in heart failure. *Nat Med* 14:510–517. doi: 10.1038/nm1750

- Lira FS, Rosa JC, Yamashita AS, et al (2009) Endurance training induces depot-specific changes in IL-10/TNF- α ratio in rat adipose tissue. *Cytokine* 45:80–85. doi: 10.1016/j.cyto.2008.10.018
- Liu R, Li B, Flanagan SW, et al (2002) Increased mitochondrial antioxidative activity or decreased oxygen free radical propagation prevent mutant SOD1-mediated motor neuron cell death and increase amyotrophic lateral sclerosis-like transgenic mouse survival. *J Neurochem* 80:488–500.
- Loader J, Montero D, Lorenzen C, et al (2015) Acute Hyperglycemia Impairs Vascular Function in Healthy and Cardiometabolic Diseased Subjects: Systematic Review and Meta-Analysis. *Arterioscler Thromb Vasc Biol* 35:2060–2072. doi: 10.1161/ATVBAHA.115.305530
- Löhn M, Dubrovska G, Lauterbach B, et al (2002) Periadventitial fat releases a vascular relaxing factor. *FASEB J* 16:1057–1063. doi: 10.1096/fj.02-0024com
- Lönnqvist F, Arner P, Nordfors L, Schalling M (1995) Overexpression of the obese (ob) gene in adipose tissue of human obese subjects. *Nat Med* 1:950–953.
- Loschen G, Azzi A (1975) On the formation of hydrogen peroxide and oxygen radicals in heart mitochondria. *Recent Adv Stud Cardiac Struct Metab* 7:3–12.
- Lozano I, Van der Werf R, Bietiger W, et al (2016) High-fructose and high-fat diet-induced disorders in rats: impact on diabetes risk, hepatic and vascular complications. *Nutr Metab (Lond)* 13:15. doi: 10.1186/s12986-016-0074-1
- Lu C, Zhao AX, Gao Y-J, Lee RMKW (2011) Modulation of vein function by perivascular adipose tissue. *European Journal of Pharmacology* 657:111–116. doi: 10.1016/j.ejphar.2010.12.028
- Lubos E, Loscalzo J, Handy DE (2010) Glutathione Peroxidase-1 in Health and Disease: From Molecular Mechanisms to Therapeutic Opportunities. *Antioxidants & Redox Signaling* 15:1957–1997. doi: 10.1089/ars.2010.3586
- Luquet S, Gaudel C, Holst D, et al (2005) Roles of PPAR delta in lipid absorption and metabolism: a new target for the treatment of type 2 diabetes. *Biochim Biophys Acta* 1740:313–317. doi: 10.1016/j.bbadis.2004.11.011

M

- Ma L, Ma S, He H, et al (2010) Perivascular fat-mediated vascular dysfunction and remodeling through the AMPK/mTOR pathway in high-fat diet-induced obese rats. *Hypertens Res* 33:446–453. doi: 10.1038/hr.2010.11
- Maeda N, Takahashi M, Funahashi T, et al (2001) PPARgamma ligands increase expression and plasma concentrations of adiponectin, an adipose-derived protein. *Diabetes* 50:2094–2099.

- Maenhaut N, Boydens C, Van de Voorde J (2010) Hypoxia enhances the relaxing influence of perivascular adipose tissue in isolated mice aorta. *Eur J Pharmacol* 641:207–212. doi: 10.1016/j.ejphar.2010.05.058
- Mah E, Bruno RS (2012) Postprandial hyperglycemia on vascular endothelial function: mechanisms and consequences. *Nutr Res* 32:727–740. doi: 10.1016/j.nutres.2012.08.002
- Mah E, Noh SK, Ballard KD, et al (2011) Postprandial hyperglycemia impairs vascular endothelial function in healthy men by inducing lipid peroxidation and increasing asymmetric dimethylarginine:arginine. *J Nutr* 141:1961–1968. doi: 10.3945/jn.111.144592
- Maiorana A, O'Driscoll G, Taylor R, Green D (2003) Exercise and the nitric oxide vasodilator system. *Sports Med* 33:1013–1035.
- Malik VS, Popkin BM, Bray GA, et al (2010) Sugar-sweetened beverages and risk of metabolic syndrome and type 2 diabetes: a meta-analysis. *Diabetes Care* 33:2477–2483. doi: 10.2337/dc10-1079
- Malliani A (1999) The Pattern of Sympathovagal Balance Explored in the Frequency Domain. *News Physiol Sci* 14:111–117.
- Mancia G, Bousquet P, Elghozi JL, et al (2007) The sympathetic nervous system and the metabolic syndrome. *J Hypertens* 25:909–920. doi: 10.1097/HJH.0b013e328048d004
- Marcell TJ, McAuley KA, Traustadóttir T, Reaven PD (2005) Exercise training is not associated with improved levels of C-reactive protein or adiponectin. *Metab Clin Exp* 54:533–541. doi: 10.1016/j.metabol.2004.11.008
- Marchesi C, Ebrahimian T, Angulo O, et al (2009) Endothelial Nitric Oxide Synthase Uncoupling and Perivascular Adipose Oxidative Stress and Inflammation Contribute to Vascular Dysfunction in a Rodent Model of Metabolic Syndrome. *Hypertension* 54:1384–1392. doi: 10.1161/HYPERTENSIONAHA.109.138305
- Martínez-Ruiz A, Villanueva L, Orduña CG de, et al (2005) S-nitrosylation of Hsp90 promotes the inhibition of its ATPase and endothelial nitric oxide synthase regulatory activities. *PNAS* 102:8525–8530. doi: 10.1073/pnas.0407294102
- Maurière P, Després JP, Prud'homme D, et al (1991) Regional variation in adipose tissue lipolysis in lean and obese men. *J Lipid Res* 32:1625–1633.
- Mauro CR, Ilonzo G, Nguyen BT, et al (2013) Attenuated adiposopathy in perivascular adipose tissue compared with subcutaneous human adipose tissue. *The American Journal of Surgery* 206:241–244. doi: 10.1016/j.amjsurg.2012.07.032
- Mehebik N, Jaubert A-M, Sabourault D, et al (2005) Leptin-induced nitric oxide production in white adipocytes is mediated through PKA and MAP kinase activation. *Am J Physiol, Cell Physiol* 289:C379-387. doi: 10.1152/ajpcell.00320.2004
- Meijer RI, Bakker W, Alta C-LAF, et al (2013) Perivascular Adipose Tissue Control of Insulin-Induced Vasoreactivity in Muscle Is Impaired in db/db Mice. *Diabetes*

- 62:590–598. doi: 10.2337/db11-1603
- Melikian N, Seddon MD, Casadei B, et al (2009) Neuronal Nitric Oxide Synthase and Human Vascular Regulation. *Trends in Cardiovascular Medicine* 19:256–262. doi: 10.1016/j.tcm.2010.02.007
- Melikian N, Wheatcroft SB, Ogah OS, et al (2007) Asymmetric dimethylarginine and reduced nitric oxide bioavailability in young Black African men. *Hypertension* 49:873–877. doi: 10.1161/01.HYP.0000258405.25330.80
- Mimura J, Yuasa F, Yuyama R, et al (2005) The effect of residential exercise training on baroreflex control of heart rate and sympathetic nerve activity in patients with acute myocardial infarction*. *Chest* 127:1108–1115. doi: 10.1378/chest.127.4.1108
- Minato K, Shiroya Y, Nakae Y, Kondo T The effect of chronic exercise on the rat pancreas. *International Journal of Pancreatology* 27:151–156. doi: 10.1385/IJGC:27:2:151
- Mitchell GF, Parise H, Vita JA, et al (2004) Local shear stress and brachial artery flow-mediated dilation: the Framingham Heart Study. *Hypertension* 44:134–139. doi: 10.1161/01.HYP.0000137305.77635.68
- Miyazaki H, Ohishi S, Ookawara T, et al Strenuous endurance training in humans reduces oxidative stress following exhausting exercise. *Eur J Appl Physiol* 84:1–6. doi: 10.1007/s004210000342
- Monnier L, Mas E, Ginet C, et al (2006) Activation of oxidative stress by acute glucose fluctuations compared with sustained chronic hyperglycemia in patients with type 2 diabetes. *JAMA* 295:1681–1687. doi: 10.1001/jama.295.14.1681
- Mora S, Cook N, Buring JE, et al (2007) Physical activity and reduced risk of cardiovascular events: potential mediating mechanisms. *Circulation* 116:2110–2118. doi: 10.1161/CIRCULATIONAHA.107.729939
- Moro MA, Russel RJ, Cellek S, et al (1996) cGMP mediates the vascular and platelet actions of nitric oxide: confirmation using an inhibitor of the soluble guanylyl cyclase. *Proc Natl Acad Sci USA* 93:1480–1485.
- Morris DL, Rui L (2009) Recent advances in understanding leptin signaling and leptin resistance. *Am J Physiol Endocrinol Metab* 297:E1247-1259. doi: 10.1152/ajpendo.00274.2009
- Motoshima H, Wu X, Mahadev K, Goldstein BJ (2004) Adiponectin suppresses proliferation and superoxide generation and enhances eNOS activity in endothelial cells treated with oxidized LDL. *Biochemical and Biophysical Research Communications* 315:264–271. doi: 10.1016/j.bbrc.2004.01.049
- Mourier A, Gautier JF, De Kerviler E, et al (1997) Mobilization of visceral adipose tissue related to the improvement in insulin sensitivity in response to physical training in NIDDM. Effects of branched-chain amino acid supplements. *Diabetes Care* 20:385–391.
- Muniyappa R, Sowers JR (2013) Role of insulin resistance in endothelial dysfunction. *Rev*

- Endocr Metab Disord 14:5–12. doi: 10.1007/s11154-012-9229-1
- Murray CI, Gebcka MA, Haile A, et al (2008) Abstract 3379: cGMP Specific Phosphodiesterase Type 5A Activity is Regulated by S-nitrosylation at Cys 181. Circulation 118:S_415-S_415.
- Myers J, Hadley D, Oswald U, et al (2007) Effects of exercise training on heart rate recovery in patients with chronic heart failure. Am Heart J 153:1056–1063. doi: 10.1016/j.ahj.2007.02.038
- N*
- Nara M, Kanda T, Tsukui S, et al (1999) Running exercise increases tumor necrosis factor- α secreting from mesenteric fat in insulin-resistant rats. Life Sciences 65:237–244. doi: 10.1016/S0024-3205(99)00242-8
- Nassis GP, Papantakou K, Skenderi K, et al (2005) Aerobic exercise training improves insulin sensitivity without changes in body weight, body fat, adiponectin, and inflammatory markers in overweight and obese girls. Metab Clin Exp 54:1472–1479. doi: 10.1016/j.metabol.2005.05.013
- Navarro A, Gomez C, Lopez-Cepero JM, Boveris A (2004) Beneficial effects of moderate exercise on mice aging: survival, behavior, oxidative stress, and mitochondrial electron transfer. American Journal of Physiology - Regulatory, Integrative and Comparative Physiology 286:R505–R511. doi: 10.1152/ajpregu.00208.2003
- Nedergaard J, Cannon B (2014) The Browning of White Adipose Tissue: Some Burning Issues. Cell Metab 20:396–407. doi: 10.1016/j.cmet.2014.07.005
- Nguyen Dinh Cat A, Montezano AC, Burger D, Touyz RM (2012) Angiotensin II, NADPH Oxidase, and Redox Signaling in the Vasculature. Antioxidants & Redox Signaling 19:1110–1120. doi: 10.1089/ars.2012.4641
- Nicklas BJ, Rogus EM, Goldberg AP (1997) Exercise blunts declines in lipolysis and fat oxidation after dietary-induced weight loss in obese older women. Am J Physiol 273:E149-155.
- Nilsson GE, Tenland T, Oberg PA (1980) Evaluation of a laser Doppler flowmeter for measurement of tissue blood flow. IEEE Trans Biomed Eng 27:597–604. doi: 10.1109/TBME.1980.326582
- Nishikawa T, Edelstein D, Du XL, et al (2000) Normalizing mitochondrial superoxide production blocks three pathways of hyperglycaemic damage. Nature 404:787–790. doi: 10.1038/35008121
- Nisoli E, Clementi E, Carruba MO, Moncada S (2007) Defective mitochondrial biogenesis: a hallmark of the high cardiovascular risk in the metabolic syndrome? Circ Res 100:795–806. doi: 10.1161/01.RES.0000259591.97107.6c

- Niu XF, Smith CW, Kubes P (1994) Intracellular oxidative stress induced by nitric oxide synthesis inhibition increases endothelial cell adhesion to neutrophils. *Circ Res* 74:1133–1140.
- Noguchi K, Matsuzaki T, Sakanashi M, et al (2015) Effect of caffeine contained in a cup of coffee on microvascular function in healthy subjects. *J Pharmacol Sci* 127:217–222. doi: 10.1016/j.jphs.2015.01.003
- Nojiri H, Shimizu T, Funakoshi M, et al (2006) Oxidative stress causes heart failure with impaired mitochondrial respiration. *J Biol Chem* 281:33789–33801. doi: 10.1074/jbc.M602118200
- Nonogaki K (2000) New insights into sympathetic regulation of glucose and fat metabolism. *Diabetologia* 43:533–549. doi: 10.1007/s001250051341
- Notarius CF, Millar PJ, Floras JS (2015) Muscle sympathetic activity in resting and exercising humans with and without heart failure. *Appl Physiol Nutr Metab* 40:1107–1115. doi: 10.1139/apnm-2015-0289
- Nour-Eldine W, Ghantous CM, Zibara K, et al (2016) Adiponectin Attenuates Angiotensin II-Induced Vascular Smooth Muscle Cell Remodeling through Nitric Oxide and the RhoA/ROCK Pathway. *Front Pharmacol* 7:86. doi: 10.3389/fphar.2016.00086



- Oates PJ (2002) Polyol pathway and diabetic peripheral neuropathy. *Int Rev Neurobiol* 50:325–392.
- O'Connor L, Imamura F, Lentjes MAH, et al (2015) Prospective associations and population impact of sweet beverage intake and type 2 diabetes, and effects of substitutions with alternative beverages. *Diabetologia* 58:1474–1483. doi: 10.1007/s00125-015-3572-1
- O'Neill S, O'Driscoll L (2015) Metabolic syndrome: a closer look at the growing epidemic and its associated pathologies. *Obes Rev* 16:1–12. doi: 10.1111/obr.12229
- Oswal A, Yeo G (2010) Leptin and the Control of Body Weight: A Review of Its Diverse Central Targets, Signaling Mechanisms, and Role in the Pathogenesis of Obesity. *Obesity* 18:221–229. doi: 10.1038/oby.2009.228
- Ouchi N, Ohishi M, Kihara S, et al (2003) Association of hypoadiponectinemia with impaired vasoreactivity. *Hypertension* 42:231–234. doi: 10.1161/01.HYP.0000083488.67550.B8
- Ouedraogo R, Wu X, Xu S-Q, et al (2006) Adiponectin suppression of high-glucose-induced reactive oxygen species in vascular endothelial cells: evidence for involvement of a cAMP signaling pathway. *Diabetes* 55:1840–1846. doi: 10.2337/db05-1174
- Owen MK, Witzmann FA, McKenney ML, et al (2013) Perivascular adipose tissue potentiates contraction of coronary vascular smooth muscle: influence of obesity.

Circulation 128:9–18. doi: 10.1161/CIRCULATIONAHA.112.001238

Ozkor MA, Rahman AM, Murrow JR, et al (2014) Differences in vascular nitric oxide and endothelium-derived hyperpolarizing factor bioavailability in blacks and whites. Arterioscler Thromb Vasc Biol 34:1320–1327. doi: 10.1161/ATVBAHA.113.303136

Oztasan N, Taysi S, Gumustekin K, et al (2004) Endurance training attenuates exercise-induced oxidative stress in erythrocytes in rat. Eur J Appl Physiol 91:622–627. doi: 10.1007/s00421-003-1029-6



Padilla J, Jenkins NT, Vieira-Potter VJ, Laughlin MH (2013) Divergent phenotype of rat thoracic and abdominal perivascular adipose tissues. American Journal of Physiology - Regulatory, Integrative and Comparative Physiology 304:R543–R552. doi: 10.1152/ajpregu.00567.2012

Palm F, Onozato ML, Luo Z, Wilcox CS (2007) Dimethylarginine dimethylaminohydrolase (DDAH): expression, regulation, and function in the cardiovascular and renal systems. American Journal of Physiology - Heart and Circulatory Physiology 293:H3227–H3245. doi: 10.1152/ajpheart.00998.2007

Panza JA, Epstein SE, Quyyumi AA (1991) Circadian variation in vascular tone and its relation to alpha-sympathetic vasoconstrictor activity. N Engl J Med 325:986–990. doi: 10.1056/NEJM199110033251402

Paolisso G, Manzella D, Rizzo MR, et al (2000) Elevated plasma fatty acid concentrations stimulate the cardiac autonomic nervous system in healthy subjects. Am J Clin Nutr 72:723–730.

Park J, Choi HJ, Lee S, et al (2000) Rac-related GTP-binding protein in elicitor-induced reactive oxygen generation by suspension-cultured soybean cells. Plant Physiol 124:725–732.

Patel KP, Zheng H (2012) Central neural control of sympathetic nerve activity in heart failure following exercise training. Am J Physiol Heart Circ Physiol 302:H527–537. doi: 10.1152/ajpheart.00676.2011

Pautz A, Art J, Hahn S, et al (2010) Regulation of the expression of inducible nitric oxide synthase. Nitric Oxide 23:75–93. doi: 10.1016/j.niox.2010.04.007

Payne GA, Bohlen HG, Dincer UD, et al (2009) Periadventitial adipose tissue impairs coronary endothelial function via PKC-beta-dependent phosphorylation of nitric oxide synthase. Am J Physiol Heart Circ Physiol 297:H460–465. doi: 10.1152/ajpheart.00116.2009

Payne GA, Borbouse L, Kumar S, et al (2010) Epicardial Perivascular Adipose-Derived Leptin Exacerbates Coronary Endothelial Dysfunction in Metabolic Syndrome via a

- Protein Kinase C-β Pathway. Arteriosclerosis, Thrombosis, and Vascular Biology 30:1711–1717. doi: 10.1161/ATVBAHA.110.210070
- Pedersen BK, Saltin B (2006) Evidence for prescribing exercise as therapy in chronic disease. Scand J Med Sci Sports 16 Suppl 1:3–63. doi: 10.1111/j.1600-0838.2006.00520.x
- Pepino MY (2015) Metabolic effects of non-nutritive sweeteners. Physiol Behav 152:450–455. doi: 10.1016/j.physbeh.2015.06.024
- Pescatello LS, Franklin BA, Fagard R, et al (2004) American College of Sports Medicine position stand. Exercise and hypertension. Med Sci Sports Exerc 36:533–553.
- Petrovic N, Walden TB, Shabalina IG, et al (2010) Chronic peroxisome proliferator-activated receptor gamma (PPARgamma) activation of epididymally derived white adipocyte cultures reveals a population of thermogenically competent, UCP1-containing adipocytes molecularly distinct from classic brown adipocytes. J Biol Chem 285:7153–7164. doi: 10.1074/jbc.M109.053942
- Piazza M, Guillemette JG, Dieckmann T (2015) Dynamics of Nitric Oxide Synthase–Calmodulin Interactions at Physiological Calcium Concentrations. Biochemistry 54:1989–2000. doi: 10.1021/bi501353s
- Pierpont GL, Stolpman DR, Gornick CC (2000) Heart rate recovery post-exercise as an index of parasympathetic activity. J Auton Nerv Syst 80:169–174.
- Pilon G, Dallaire P, Marette A (2004) Inhibition of inducible nitric-oxide synthase by activators of AMP-activated protein kinase: a new mechanism of action of insulin-sensitizing drugs. J Biol Chem 279:20767–20774. doi: 10.1074/jbc.M401390200
- Polak J, Klimcakova E, Moro C, et al (2006) Effect of aerobic training on plasma levels and subcutaneous abdominal adipose tissue gene expression of adiponectin, leptin, interleukin 6, and tumor necrosis factor alpha in obese women. Metab Clin Exp 55:1375–1381. doi: 10.1016/j.metabol.2006.06.008
- Police SB, Thatcher SE, Charnigo R, et al (2009) Obesity Promotes Inflammation in Periaortic Adipose Tissue and Angiotensin II-Induced Abdominal Aortic Aneurysm Formation. Arteriosclerosis, Thrombosis, and Vascular Biology 29:1458–1464. doi: 10.1161/ATVBAHA.109.192658
- Potenza MA, Addabbo F, Montagnani M (2009a) Vascular actions of insulin with implications for endothelial dysfunction. Am J Physiol Endocrinol Metab 297:E568–577. doi: 10.1152/ajpendo.00297.2009
- Potenza MA, Addabbo F, Montagnani M (2009b) Vascular actions of insulin with implications for endothelial dysfunction. American Journal of Physiology - Endocrinology and Metabolism 297:E568–E577. doi: 10.1152/ajpendo.00297.2009
- Procopio C, Andreozzi F, Laratta E, et al (2009) Leptin-Stimulated Endothelial Nitric-Oxide Synthase via an Adenosine 5'-Monophosphate-Activated Protein Kinase/Akt Signaling Pathway Is Attenuated by Interaction with C-Reactive Protein. Endocrinology 150:3584–3593. doi: 10.1210/en.2008-0921

Q

Qi C, Pekala PH (2000) Tumor Necrosis Factor- α -Induced Insulin Resistance in Adipocytes. Proceedings of the Society for Experimental Biology and Medicine 223:128–135. doi: 10.1111/j.1525-1373.2000.22318.x

R

Radomski MW, Moncada S (1993) Regulation of vascular homeostasis by nitric oxide. Thromb Haemost 70:36–41.

Ranallo RF, Rhodes EC (1998) Lipid metabolism during exercise. Sports Med 26:29–42.

Randle PJ, Garland PB, Hales CN, Newsholme EA (1963) The glucose fatty-acid cycle. Its role in insulin sensitivity and the metabolic disturbances of diabetes mellitus. Lancet 1:785–789.

Rao GH, Krishnamurthi S, Raji L, White JG (1990) Influence of nitric oxide on agonist-mediated calcium mobilization in platelets. Biochem Med Metab Biol 43:271–275.

Ravi K, Brennan LA, Levic S, et al (2004) S-nitrosylation of endothelial nitric oxide synthase is associated with monomerization and decreased enzyme activity. PNAS 101:2619–2624. doi: 10.1073/pnas.0300464101

Reaven GM (1988) Banting lecture 1988. Role of insulin resistance in human disease. Diabetes 37:1595–1607.

Rebolledo A, Rebolledo OR, Marra CA, et al (2010) Early alterations in vascular contractility associated to changes in fatty acid composition and oxidative stress markers in perivascular adipose tissue. Cardiovascular Diabetology 9:65. doi: 10.1186/1475-2840-9-65

Reifenberger MS, Turk JR, Newcomer SC, et al (2007) Perivascular fat alters reactivity of coronary artery: effects of diet and exercise. Med Sci Sports Exerc 39:2125–2134. doi: 10.1249/mss.0b013e318156e9df

Rengo G, Pagano G, Parisi V, et al (2014) Changes of plasma norepinephrine and serum N-terminal pro-brain natriuretic peptide after exercise training predict survival in patients with heart failure. Int J Cardiol 171:384–389. doi: 10.1016/j.ijcard.2013.12.024

Reseland JE, Anderssen SA, Solvoll K, et al (2001) Effect of long-term changes in diet and

- exercise on plasma leptin concentrations. *Am J Clin Nutr* 73:240–245.
- Ribiere C, Jaubert AM, Gaudiot N, et al (1996) White adipose tissue nitric oxide synthase: a potential source for NO production. *Biochem Biophys Res Commun* 222:706–712. doi: 10.1006/bbrc.1996.0824
- Richterova B, Stich V, Moro C, et al (2004) Effect of endurance training on adrenergic control of lipolysis in adipose tissue of obese women. *J Clin Endocrinol Metab* 89:1325–1331. doi: 10.1210/jc.2003-031001
- Ristow M, Zarse K, Oberbach A, et al (2009) Antioxidants prevent health-promoting effects of physical exercise in humans. *Proc Natl Acad Sci USA* 106:8665–8670. doi: 10.1073/pnas.0903485106
- Rittig K, Staib K, Machann J, et al (2008) Perivascular fatty tissue at the brachial artery is linked to insulin resistance but not to local endothelial dysfunction. *Diabetologia* 51:2093–2099. doi: 10.1007/s00125-008-1128-3
- Roque FR, Hernanz R, Salaices M, Briones AM (2013) Exercise training and cardiometabolic diseases: focus on the vascular system. *Curr Hypertens Rep* 15:204–214. doi: 10.1007/s11906-013-0336-5
- Ross R (1999) Atherosclerosis is an inflammatory disease. *Am Heart J* 138:S419–420.
- Rossi M, Maurizio S, Carpi A (2005) Skin blood flow motion response to insulin iontophoresis in normal subjects. *Microvascular Research* 70:17–22. doi: 10.1016/j.mvr.2005.05.001
- Rothwell NJ, Stock MJ (1979) A role for brown adipose tissue in diet-induced thermogenesis. *Nature* 281:31–35.
- Roustit M, Blaise S, Millet C, Cracowski JL (2010) Reproducibility and methodological issues of skin post-occlusive and thermal hyperemia assessed by single-point laser Doppler flowmetry. *Microvasc Res* 79:102–108. doi: 10.1016/j.mvr.2010.01.001
- Roustit M, Cracowski J-L (2013) Assessment of endothelial and neurovascular function in human skin microcirculation. *Trends Pharmacol Sci* 34:373–384. doi: 10.1016/j.tips.2013.05.007
- Roy D, Perreault M, Marette A (1998) Insulin stimulation of glucose uptake in skeletal muscles and adipose tissues in vivo is NO dependent. *Am J Physiol* 274:E692–699.



Sakurai T, Izawa T, Kizaki T, et al (2009) Exercise training decreases expression of inflammation-related adipokines through reduction of oxidative stress in rat white adipose tissue. *Biochemical and Biophysical Research Communications* 379:605–609. doi: 10.1016/j.bbrc.2008.12.127

Sakurai T, Takei M, Ogasawara J, et al (2005) Exercise Training Enhances Tumor Necrosis

- Factor- α -Induced Expressions of Anti-Apoptotic Genes without Alterations in Caspase-3 Activity in Rat Epididymal Adipocytes. *The Japanese Journal of Physiology* 55:181–189. doi: 10.2170/jjphysiol.R2096
- Sander M, Chavoshan B, Victor RG (1999) A large blood pressure-raising effect of nitric oxide synthase inhibition in humans. *Hypertension* 33:937–942.
- Sato Y, Nagasaki M, Nakai N, Fushimi T (2003) Physical exercise improves glucose metabolism in lifestyle-related diseases. *Exp Biol Med (Maywood)* 228:1208–1212.
- Scherrer U, Sartori C (1997) Insulin as a vascular and sympathoexcitatory hormone: implications for blood pressure regulation, insulin sensitivity, and cardiovascular morbidity. *Circulation* 96:4104–4113.
- Schlaich MP, Lambert E, Kaye DM, et al (2004) Sympathetic augmentation in hypertension: role of nerve firing, norepinephrine reuptake, and Angiotensin neuromodulation. *Hypertension* 43:169–175. doi: 10.1161/01.HYP.0000103160.35395.9E
- Schlossmann J, Ammendola A, Ashman K, et al (2000) Regulation of intracellular calcium by a signalling complex of IRAG, IP3 receptor and cGMP kinase I β . *Nature* 404:197–201. doi: 10.1038/35004606
- Schwartz RS, Shuman WP, Larson V, et al (1991) The effect of intensive endurance exercise training on body fat distribution in young and older men. *Metab Clin Exp* 40:545–551.
- Schwarz PM, Kleinert H, Förstermann U (1999) Potential functional significance of brain-type and muscle-type nitric oxide synthase I expressed in adventitia and media of rat aorta. *Arterioscler Thromb Vasc Biol* 19:2584–2590.
- Sebai M, Lu S, Xiang L, Hester RL (2011) Improved functional vasodilation in obese Zucker rats following exercise training. *Am J Physiol Heart Circ Physiol* 301:H1090–1096. doi: 10.1152/ajpheart.00233.2011
- Seddon M, Melikian N, Dworakowski R, et al (2009) Effects of Neuronal Nitric Oxide Synthase on Human Coronary Artery Diameter and Blood Flow In Vivo. *Circulation* 119:2656–2662. doi: 10.1161/CIRCULATIONAHA.108.822205
- Seddon MD, Chowienczyk PJ, Brett SE, et al (2008) Neuronal Nitric Oxide Synthase Regulates Basal Microvascular Tone in Humans In Vivo. *Circulation* 117:1991–1996. doi: 10.1161/CIRCULATIONAHA.107.744540
- Segal SS (1992) Communication Among Endothelial and Smooth Muscle Cells Coordinates Blood Flow Control During Exercise. *Physiology* 7:152–156.
- Segal SS, Kurjiaka DT (1995) Coordination of blood flow control in the resistance vasculature of skeletal muscle. *Med Sci Sports Exerc* 27:1158–1164.
- Seligman BGS, Polanczyk CA, Santos ASB, et al (2011) Intensive practical lifestyle intervention improves endothelial function in metabolic syndrome independent of weight loss: a randomized controlled trial. *Metab Clin Exp* 60:1736–1740. doi: 10.1016/j.metabol.2011.05.006
- Serné EH, IJzerman RG, Gans ROB, et al (2002) Direct evidence for insulin-induced

- capillary recruitment in skin of healthy subjects during physiological hyperinsulinemia. *Diabetes* 51:1515–1522.
- Shima K, Zhu M, Noma Y, et al (1997) Exercise training in Otsuka Long-Evans Tokushima Fatty rat, a model of spontaneous non-insulin-dependent diabetes mellitus: effects on the B-cell mass, insulin content and fibrosis in the pancreas. *Diabetes Res Clin Pract* 35:11–19.
- Shimabukuro M, Higa N, Asahi T, et al (2003) Hypoadiponectinemia is closely linked to endothelial dysfunction in man. *J Clin Endocrinol Metab* 88:3236–3240. doi: 10.1210/jc.2002-021883
- Shimomura I, Matsuda M, Hammer RE, et al (2000) Decreased IRS-2 and increased SREBP-1c lead to mixed insulin resistance and sensitivity in livers of lipodystrophic and ob/ob mice. *Mol Cell* 6:77–86.
- Shulman GI (2000) Cellular mechanisms of insulin resistance. *J Clin Invest* 106:171–176. doi: 10.1172/JCI10583
- Shyy JY-J, Chien S (2002) Role of integrins in endothelial mechanosensing of shear stress. *Circ Res* 91:769–775.
- Sigal RJ, Kenny GP, Wasserman DH, Castaneda-Sceppa C (2004) Physical activity/exercise and type 2 diabetes. *Diabetes Care* 27:2518–2539.
- Silva BR, Pernomian L, Grando MD, Bendhack LM (2014) Phenylephrine activates eNOS Ser 1177 phosphorylation and nitric oxide signaling in renal hypertensive rat aorta. *Eur J Pharmacol* 738:192–199. doi: 10.1016/j.ejphar.2014.05.040
- Simmerman HK, Jones LR (1998) Phospholamban: protein structure, mechanism of action, and role in cardiac function. *Physiol Rev* 78:921–947.
- Sindler AL, Delp MD, Reyes R, et al (2009) Effects of ageing and exercise training on eNOS uncoupling in skeletal muscle resistance arterioles. *J Physiol (Lond)* 587:3885–3897. doi: 10.1113/jphysiol.2009.172221
- Singh GM, Micha R, Khatibzadeh S, et al (2015) Estimated Global, Regional, and National Disease Burdens Related to Sugar-Sweetened Beverage Consumption in 2010. *Circulation* 132:639–666. doi: 10.1161/CIRCULATIONAHA.114.010636
- Singh JP, Kandala J, Camm AJ (2014) Non-pharmacological modulation of the autonomic tone to treat heart failure. *Eur Heart J* 35:77–85. doi: 10.1093/eurheartj/eht436
- Skalicky J, Muzakova V, Kandar R, et al (2008) Evaluation of oxidative stress and inflammation in obese adults with metabolic syndrome. *Clin Chem Lab Med* 46:499–505. doi: 10.1515/CCLM.2008.096
- Skrapari I, Tentolouris N, Perrea D, et al (2007) Baroreflex sensitivity in obesity: relationship with cardiac autonomic nervous system activity. *Obesity (Silver Spring)* 15:1685–1693. doi: 10.1038/oby.2007.201
- Smith CJ, Sun D, Hoegler C, et al (1996) Reduced gene expression of vascular endothelial NO synthase and cyclooxygenase-1 in heart failure. *Circ Res* 78:58–64.

- Soltis EE, Cassis LA (1991) Influence of perivascular adipose tissue on rat aortic smooth muscle responsiveness. *Clin Exp Hypertens A* 13:277–296.
- Souza EG, De Lorenzo A, Huguenin G, et al (2014) Impairment of systemic microvascular endothelial and smooth muscle function in individuals with early-onset coronary artery disease: studies with laser speckle contrast imaging. *Coron Artery Dis* 25:23–28. doi: 10.1097/MCA.000000000000055
- Sowers JR (2002) Hypertension, angiotensin II, and oxidative stress. *N Engl J Med* 346:1999–2001. doi: 10.1056/NEJMMe020054
- Sriram K, Laughlin JG, Rangamani P, Tartakovsky DM (2016) Shear-Induced Nitric Oxide Production by Endothelial Cells. *Biophysical Journal* 111:208–221. doi: 10.1016/j.bpj.2016.05.034
- Stallknecht B, Vinent J, Ploug T, Galbo H (1991) Increased activities of mitochondrial enzymes in white adipose tissue in trained rats. *American Journal of Physiology - Endocrinology and Metabolism* 261:E410–E414.
- Stamler JS, Meissner G (2001) Physiology of nitric oxide in skeletal muscle. *Physiol Rev* 81:209–237.
- Stanford KI, Middelbeek RJW, Goodyear LJ (2015) Exercise Effects on White Adipose Tissue: Beiging and Metabolic Adaptations. *Diabetes* 64:2361–2368. doi: 10.2337/db15-0227
- Stas SN, El-Atat FA, Sowers JR (2004) Pathogenesis of hypertension in diabetes. *Rev Endocr Metab Disord* 5:221–225. doi: 10.1023/B:REMD.0000032410.75638.da
- Steinberg HO, Chaker H, Leaming R, et al (1996) Obesity/insulin resistance is associated with endothelial dysfunction. Implications for the syndrome of insulin resistance. *J Clin Invest* 97:2601–2610. doi: 10.1172/JCI118709
- Straub AC, Billaud M, Johnstone SR, et al (2011) Compartmentalized connexin 43 s-nitrosylation/denitrosylation regulates heterocellular communication in the vessel wall. *Arterioscler Thromb Vasc Biol* 31:399–407. doi: 10.1161/ATVBAHA.110.215939
- Straub AC, Butcher JT, Billaud M, et al (2014) Hemoglobin α /eNOS coupling at myoendothelial junctions is required for nitric oxide scavenging during vasoconstriction. *Arterioscler Thromb Vasc Biol* 34:2594–2600. doi: 10.1161/ATVBAHA.114.303974
- Straznicky NE, Lambert EA, Lambert GW, et al (2005) Effects of dietary weight loss on sympathetic activity and cardiac risk factors associated with the metabolic syndrome. *J Clin Endocrinol Metab* 90:5998–6005. doi: 10.1210/jc.2005-0961
- Sun J, Xu Y, Dai Z, Sun Y (2009) Intermittent high glucose enhances proliferation of vascular smooth muscle cells by upregulating osteopontin. *Mol Cell Endocrinol* 313:64–69. doi: 10.1016/j.mce.2009.08.019
- Sun X, Hou N, Han F, et al (2013) Effect of high free fatty acids on the anti-contractile

- response of perivascular adipose tissue in rat aorta. *Journal of Molecular and Cellular Cardiology* 63:169–174. doi: 10.1016/j.yjmcc.2013.07.018
- Sutherland LN, Bomhof MR, Capozzi LC, et al (2009) Exercise and adrenaline increase PGC-1 α mRNA expression in rat adipose tissue. *J Physiol (Lond)* 587:1607–1617. doi: 10.1113/jphysiol.2008.165464
- Suzuki K, Olah G, Modis K, et al (2011) Hydrogen sulfide replacement therapy protects the vascular endothelium in hyperglycemia by preserving mitochondrial function. *PNAS* 108:13829–13834. doi: 10.1073/pnas.1105121108
- Szasz T, Bomfim GF, Webb RC (2013) The influence of perivascular adipose tissue on vascular homeostasis. *Vasc Health Risk Manag* 9:105–116. doi: 10.2147/VHRM.S33760

T

- Tan KCB, Xu A, Chow WS, et al (2004) Hypoadiponectinemia Is Associated with Impaired Endothelium-Dependent Vasodilation. *The Journal of Clinical Endocrinology & Metabolism* 89:765–769. doi: 10.1210/jc.2003-031012
- Tanaka T, Nakatani K, Morioka K, et al (2003) Nitric oxide stimulates glucose transport through insulin-independent GLUT4 translocation in 3T3-L1 adipocytes. *Eur J Endocrinol* 149:61–67.
- Tankersley CG, Campen M, Bierman A, et al (2004) Particle effects on heart-rate regulation in senescent mice. *Inhal Toxicol* 16:381–390. doi: 10.1080/08958370490439551
- Tanoue A, Koba M, Miyawaki S, et al (2002a) Role of the alpha1D-adrenergic receptor in the development of salt-induced hypertension. *Hypertension* 40:101–106.
- Tanoue A, Nasa Y, Koshimizu T, et al (2002b) The alpha(1D)-adrenergic receptor directly regulates arterial blood pressure via vasoconstriction. *J Clin Invest* 109:765–775. doi: 10.1172/JCI14001
- Tarvainen MP, Niskanen J-P, Lipponen JA, et al (2014) Kubios HRV--heart rate variability analysis software. *Comput Methods Programs Biomed* 113:210–220. doi: 10.1016/j.cmpb.2013.07.024
- Tedesco L, Valerio A, Dossena M, et al (2010) Cannabinoid receptor stimulation impairs mitochondrial biogenesis in mouse white adipose tissue, muscle, and liver: the role of eNOS, p38 MAPK, and AMPK pathways. *Diabetes* 59:2826–2836. doi: 10.2337/db09-1881
- Teixeira de Lemos E, Reis F, Baptista S, et al (2009) Exercise training decreases proinflammatory profile in Zucker diabetic (type 2) fatty rats. *Nutrition* 25:330–339. doi: 10.1016/j.nut.2008.08.014
- Tentolouris N, Liatis S, Katsilambros N (2006) Sympathetic system activity in obesity and

- metabolic syndrome. *Ann N Y Acad Sci* 1083:129–152. doi: 10.1196/annals.1367.010
- Tesfamariam B, Weisbrod RM, Cohen RA (1987) Endothelium inhibits responses of rabbit carotid artery to adrenergic nerve stimulation. *Am J Physiol* 253:H792-798.
- Thijssen DHJ, Black MA, Pyke KE, et al (2011) Assessment of flow-mediated dilation in humans: a methodological and physiological guideline. *Am J Physiol Heart Circ Physiol* 300:H2-12. doi: 10.1152/ajpheart.00471.2010
- Thijssen DHJ, Maiorana AJ, O'Driscoll G, et al (2010) Impact of inactivity and exercise on the vasculature in humans. *Eur J Appl Physiol* 108:845–875. doi: 10.1007/s00421-009-1260-x
- Thireau J, Aimond F, Poisson D, et al (2010) New insights into sexual dimorphism during progression of heart failure and rhythm disorders. *Endocrinology* 151:1837–1845. doi: 10.1210/en.2009-1184
- Thireau J, Karam S, Roberge S, et al (2014) B-adrenergic blockade combined with subcutaneous B-type natriuretic peptide: a promising approach to reduce ventricular arrhythmia in heart failure? *Heart* 100:833–841. doi: 10.1136/heartjnl-2013-305167
- Thireau J, Poisson D, Zhang BL, et al (2008a) Increased heart rate variability in mice overexpressing the Cu/Zn superoxide dismutase. *Free Radic Biol Med* 45:396–403. doi: 10.1016/j.freeradbiomed.2008.04.020
- Thireau J, Zhang BL, Poisson D, Babuty D (2008b) Heart rate variability in mice: a theoretical and practical guide. *Exp Physiol* 93:83–94. doi: 10.1113/expphysiol.2007.040733
- Thomas EL, Brynes AE, McCarthy J, et al (2000) Preferential loss of visceral fat following aerobic exercise, measured by magnetic resonance imaging. *Lipids* 35:769–776.
- Tilg H, Moschen AR (2006) Adipocytokines: mediators linking adipose tissue, inflammation and immunity. *Nat Rev Immunol* 6:772–783. doi: 10.1038/nri1937
- Tominaga M, Eguchi H, Manaka H, et al (1999) Impaired glucose tolerance is a risk factor for cardiovascular disease, but not impaired fasting glucose. The Funagata Diabetes Study. *Diabetes Care* 22:920–924.
- Touati S, Meziri F, Devaux S, et al (2011) Exercise reverses metabolic syndrome in high-fat diet-induced obese rats. *Med Sci Sports Exerc* 43:398–407. doi: 10.1249/MSS.0b013e3181eeb12d
- Townsend N, Nichols M, Scarborough P, Rayner M (2015) Cardiovascular disease in Europe — epidemiological update 2015. *European Heart Journal* ehv428. doi: 10.1093/eurheartj/ehv428
- Trevellin E, Scorzeto M, Olivieri M, et al (2014) Exercise Training Induces Mitochondrial Biogenesis and Glucose Uptake in Subcutaneous Adipose Tissue Through eNOS-Dependent Mechanisms. *Diabetes* 63:2800–2811. doi: 10.2337/db13-1234
- Triposkiadis F, Parissis JT, Starling RC, et al (2009) Current drugs and medical treatment algorithms in the management of acute decompensated heart failure. *Expert Opin*

Investig Drugs 18:695–707. doi: 10.1517/13543780902922660

Tsuchida A, Yamauchi T, Ito Y, et al (2004) Insulin/Foxo1 pathway regulates expression levels of adiponectin receptors and adiponectin sensitivity. J Biol Chem 279:30817–30822. doi: 10.1074/jbc.M402367200

Tuomilehto J, Lindström J, Eriksson JG, et al (2001) Prevention of type 2 diabetes mellitus by changes in lifestyle among subjects with impaired glucose tolerance. N Engl J Med 344:1343–1350. doi: 10.1056/NEJM200105033441801

Tuttle JL, Falcone JC (2001) Nitric oxide release during alpha1-adrenoceptor-mediated constriction of arterioles. Am J Physiol Heart Circ Physiol 281:H873-881.



Vallance P, Leiper J (2004) Cardiovascular Biology of the Asymmetric Dimethylarginine:Dimethylarginine Dimethylaminohydrolase Pathway. Arteriosclerosis, Thrombosis, and Vascular Biology 24:1023–1030. doi: 10.1161/01.ATV.0000128897.54893.26

Verlohren S, Dubrovska G, Tsang S-Y, et al (2004) Visceral periadventitial adipose tissue regulates arterial tone of mesenteric arteries. Hypertension 44:271–276. doi: 10.1161/01.HYP.0000140058.28994.ec

Vernochet C, Mourier A, Bezy O, et al (2012) Adipose-Specific Deletion of TFAM Increases Mitochondrial Oxidation and Protects Mice against Obesity and Insulin Resistance. Cell Metabolism 16:765–776. doi: 10.1016/j.cmet.2012.10.016

Vieira VJ, Valentine RJ, Wilund KR, et al (2009) Effects of exercise and low-fat diet on adipose tissue inflammation and metabolic complications in obese mice. American Journal of Physiology - Endocrinology and Metabolism 296:E1164–E1171. doi: 10.1152/ajpendo.00054.2009

Villarroya F, Peyrou M, Giralt M (2016) Transcriptional regulation of the uncoupling protein-1 gene. Biochimie. doi: 10.1016/j.biichi.2016.09.017

Virdis A, Duranti E, Rossi C, et al (2015) Tumour necrosis factor-alpha participates on the endothelin-1/nitric oxide imbalance in small arteries from obese patients: role of perivascular adipose tissue. European Heart Journal 36:784–794. doi: 10.1093/eurheartj/ehu072

Vollmer RR (1996) Selective neural regulation of epinephrine and norepinephrine cells in the adrenal medulla -- cardiovascular implications. Clin Exp Hypertens 18:731–751.



- Wagner R, Machann J, Lehmann R, et al (2012) Exercise-induced albuminuria is associated with perivascular renal sinus fat in individuals at increased risk of type 2 diabetes. *Diabetologia* 55:2054–2058. doi: 10.1007/s00125-012-2551-z
- Walther G, Obert P, Dutheil F, et al (2015) Metabolic syndrome individuals with and without type 2 diabetes mellitus present generalized vascular dysfunction: cross-sectional study. *Arterioscler Thromb Vasc Biol* 35:1022–1029. doi: 10.1161/ATVBAHA.114.304591
- Wang X, You T, Murphy K, et al (2015) Addition of Exercise Increases Plasma Adiponectin and Release from Adipose Tissue. *Med Sci Sports Exerc* 47:2450–2455. doi: 10.1249/MSS.0000000000000670
- Weiss EP, Arif H, Villareal DT, et al (2008) Endothelial function after high-sugar-food ingestion improves with endurance exercise performed on the previous day. *Am J Clin Nutr* 88:51–57.
- Wen Y, Skidmore JC, Porter-Turner MM, et al (2002) Relationship of glycation, antioxidant status and oxidative stress to vascular endothelial damage in diabetes. *Diabetes Obes Metab* 4:305–308.
- Weston AH, Egner I, Dong Y, et al (2013) Stimulated release of a hyperpolarizing factor (ADHF) from mesenteric artery perivascular adipose tissue: involvement of myocyte BKCa channels and adiponectin. *Br J Pharmacol* 169:1500–1509. doi: 10.1111/bph.12157
- Widder JD, Chen W, Li L, et al (2007) Regulation of Tetrahydrobiopterin Biosynthesis by Shear Stress. *Circulation Research* 101:830–838. doi: 10.1161/CIRCRESAHA.107.153809
- Woerdeman J, Meijer RI, Eringa EC, et al (2016) Insulin Sensitivity Determines Effects of Insulin and Meal Ingestion on Systemic Vascular Resistance in Healthy Subjects. *Microcirculation* 23:62–68. doi: 10.1111/micc.12258
- Wójcicka G, Jamroz-Wiśniewska A, Atanasova P, et al (2011) Differential effects of statins on endogenous H₂S formation in perivascular adipose tissue. *Pharmacol Res* 63:68–76. doi: 10.1016/j.phrs.2010.10.011
- Wolf AM, Wolf D, Rumpold H, et al (2004) Adiponectin induces the anti-inflammatory cytokines IL-10 and IL-1RA in human leukocytes. *Biochemical and Biophysical Research Communications* 323:630–635. doi: 10.1016/j.bbrc.2004.08.145
- Woodman CR, Muller JM, Laughlin MH, Price EM (1997) Induction of nitric oxide synthase mRNA in coronary resistance arteries isolated from exercise-trained pigs. *Am J Physiol* 273:H2575-2579.
- Wray DW, Uberoi A, Lawrenson L, Richardson RS (2005) Heterogeneous limb vascular responsiveness to shear stimuli during dynamic exercise in humans. *J Appl Physiol* 99:81–86. doi: 10.1152/japplphysiol.01285.2004
- Wu G, Collins JK, Perkins-Veazie P, et al (2007) Dietary supplementation with watermelon pomace juice enhances arginine availability and ameliorates the metabolic syndrome

in Zucker diabetic fatty rats. *J Nutr* 137:2680–2685.

Wu G, Morris SM (1998) Arginine metabolism: nitric oxide and beyond. *Biochem J* 336 (Pt 1):1–17.

Wu J, Boström P, Sparks LM, et al (2012) Beige adipocytes are a distinct type of thermogenic fat cell in mouse and human. *Cell* 150:366–376. doi: 10.1016/j.cell.2012.05.016

X

Xi B, Huang Y, Reilly KH, et al (2015) Sugar-sweetened beverages and risk of hypertension and CVD: a dose-response meta-analysis. *Br J Nutr* 113:709–717. doi: 10.1017/S0007114514004383

Xi W, Satoh H, Kase H, et al (2005) Stimulated HSP90 binding to eNOS and activation of the PI3-Akt pathway contribute to globular adiponectin-induced NO production: vasorelaxation in response to globular adiponectin. *Biochem Biophys Res Commun* 332:200–205. doi: 10.1016/j.bbrc.2005.04.111

Xia N, Horke S, Habermeier A, et al (2016) Uncoupling of Endothelial Nitric Oxide Synthase in Perivascular Adipose Tissue of Diet-Induced Obese MiceSignificance. *Arteriosclerosis, Thrombosis, and Vascular Biology* 36:78–85. doi: 10.1161/ATVBAHA.115.306263

Xiang L, Naik J, Hester RL (2005) Exercise-induced increase in skeletal muscle vasodilatory responses in obese Zucker rats. *Am J Physiol Regul Integr Comp Physiol* 288:R987–991. doi: 10.1152/ajpregu.00702.2004

Xu A, Vanhoutte PM (2012) Adiponectin and adipocyte fatty acid binding protein in the pathogenesis of cardiovascular disease. *Am J Physiol Heart Circ Physiol* 302:H1231–1240. doi: 10.1152/ajpheart.00765.2011

Y

Yang A-L, Lo C-W, Lee J-T, Su C-T (2011) Enhancement of vasorelaxation in hypertension following high-intensity exercise. *Chin J Physiol* 54:87–95.

Yang H, Roberts LJ, Shi MJ, et al (2004) Retardation of Atherosclerosis by Overexpression of Catalase or Both Cu/Zn-Superoxide Dismutase and Catalase in Mice Lacking Apolipoprotein E. *Circulation Research* 95:1075–1081. doi: 10.1161/01.RES.0000149564.49410.0d

Yang H, Zhou L, Wang Z, et al (2009) Overexpression of antioxidant enzymes in ApoE-deficient mice suppresses benzo(a)pyrene-accelerated atherosclerosis. *Atherosclerosis* 207:51–58. doi: 10.1016/j.atherosclerosis.2009.03.052

- Ye L, Wu J, Cohen P, et al (2013) Fat cells directly sense temperature to activate thermogenesis. *Proc Natl Acad Sci USA* 110:12480–12485. doi: 10.1073/pnas.1310261110
- You T, Murphy KM, Lyles MF, et al (2006) Addition of aerobic exercise to dietary weight loss preferentially reduces abdominal adipocyte size. *Int J Obes (Lond)* 30:1211–1216. doi: 10.1038/sj.ijo.0803245
- Young CN, Fisher JP, Gallagher KM, et al (2009) Inhibition of nitric oxide synthase evokes central sympatho-excitation in healthy humans. *J Physiol (Lond)* 587:4977–4986. doi: 10.1113/jphysiol.2009.177204
- Zahlten RN, Hagler HK, Nejtek ME, Day CJ (1978) Morphological characterization of Kupffer and endothelial cells of rat liver isolated by counterflow elutriation. *Gastroenterology* 75:80–87.
- Zhang H, Zhao J, Yu H, Guo D (2016) Genistein ameliorated endothelial nitric oxidase synthase uncoupling by stimulating sirtuin-1 pathway in ox-LDL-injured HUVECs. *Environmental Toxicology and Pharmacology* 42:118–124. doi: 10.1016/j.etap.2016.01.011
- Zhang K, Zucker IH, Patel KP (1998) Altered number of diaphorase (NOS) positive neurons in the hypothalamus of rats with heart failure. *Brain Res* 786:219–225.
- Zhang Q-J, McMillin SL, Tanner JM, et al (2009) Endothelial nitric oxide synthase phosphorylation in treadmill-running mice: role of vascular signalling kinases. *The Journal of Physiology* 587:3911–3920. doi: 10.1113/jphysiol.2009.172916
- Zhao Y, Vanhoutte PM, Leung SWS (2015) Vascular nitric oxide: Beyond eNOS. *J Pharmacol Sci* 129:83–94. doi: 10.1016/j.jphs.2015.09.002
- Zheng F, Lu W, Jia C, et al (2010) Relationships between glucose excursion and the activation of oxidative stress in patients with newly diagnosed type 2 diabetes or impaired glucose regulation. *Endocrine* 37:201–208. doi: 10.1007/s12020-009-9296-6
- Zhu M, Wen M, Sun X, et al (2015) Propofol protects against high glucose-induced endothelial apoptosis and dysfunction in human umbilical vein endothelial cells. *Anesth Analg* 120:781–789. doi: 10.1213/ANE.0000000000000616
- Zhu W, Cheng KKY, Vanhoutte PM, et al (2008) Vascular effects of adiponectin: molecular mechanisms and potential therapeutic intervention. *Clinical Science* 114:361–374. doi: 10.1042/CS20070347
- Zouhal H, Lemoine-Morel S, Mathieu M-E, et al (2013) Catecholamines and obesity: effects of exercise and training. *Sports Med* 43:591–600. doi: 10.1007/s40279-013-0039-8

ANNEXES

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Acute Hyperglycemia Impairs Vascular Function in Healthy and Cardiometabolic Diseased Subjects: Systematic Review and Meta-Analysis

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Acute Hyperglycemia Impairs Vascular Function in Healthy and Cardiometabolic Diseased Subjects

Systematic Review and Meta-Analysis

Jordan Loader, David Montero, Christian Lorenzen, Rani Watts, Cindy Méziat, Cyril Reboul, Simon Stewart, Guillaume Walther

Objectives—Controversy exists over the effect of acute hyperglycemia on vascular function. In this systematic review, we compared the effect of acute hyperglycemia on endothelial and vascular smooth muscle functions across healthy and cardiometabolic diseased subjects.

Approach and Results—A systematic search of MEDLINE, EMBASE, and Web of Science from inception until July 2014 identified articles evaluating endothelial or vascular smooth muscle function during acute hyperglycemia and normoglycemia. Meta-analyses compared the standardized mean difference (SMD) in endothelial and vascular smooth muscle functions between acute hyperglycemia and normoglycemia. Subgroup analyses and metaregression identified sources of heterogeneity. Thirty-nine articles (525 healthy and 540 cardiometabolic subjects) were analyzed. Endothelial function was decreased (39 studies; n=1065; SMD, -1.25; 95% confidence interval, -1.52 to -0.98; $P<0.01$), whereas vascular smooth muscle function was preserved (6 studies; n=144; SMD, -0.07; 95% confidence interval, -0.30 to 0.16; $P=0.55$) during acute hyperglycemia compared with normoglycemia. Significant heterogeneity was detected among endothelial function studies ($P<0.01$). A subgroup analysis revealed that endothelial function was decreased in the macrocirculation (30 studies; n=884; SMD, -1.40; 95% confidence interval, -1.68 to -1.12; $P<0.01$) but not in the microcirculation (9 studies; n=181; SMD, -0.63; 95% confidence interval, -1.36 to 0.11; $P=0.09$). Similar results were observed according to health status. Macrovascular endothelial function was inversely associated with age, blood pressure, and low-density lipoprotein cholesterol and was positively associated with the postocclusion interval of vascular assessment.

Conclusions—To our knowledge, this is the first systematic review and meta-analysis of its kind. In healthy and diseased subjects, we found evidence for macrovascular but not microvascular endothelial dysfunction during acute hyperglycemia. (*Arterioscler Thromb Vasc Biol*. 2015;35:2060-2072. DOI: 10.1161/ATVBAHA.115.305530.)

Key Words: cardiovascular diseases ■ hyperglycemia ■ meta-analysis ■ microcirculation ■ nitric oxide ■ vascular

The prevalence of type 2 diabetes mellitus represents a major public health issue, directly affecting an estimated 312 million people worldwide.¹ This burden is projected to worsen due, in part, to increasingly sedentary lifestyles and unhealthy dietary habits predominantly characterized by an excess consumption of added sugars.²⁻⁴ Habitual consumption of added sugars, most commonly in the form of sugar-sweetened beverages, is strongly associated with an increased risk in developing type 2 diabetes mellitus, as well as metabolic syndrome and obesity.⁵⁻⁸ In addition, consumption of added sugars has been linked to an increased risk of developing cardiovascular disease (CVD), which is the leading cause of mortality among those with cardiometabolic disease.^{1,9,10}

Consumption of excess added sugars leads to acute hyperglycemia, which is considered a better predictor of future CVD events than fasting glycemia in healthy and diabetic populations.^{11,12} Indeed, such acute hyperglycemic stress has also been proposed to contribute to vascular dysfunction,¹³ which represents one of the main precursors to CVD.¹⁴

In normal vascular function, the endothelium and vascular smooth muscle (VSM) cells continuously interact to regulate vasodilation and vasoconstriction, maintaining optimal organ perfusion and vascular tone.^{15,16} During acute hyperglycemia, increased oxidative stress has been proposed as a key trigger of vascular dysfunction by reducing nitric oxide (NO) production or NO bioavailability.^{15,17,18} Furthermore, animal and

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Nonstandard Abbreviations and Acronyms

CI	confidence interval
CVD	cardiovascular disease
NO	nitric oxide
SMD	standardized mean difference
VSM	vascular smooth muscle

in vitro studies suggest that acute hyperglycemia may also impair VSM function by disrupting VSM cell apoptosis, causing subsequent VSM cell proliferation and desensitization to NO.^{19–21} However, whether endothelial and VSM functions are transiently impaired during acute hyperglycemia in humans is unclear because of discrepant results. Given this, we conducted a systematic review and meta-analysis of available studies comparing endothelial function alone or in combination with VSM function during acute hyperglycemia in healthy and cardiometabolic diseased individuals. To our knowledge, this represents the first systematic review and meta-analysis to assess the effect of acute hyperglycemia on vascular function.

Materials and Methods

Materials and methods are available in the online-only Data Supplement.

Results

Study Selection and Characteristics

A flowchart of study selection is shown in Figure 1. The systematic search resulted in the inclusion of 39 from 394 potential articles.^{22–60} Fourteen of these articles reported vascular data for multiple subgroups of a given or diverse health status; thus, they were assessed as individual studies.^{22,25–28,31,37,38,45,52,56–59} The main characteristics and clinical data for these studies are shown in Tables 1 and 2, respectively. Three potentially relevant studies were not available for full text reading and thus could not be included.^{61–63} All studies assessed endothelial or VSM function during acute hyperglycemia and normoglycemia in a total of 1065 individuals classified as healthy ($n=525$), obese ($n=72$), impaired glucose tolerance ($n=104$), type 2 diabetes mellitus ($n=229$), hypertensive ($n=94$), metabolic syndrome ($n=30$), or type 1 diabetic mellitus ($n=11$).

Quality Assessment and Potential Bias

The quality of the studies was moderate-to-high. The mean score was 9.4 ± 1.5 of possible 12 points (Table 1). The quality of evidence for outcomes demonstrating the effect of acute hyperglycemia on vascular function was low-to-moderate (Table 3). As for the evaluation of potential bias, the funnel plot (Figure 2), Begg and Mazumdar rank correlation test, and the Egger regression test suggested the presence of publication bias or other biases for the standardized mean difference (SMD) in endothelial function in the studies included in the meta-analysis ($P<0.01$ and $P<0.01$, respectively). There was no evidence of publication or other biases when assessing the SMD in VSM function in the studies included in the meta-analysis.

Endothelial Function

After data pooling, endothelial function was significantly decreased during acute hyperglycemia compared with normoglycemia (39 studies; $n=1065$; SMD, -1.25 ; 95% confidence interval [CI], -1.52 to -0.98 ; $P<0.01$; Figure 3). There was no difference between health groups in the SMD in endothelial function ($P=0.13$), but significant heterogeneity was detected ($I^2=87\%$; $P<0.01$). Subgroup analysis of the SMD in endothelial function revealed that macrovascular function was significantly decreased during acute hyperglycemia compared with normoglycemia (30 studies; $n=884$; SMD, -1.40 ; 95% CI, -1.68 to -1.12 ; $P<0.01$), whereas no significant decrease was found in the studies that assessed microvascular endothelial function (9 studies; $n=181$; SMD, -0.63 ; 95% CI, -1.36 to 0.11 ; $P=0.09$). Heterogeneity was detected in the SMD in endothelial function for both macrovascular ($I^2=84\%$; $P<0.01$) and microvascular function studies ($P=90\%$; $P<0.01$). Of note, the heterogeneity about microvascular endothelial function was primarily explained ($\approx 40\%$) by a single study,³⁰ and the exclusion of such study did not significantly alter the pooled effect size (8 studies; $n=147$; SMD, -0.18 ; 95% CI, -0.53 to 0.17 ; $P=0.30$).

VSM Function

After data pooling, VSM function was preserved during acute hyperglycemia versus normoglycemia (6 studies; $n=144$; SMD, -0.07 ; 95% CI, -0.30 to 0.16 ; $P=0.55$; Figure 4). There was no significant difference between health groups in the SMD in VSM function ($P=0.49$), and no heterogeneity was observed ($I^2=0\%$; $P=0.85$). Because of limited data availability, it was not possible to analyze macro-VSM and micro-VSM function separately.

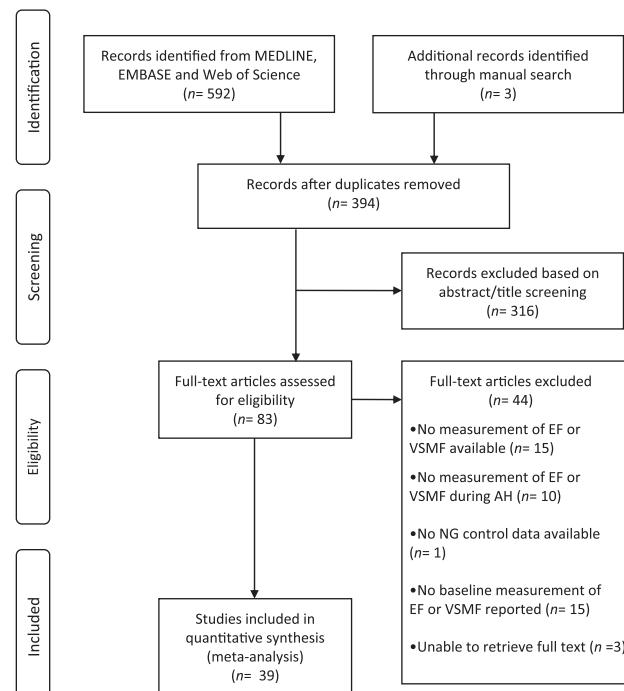


Figure 1. Flow diagram of the study selection process. AH indicates acute hyperglycemia; EF, endothelial function; NG, normoglycemia; and VSMF, vascular smooth muscle function.

Table 1. Main Characteristics of Studies Included in the Meta-Analysis

Study, Year of Publication	Study Design	Health Status	Medication	Quality Score (0–12)	Vascular Region	Inducing AH		Vascular EF During AH	VSMF During AH
						Method	Dose		
Grasser et al, ³⁴ 2014	RCT	Healthy	None	9	Micro	Energy drink	Sucrose/glucose, 39.1 g	↑ ACh	NA
Nakayama et al, ⁴³ 2013	OBS	Healthy	None	6	Macro	Sugar drink	Maltose, 75 g	↓ FMD	NA
Mah et al, ⁴² 2013	RCT	Healthy	None	11	Macro	OGTT	Glucose, 75 g	↓ FMD	NA
Wang et al A, ⁵² 2013	RCT	Healthy	None	9	Macro	OGTT	Glucose, 75 g	↓ FMD	NA
Zhang et al A, ⁵⁹ 2013	RCT	Healthy	None	10	Macro	OGTT	Glucose, 75 g	↓ FMD	NA
De Marchi et al, ³⁰ 2012	RCT	Healthy	None	8	Micro	OGTT	Glucose, 75 g	↓ ACh	↔ SNP
Grassi et al, ³⁵ 2012	RCT	Healthy	None	9	Macro	OGTT	Glucose, 75 g	↓ FMD	NA
Suzuki et al, ⁴⁹ 2012	OBS	Healthy	None	8	Macro	OGTT	Glucose, 75 g	↓ FMD	NA
Ceriello et al A, ²⁸ 2011	RCT	Healthy	None	6	Macro	OGTT	Glucose, 75 g	↓ FMD	NA
Mah et al, ⁴¹ 2011	RCT	Healthy	None	11	Macro	OGTT	Glucose, 75 g	↓ FMD	NA
Watanabe et al, ⁵³ 2011	OBS	Healthy	None	11	Macro	OGTT	Glucose 75 g	↓ FMD	NA
Baynard et al A, ²⁵ 2009	OBS	Healthy	3 statin, 1 hydrochlorothiazide, 1 angiotensin II receptor blocker, 1 ACE inhibitor	9	Macro	Test meal	Carbohydrate, 80 g	↓ FMD	NA
Ceriello et al (1) A, ²⁷ 2008	OBS	Healthy	None	8	Macro	IV infusion	Glucose, 15 mmol/L	↓ FMD	NA
Ceriello et al (2) A, ²⁶ 2008	RCT	Healthy	None	8	Macro	IV infusion	Glucose, 15 mmol/L	↓ FMD	NA
Natali et al A, ⁴⁵ 2008	OBS	Healthy	Unknown	11	Micro	OGTT	Glucose, 75 g	↔ ACh	↓ SNP
Weiss et al, ⁵⁴ 2008	RCT	Healthy	None	10	Macro	Candy/sugar drink	Carbohydrate, 101 g	↓ FMD	NA
Xiang et al (1) A, ⁵⁷ 2008	RCT	Healthy	None	11	Macro	OGTT	Glucose, 75 g	↔ FMD	NA
Xiang et al (2) A, ⁵⁶ 2008	OBS	Healthy	None	11	Macro	OGTT	Glucose, 75 g	↓ FMD	↔ NMD
Xiang et al (2) B, ⁵⁶ 2008	OBS	Healthy	None	11	Macro	OGTT	Glucose, 75 g	↔ FMD	↔ NMD
Dengel et al A, ³¹ 2007	OBS	Healthy	None	11	Macro	OGTT	Glucose, 75 g	↑ FMD	NA
Zhu et al, ⁶⁰ 2007	RCT	Healthy	None	7	Macro	OGTT	Glucose, 75 g	↓ FMD	NA
Arora et al A, ²² 2006	OBS	Healthy	None	10	Micro	OGTT	Glucose 75 g	↓ PORH	NA
Fujimoto et al, ³³ 2006	OBS	Healthy	None	10	Macro	OGTT	Glucose, 75 g	↓ FMD	NA
Tushuizen et al, ⁵¹ 2006	RCT	Healthy	None	11	Macro	Test meal	Carbohydrate, 55 g	↓ FMD	NA
Napoli et al, ⁴⁴ 2004	RCT	Healthy	None	6	Micro	Test meal	Carbohydrate, 60 g	↔ ACh	↔ SNP
Siafarikas et al, ⁴⁷ 2004	RCT	Healthy	None	9	Macro	OGTT	Glucose, 75 g	↔ FMD	NA
Ihlemann et al, ³⁶ 2003	RCT	Healthy	None	7	Micro	OGTT	Glucose, 75 g	↓ Serotonin	↓ SNP
Bagg et al, ²³ 2000	RCT	Healthy	None	9	Macro	IV infusion	Dextrose, 10% 238 mL	↔ FMD	NA
Title et al, ⁵⁰ 2000	RCT	Healthy	None	11	Macro	OGTT	Glucose, 75 g	↓ FMD	NA
Kawano et al A, ³⁸ 1999	OBS	Healthy	None	10	Macro	OGTT	Glucose, 75 g	↔ FMD	NA
Williams et al, ⁵⁵ 1998	OBS	Healthy	None	10	Micro	IV infusion	Glucose, 16.7 mmol/L	↔ Metacholine	NA
Lavi et al, ⁴⁰ 2009	RCT	Obese	None	8	Macro	Sugar drink	Glucose, 50 g	↓ FMD	NA
Dengel et al B, ³¹ 2007	OBS	Obese	None	11	Macro	OGTT	Glucose, 75 g	↑ FMD	NA
Wang et al B, ⁵² 2013	RCT	IGT	None	9	Macro	OGTT	Glucose, 75 g	↓ FMD	NA
Natali et al B, ⁴⁵ 2008	OBS	IGT	Unknown	11	Micro	OGTT	Glucose, 75 g	↔ ACh	↓ SNP
Xiang et al (1) B, ⁵⁷ 2008	RCT	IGT	None	11	Macro	OGTT	Glucose, 75 g	↓ FMD	NA
Arora et al B, ²² 2006	OBS	IGT	None	10	Micro	OGTT	Glucose, 75 g	↓ PORH	NA
Kawano et al B, ³⁸ 1999	OBS	IGT	None	10	Macro	OGTT	Glucose, 75 g	↓ FMD	NA
Wang et al C, ⁵² 2013	RCT	T2DM	None	9	Macro	OGTT	Glucose 75 g	↓ FMD	NA

(Continued)

Table 1. Continued

Study, Year of Publication	Study Design	Health Status	Medication	Quality Score (0–12)	Vascular Region	Inducing AH		Vascular EF During AH	VSMF During AH
						Method	Dose		
Ceriello et al B, ²⁸ 2011	RCT	T2DM	6 metformin discontinued 4 wk before 5 excluded ACE inhibitors	6	Macro	OGTT	Glucose 75 g	↓ FMD	NA
Chittari et al, ²⁹ 2011	OBS	T2DM	17 oral agents	10	Macro	OGTT	Glucose, 75 g	↓ FMD	NA
Kato et al A, ³⁷ 2010	RCT	T2DM	None	9	Macro	Cookie	Carbohydrate, 75 g	↓ FMD	NA
Kato et al B, ³⁷ 2010	RCT	T2DM	None	9	Macro	Cookie	Carbohydrate, 75 g	↓ FMD	NA
Kato et al C, ³⁷ 2010	RCT	T2DM	None	9	Macro	Cookie	Carbohydrate, 75 g	↓ FMD	NA
Ceriello et al (1) B, ²⁷ 2008	OBS	T2DM	None	8	Macro	IV infusion	Glucose 15, mmol/L	↓ FMD	NA
Ceriello et al (2) B, ²⁶ 2008	RCT	T2DM	None	8	Macro	IV infusion	Glucose 15, mmol/L	↓ FMD	NA
Ceriello et al (2) C, ²⁶ 2008	RCT	T2DM	None	8	Macro	IV infusion	Glucose 10, mmol/L	↓ FMD	NA
Ceriello et al (2) D, ²⁶ 2008	RCT	T2DM	None	8	Macro	IV infusion	Glucose 10, mmol/L	↓ FMD	NA
Natali et al C, ⁴⁵ 2008	OBS	T2DM	Unknown	11	Micro	OGTT	Glucose, 75 g	↔ ACh	↓ SNP
Stirban et al, ⁴⁸ 2006	RCT	T2DM	13 insulin, 11 aspirin, 9 ACE inhibitors, 1 angiotensin receptor blocker, 6 hydroxymethylglutaryl-CoA inhibitors, 5 β-blockers, 5 diuretics, 3 calcium channel blockers	11	Macro	Test meal	Carbohydrate, 48 g	↓ FMD	NA
Kim et al, ³⁹ 2003	OBS	T2DM	None	9	Micro	IV infusion	Glucose, 12/mg/kg per min	↓ PORH	NA
Kawano et al C, ³⁸ 1999	OBS	T2DM	None	10	Macro	OGTT	Glucose, 75 g	↓ FMD	NA
Shige et al, ⁴⁶ 1999	OBS	T2DM	None	8	Macro	Test meal	Sucrose, 75 g	↓ FMD	↔ NMD
Zhang et al B, ⁵⁹ 2013	RCT	Hypertensive	None	10	Macro	OGTT	Glucose, 75 g	↓ FMD	NA
Zhang et al A, ³⁸ 2012	RCT	Hypertensive	None	11	Macro	OGTT	Glucose, 75 g	↓ FMD	NA
Zhang et al B, ⁵⁸ 2012	RCT	Hypertensive	None	11	Macro	OGTT	Glucose, 75 g	↓ FMD	NA
Ballard et al, ²⁴ 2013	RCT	MetS	None	11	Macro	Rice milk	Mixed sugars, 23 g	↓ FMD	NA
Baynard et al B, ²⁵ 2009	OBS	MetS	3 metformin, 2 sulfonylurea, 4 statin	11	Macro	Test meal	Carbohydrate, 80 g	↓ FMD	NA
Dye et al, ³² 2012	OBS	T1DM	Insulin monotherapy	9	Micro	IV infusion	Dextrose, 200 mg/dL	↓ PORH	NA

Some studies presented multiple health groups comparing normoglycemia and acute hyperglycemia vascular function and were therefore evaluated as individual studies (distinguished by A, B, C, or D). Authors who published multiple studies in a single year were distinguished by (1) or (2). ↓ indicates significant decrease of vascular function during acute hyperglycemia compared with normoglycemia; ↔, no significant difference in vascular function between acute hyperglycemia and normoglycemia; ↑, significant increase of vascular function during acute hyperglycemia compared with normoglycemia; ACE, angiotensin-converting enzyme; ACh, acetylcholine; AH, acute hyperglycemia; EF, endothelial function; FMD, flow-mediated dilation; IGT, impaired glucose tolerance; IV, intravenous; MetS, metabolic syndrome; NA, vascular data not available; NMD, nitroglycerin-mediated dilation; OBS, observational study; OGTT, oral glucose tolerance test; PORH, postocclusive reactive hyperemia; RCT, randomized controlled trial; SNP, sodium nitroprusside; T1DM, type 1 diabetic mellitus; T2DM, type 2 diabetic mellitus; and VSMF, vascular smooth muscle function.

Metaregression Analyses

The SMD in macrovascular endothelial function was inversely associated with age ($\beta=-0.03$; $P<0.01$), systolic blood pressure ($\beta=-0.04$; $P<0.01$), diastolic blood pressure ($\beta=-0.05$; $P<0.01$), mean arterial pressure

($\beta=-0.05$; $P<0.01$), and low-density lipoprotein cholesterol ($\beta=-0.93$; $P<0.01$; Figure 5). In turn, the SMD in macrovascular endothelial function was positively associated with the postocclusion interval of vascular assessment ($\beta=0.36$; $P=0.01$).

Table 2. Clinical Data for Each Study Included in This Meta-Analysis

Study, Year of Publication	Health Status	n (%) Women	Age, y	BMI, kg/m ²	Blood Pressure, mm Hg			Fasting Plasma Insulin, pmol/L	Fasting Plasma Glucose, mmol/L	Cholesterol, mmol/L			Triglycerides, mmol/L	
					SBP	DBP	MAP			HbA _{1c} , %	Total	HDL	LDL	
Grasser et al, ³⁴ 2014	Healthy	25 (48)	22.5±3	23.3±10	114±10	87±5	96±7	NA	NA	NA	NA	NA	NA	NA
Nakayama et al, ⁴³ 2013	Healthy	23 (0)	44±10	23±2.1	111±8	71±6	84±7	NA	5.1±0.4	NA	NA	NA	NA	NA
Mah et al, ⁴² 2013	Healthy	16 (0)	21.8±3.2	24.8±NA	117±4	79±4	92±4	142.5±94.4	5.3±0.4	NA	3.7±0.8	NA	NA	3.3±5
Wang et al A, ⁵² 2013	Healthy	33 (64)	51.36±7.15	24.76±3.6	NA	NA	NA	NA	5.4±0.5	6±0.3	5.5±0.6	1.4±0.4	3.7±0.6	1.5±0.5
Zhang et al A, ⁵⁹ 2013	Healthy	31 (48)	47.87±10.95	23.9±2.1	124±8	79±5	94±6	NA	4.8±1.1	NA	4.7±0.4	1.4±0.3	2.9±0.4	1.3±0.3
De Marchi et al, ³⁰ 2012	Healthy	34 (50)	32.4±3.5	19±3	119±4	79±9	92±7	34.8±6.6	5.1±0.6	NA	4.3±0.7	1.5±0.2	NA	1.5±0.1
Grassi et al, ³⁵ 2012	Healthy	12 (58)	28±2.7	23.2±4.2	NA	NA	NA	NA	4.2±0.4	NA	NA	1.5±0.4	2.5±0.4	0.7±0.3
Suzuki et al, ⁴⁹ 2012	Healthy	14 (57)	33.4±11.9	20.7±2.3	106±9	64±6	78±7	NA	4.8±0.6	5.4±0.3	5±0.6	1.7±0.4	2.7±0.5	0.8±0.4
Ceriello et al A, ²⁸ 2011	Healthy	12 (50)	50.5±8.66	28.5±10.7	117±19	78±8	91±11	73.4±15.2	4.5±1.4	4.8±0.7	4.5±2.1	1.4±0.7	2.5±1	0.9±0.7
Mah et al, ⁴¹ 2011	Healthy	16 (0)	21.6±3.2	28.7±NA	117±4	79±4	92±4	147±96	5.3±0.4	NA	3.6±0.7	NA	NA	NA
Watanabe et al, ⁵³ 2011	Healthy	14 (43)	33.4±11.9	20.7±2.3	106±9	64±6	78±7	29.7±11.9	4.7±0.4	NA	5±0.6	1.7±0.4	2.7±0.5	0.8±0.4
Baynard et al A, ²⁵ 2009	Healthy	10 (NA)	53±3.32	32.7±3.5	117±13	72±3	87±6	NA	4.6±0.3	5.2±0.4	5.2±1	1.3±0.3	3.2±0.95	1±0.3
Ceriello et al (1) A, ²⁷ 2008	Healthy	22 (45)	50.5±11.73	28.5±14.5	117±26	78±10	91±15	NA	4.5±1.4	4.8±0.9	4.5±2.8	1.4±0.9	2.5±1.4	0.9±0.9
Ceriello et al (2) A, ²⁶ 2008	Healthy	10 (40)	50.3±8.22	27.5±9.8	115±14	76±11	89±12	NA	4.5±1	4.8±0.6	4.8±1.9	1.4±1.6	2.4±1.3	0.9±1.6
Natali et al A, ⁴⁵ 2008	Healthy	20 (70)	49±8.94	27.9±4	129±13	78±9	95±10	NA	5.3±NA	5.6±0.5	4.9±0.9	1.2±0.2	3.3±0.8	1.1±0.5
Weiss et al, ⁵⁴ 2008	Healthy	13 (62)	48±17	24±2.2	114±14	66±7	82±10	NA	5.5±1.4	NA	NA	NA	NA	NA
Xiang et al (1) A, ⁵⁷ 2008	Healthy	26 (46)	50±6	24.2±2.3	114±7	72±6	86±7	NA	4.6±0.5	NA	4.4±0.9	1.2±0.1	1.9±0.7	1.4±1.2
Xiang et al (2) A, ⁵⁶ 2008	Healthy	17 (41)	39±12.37	24.1±6.6	115±20	68±12	84±15	NA	5.1±1.7	5.1±0.4	4.7±2.7	1.2±0.7	1.9±2.4	1.4±3.8
Xiang et al (2) B, ⁵⁶ 2008	Healthy	15 (47)	40±11.62	23.7±7.4	111±20	24±7	53±12	NA	4.6±1.9	4.8±0.8	4.5±1.9	1.2±0.7	1.9±1.8	1.4±3
Dengel et al A, ³¹ 2007	Healthy	15 (53)	11.3±1.55	17.5±1.9	110±12	59±8	76±9	41.2±15.1	4.8±0.3	NA	3.8±0.8	1.2±0.3	2.3±0.6	0.8±0.4
Zhu et al, ⁶⁰ 2007	Healthy	11 (0)	22.6±2.3	22.5±1.5	113±7	79±6	90±6	NA	5.2±0.2	NA	4±0.8	1.4±0.2	2.1±0.6	0.9±0.3
Arora et al A, ²² 2006	Healthy	10 (0)	27±NA	22.4±NA	122±NA	68±NA	86±NA	NA	4.8±NA	NA	4.1±NA	1.8±NA	2.2±NA	1.1±NA
Fujimoto et al, ³³ 2006	Healthy	10 (0)	30±2	NA	111±12	65±8	80±9	NA	5.1±0.6	NA	4.3±0.7	1.2±0.3	NA	1.2±0.7
Tushuizen et al, ⁵¹ 2006	Healthy	17 (0)	25.4±3	23.6±1.8	116±8	75±7	89±7	33±10	4.8±0.3	5.1±0.2	4±0.6	1.4±0.2	2.2±0.6	0.8±0.3
Napoli et al, ⁴⁴ 2004	Healthy	10 (40)	23±3.16	23.6±1.9	124±6	60±3	81±4	NA	5±NA	NA	NA	NA	NA	NA
Siafarikas et al, ⁴⁷ 2004	Healthy	32 (66)	19.1±1.7	22.9±4.2	NA	NA	NA	NA	NA	5±0.3	4±0.8	1.3±0.4	2.2±0.7	1.2±0.7
Ihlemann et al, ³⁶ 2003	Healthy	10 (40)	53±6.96	22.7±1.9	144±17	75±11	98±13	NA	NA	5.2±0.3	5.2±0.6	1.7±0.3	3.2±0.6	0.8±0.3
Bagg et al, ²³ 2000	Healthy	10 (20)	26±6	22±2	111±10	65±8	80±9	34.2±11.4	5.2±0.3	NA	4.7±1	NA	NA	NA
Title et al, ⁵⁰ 2000	Healthy	10 (40)	25.5±3.1	24±3	118±8	72±7	87±7	NA	5.3±0.7	NA	5.1±1.1	1.3±0.1	3.3±0.9	1.2±0.5
Kawano et al A, ³⁸ 1999	Healthy	17 (35)	52.6±7.42	NA	NA	NA	NA	51.6±9.9	5±0.3	NA	4.9±0.4	1.2±0.1	3±0.4	1.5±0.2

(Continued)

Table 2. Continued

Study, Year of Publication	Health Status	n (%) Women	Age, y	BMI, kg/m ²	Blood Pressure, mm Hg			Fasting Plasma Insulin, pmol/L	Fasting Plasma Glucose, mmol/L	Cholesterol, mmol/L			Triglycerides, mmol/L	
					SBP	DBP	MAP			HbA _{1c} , %	Total	HDL	LDL	
Williams et al, ⁵⁵ 1998	Healthy	10 (30)	33±6.32	NA	NA	NA	NA	NA	3.9±1.2	3.6±0.6	4.2±0.7	1.1±0.2	2.6±0.7	1±0.4
Lavi et al, ⁴⁰ 2009	Obese	56 (0)	47.9±5.8	32.1±4.3	134±13	82±6	99±8	NA	5.4±0.2	NA	5.1±0.7	1.1±0.2	3.2±0.7	1.7±0.9
Dengel et al B, ³¹ 2007	Obese	16 (56)	10.1±1.6	19.3±6.4	120±12	65±8	83±9	61.6±33.2	4.8±0.2	NA	4.2±0.6	1.1±0.2	2.6±0.5	1±0.6
Wang et al B, ⁵² 2013	IGT	33 (64)	52.88±9.2	27.8±3.1	NA	NA	NA	NA	6.1±0.5	6.5±0	5.5±1	1.2±0.3	3.1±0.8	1.8±0.9
Natali et al B, ⁴⁵ 2008	IGT	16 (63)	52±20	29.5±4.8	122±12	79±8	93±9	NA	5.7±NA	5.9±1.2	5.6±1.4	1.3±0.5	3.7±1.3	1.3±0.6
Xiang et al (1) B, ⁵⁷ 2008	IGT	21 (48)	51±6	24.8±3.1	110±8	72±6	85±7	NA	5.9±0.9	NA	5.2±1.1	1.2±0.2	2.3±0.7	2±1.1
Arora et al B, ²² 2006	IGT	10 (0)	65±NA	23.2±NA	134±NA	72±NA	93±NA	NA	5.3±NA	NA	4.3±NA	1.2±NA	2.1±NA	2.6±NA
Kawano et al B, ³⁸ 1999	IGT	24 (38)	58.5±7.84	NA	NA	NA	NA	66±11.8	5.8±0.8	NA	5.3±1	1.1±0.1	3.4±0.5	1.7±0.2
Wang et al C, ⁵² 2013	T2DM	43 (42)	53.4±8.99	25.7±2.9	NA	NA	NA	NA	7.5±1.2	7.4±0.1	5.8±1.7	1.2±0.3	3.2±0.7	2.2±1.2
Ceriello et al B, ²⁸ 2011	T2DM	16 (44)	51.3±10.4	29.5±13.2	123±26	80±14	95±18	107.3±15.2	7.8±8.8	8.4±1.2	5.1±3.2	1.2±0.3	2.6±0.4	1.2±1.6
Chittari et al, ²⁹ 2011	T2DM	21 (43)	46.4±9.62	30.1±5	NA	76±8	NA	NA	7.8±1.8	7.8±1.4	4.5±0.9	1.2±0.5	2.5±0.9	1.6±0.5
Kato et al A, ³⁷ 2010	T2DM	10 (50)	68±7.7	26.8±3.2	144±29	89±19	107±22	45±17.4	6.1±0.8	5.8±0.6	5.5±0.4	1.4±0.3	3.5±1.3	1.5±0.1
Kato et al B, ³⁷ 2010	T2DM	10 (30)	67.6±6.2	25.8±2.5	141±26	88±23	106±24	57.6±33.6	6.1±1	6±0.3	5.2±0.7	1.5±0.3	3.7±1.8	1.3±0.2
Kato et al C, ³⁷ 2010	T2DM	10 (30)	67.8±8.6	25.8±3.3	141±29	88±12	106±17	39.6±24.6	6.6±1	6.1±0.6	5.3±0.7	1.4±0.3	3.9±2	1.4±0.2
Ceriello et al (1) B, ²⁷ 2008	T2DM	27 (48)	51.3±13.51	29.5±17.2	123±33	80±19	95±24	NA	7.8±11.4	7.7±1.6	5.1±4.2	1.2±0.3	2.6±0.4	1.2±1.6
Ceriello et al (2) B, ²⁶ 2008	T2DM	10 (50)	50.3±6.96	27.5±10.1	118±17	77±12	91±14	NA	6.8±7	7.3±1	5±2.5	1.3±0.2	2±0.5	1±1.9
Ceriello et al (2) C, ²⁶ 2008	T2DM	10 (50)	50.2±14.23	28.4±13	121±7	79±9	93±8	NA	7.7±1.3	7.9±1.6	5±2.2	1.2±0.3	2.7±0.4	1.3±0.95
Ceriello et al (2) D, ²⁶ 2008	T2DM	10 (60)	51±17.71	28.6±11.7	122±11	80±12	94±12	NA	6±1	7.7±1.9	5.1±2.9	1.2±0.3	2.6±0.8	1.2±2.2
Natali et al C, ⁴⁵ 2008	T2DM	17 (71)	58±8.25	29.3±4.1	139±17	81±8	100±11	NA	8.3±NA	7.2±2.1	4.9±1.2	1±0.3	3.29±1.1	1.5±0.4
Stirban et al, ⁴⁸ 2006	T2DM	13 (NA)	56.9±10.1	30.3±3.2	136±22	79±13	98±16	NA	NA	8.5±1.8	NA	NA	NA	NA
Kim et al, ³⁹ 2003	T2DM	8 (50)	55.9±3.7	25.4±3.2	134±7	82±6	99±7	NA	7.2±2.6	6.6±0.8	5.5±0.3	1.3±0.2	NA	2.1±0.6
Kawano et al C, ³⁸ 1999	T2DM	17 (29)	62.2±4.95	NA	NA	NA	NA	84±22.3	7.1±0.3	NA	5.6±0.4	1.1±0.1	3.7±0.4	1.9±0.2
Shige et al, ⁴⁶ 1999	T2DM	7 (29)	49.3±8	26±5	128±9	72±9	91±9	47.4±16.8	7.1±1.3	NA	4.9±13	1±0.2	NA	1.6±0.6
Zhang et al B, ⁵⁹ 2013	Hypertensive	34 (50)	47.44±10.96	24.7±3.5	156±9	96±8	116±8	NA	4.7±0.6	NA	4.7±0.5	1.3±0.2	3±0.4	1.3±0.3
Zhang et al A, ⁵⁸ 2012	Hypertensive	26 (50)	47.92±8.05	24.6±2.6	160±8	97±6	118±7	NA	5.3±0.5	NA	4.7±0.6	1.4±0.3	2.6±0.4	1.4±0.2
Zhang et al B, ⁵⁸ 2012	Hypertensive	34 (47)	49.29±8	24.8±2.5	159±9	99±6	119±7	NA	5.2±0.6	NA	4.9±0.5	1.3±0.3	2.7±0.5	1.3±0.3
Ballard et al, ²⁴ 2013	MetS	19 (26)	28.5±9.59	35±3.9	125±11	85±8	98±8	55±25.7	6±0.9	NA	4.7±0.9	1±0.1	2.8±0.2	1.9±0.9
Baynard et al B, ²⁵ 2009	MetS	11 (NA)	52±3.16	34.4±5	123±7	78±3	93±4	NA	6.1±1.3	5.9±1.1	5.1±1	1±NA	3.1±0.7	2.4±1.3
Dye et al, ³² 2012	T1DM	11 (36)	14.5±3.32	21.5±9.6	105±7	61±10	76±9	NA	NA	8.3±1.2	NA	NA	NA	NA

Data are expressed as mean±SD or n. Some studies presented multiple health groups comparing normoglycemia and acute hyperglycemia vascular function and were therefore evaluated as individual studies (distinguished by A, B, C, or D). Authors who published multiple studies in a single year were distinguished by (1) or (2). BMI indicates body mass index; DBP, diastolic blood pressure; HDL, high-density lipoprotein; IGT, impaired glucose tolerance; LDL, low-density lipoprotein; MAP, mean arterial pressure; MetS, metabolic syndrome; NA, data not available; T1DM, type 1 diabetic mellitus; T2DM, type 2 diabetic mellitus; and SBP, systolic blood pressure.

Table 3. Effect of Acute Hyperglycemia on Vascular Function in Healthy and Cardiometabolic Populations: Quality of Evidence

Outcome Among Participants	Design (No. of Studies)	Risk of Bias	Inconsistency	Indirectness	Imprecision	Other Considerations*	Quality of Evidence (GRADE)
Decreased EF	RCT (23)	Serious†	Not serious	Not serious	Not serious	Publication bias likely¶	Low
Decreased EF	OBS (16)	Not serious	Not serious	Not serious	Not serious	None	Low
Decreased macro-EF	RCT (19)	Serious†	Not serious	Not serious	Not serious	Publication bias likely¶	Low
Decreased macro-EF	OBS (11)	Not serious	Not serious	Not serious	Not serious	None	Low
Preserved micro-EF	RCT (4)	Very serious†	Serious§	Not serious	Very serious	Publication bias likely¶	Very low
Preserved micro-EF	OBS (5)	Serious‡	Not serious	Not serious	Serious	None	Very low
Preserved VSMF	RCT (3)	Serious†	Not serious	Not serious	Not serious	None	Moderate
Preserved VSMF	OBS (3)	Not serious	Not serious	Not serious	Not serious	None	Low

EF indicates endothelial function; GRADE, Grading of Recommendations Assessment, Development and Evaluation; OBS, observational study; RCT, randomized controlled trial; and VSMF, vascular smooth muscle function.

*Large magnitude of effect, dose-response, plausible biases decreasing the magnitude of effect, publication bias.

†Method for allocation of concealment and participant/assessor blinding is unclear or not performed. Incomplete outcome data and selective reporting when assessing endothelial, macrovascular endothelial, microvascular endothelial, and vascular smooth muscle function during acute hyperglycemia.

‡No control population included, failure to adequately control for confounding, and incomplete follow-up when assessing microvascular endothelial function.

§Large I^2 and point estimates vary widely across studies assessing microvascular endothelial function suggesting benefit, harm, and no effect of acute hyperglycemia.

||The 95% confidence interval of the pooled risk ratio includes both positive and negative effects of acute hyperglycemia.

¶Publication bias is strongly suspected because of the presence of asymmetry in funnel plots for randomized control trials assessing endothelial, macrovascular endothelial, and microvascular endothelial function during hyperglycemia.

Discussion

To our knowledge, this is the first systematic review and meta-analysis to assess the effect of acute hyperglycemia on vascular function. Data from 39 studies assessing endothelial function alone or in combination with VSM function during acute hyperglycemia in healthy and cardiometabolic diseased individuals were pooled and analyzed. The meta-analysis provided evidence that the average effect of acute hyperglycemia on endothelial function is decreased function, whereas VSM function was preserved in healthy and diseased individuals. Because of evidence of heterogeneity, the interpretation of the pooled effects of hyperglycemia on endothelial function should be made cautiously. Considerable heterogeneity was identified for most subgroups suggesting that the variability across studies was because of not only sampling variability but also differences in treatment effect within each study.⁶⁴ Nevertheless, the large effect sizes and 95% CIs consistently

favored normoglycemia, providing evidence of treatment effect.⁶⁴ Exploration of heterogeneity with metaregression indicated that the variability across studies could be explained by differences in age, blood pressure, and low-density lipoprotein cholesterol and the postocclusion interval of vascular assessment.

Currently, there is no consensus on the effect of acute hyperglycemia on endothelial function and VSM function as studies assessing vascular function during acute hyperglycemia have presented confounding results. This meta-analysis demonstrated evidence of macrovascular endothelial dysfunction during acute hyperglycemia in healthy people, as well as cardiometabolic diseased subjects, suggesting that the pathogenesis of CVD may begin, among others, with acute hyperglycemia-mediated transient decreases in endothelial function long before the onset of morbidities, such as obesity, hypertension, or type 2 diabetes mellitus. Interestingly, the inverse

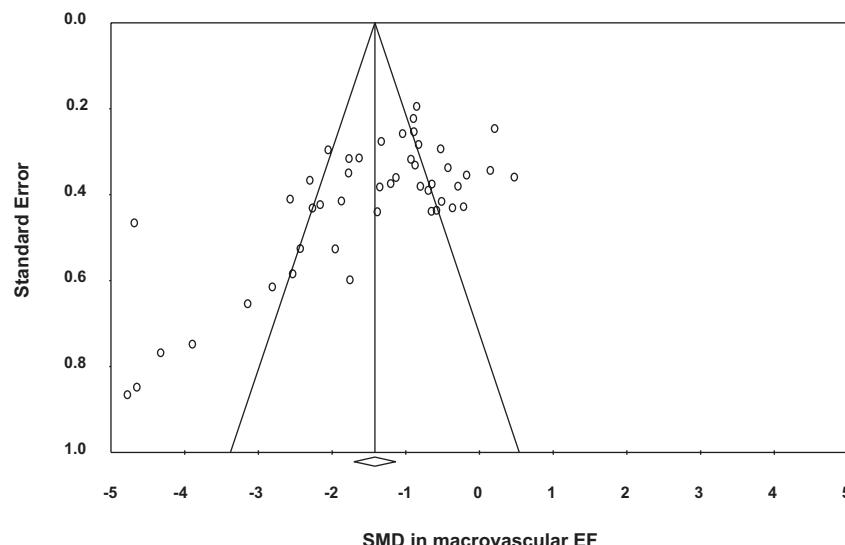


Figure 2. Funnel plot of the standardized mean difference (SMD) in macrovascular endothelial function in studies included in the meta-analysis. Funnel plot asymmetry: $P=0.0002$ and $P=0.00005$ according to Begg and Mazumdar rank correlation test and Egger test, respectively. EF indicates endothelial function.

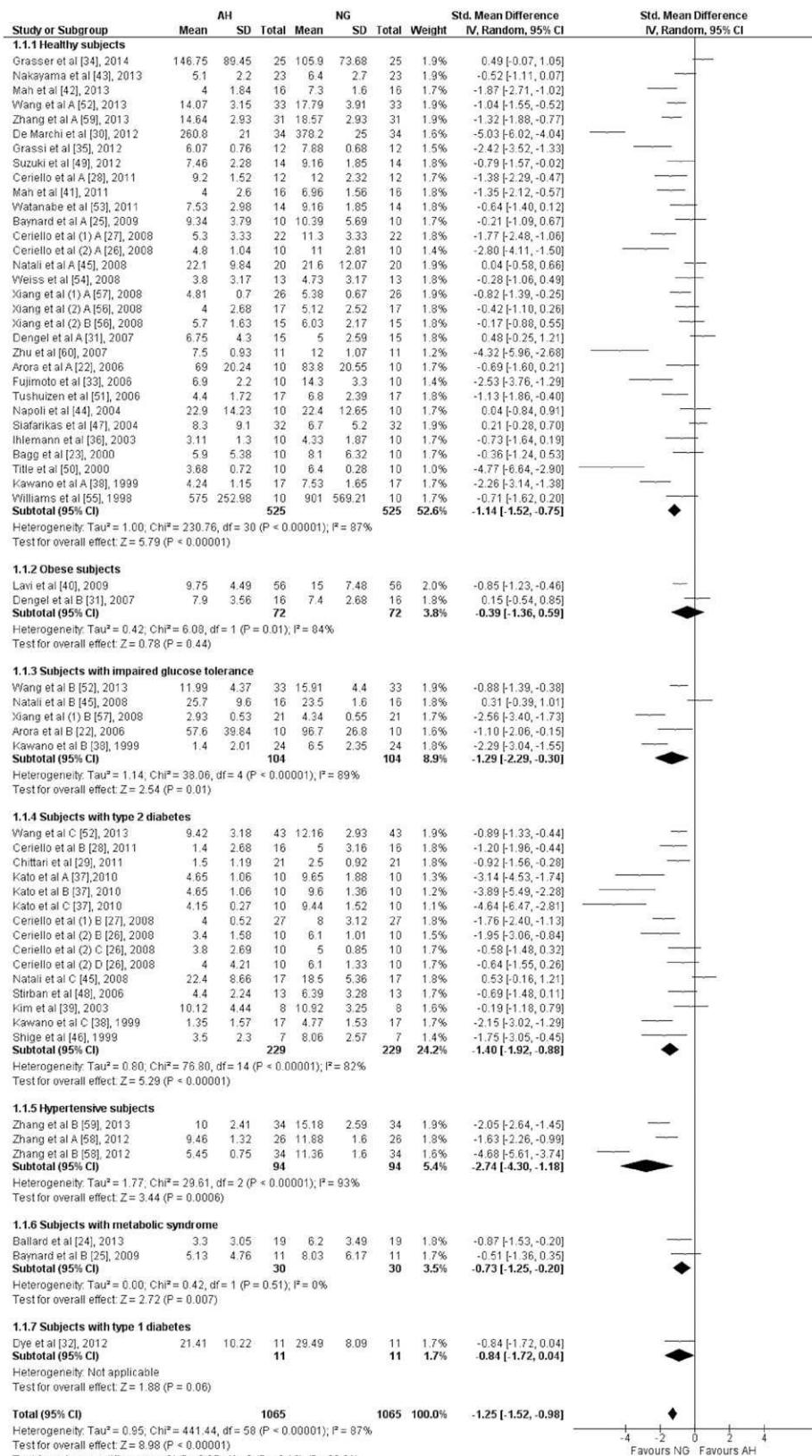


Figure 3. Forest plot of standardized mean difference (SMD) in endothelial function between acute hyperglycemic and normoglycemic states for all health groups included. Squares represent the SMD in endothelial function of each study. The diamond represents the pooled SMD in endothelial function by health group and overall. Some studies presented multiple subgroups according to health status; thus, they were evaluated as individual studies (distinguished by A, B, C, or D). Authors who published multiple studies in a single year had studies distinguished by numeric values (1 and 2). AH indicates acute hyperglycemia; CI, confidence interval; IV, inverse variance; and NG, normoglycemia.

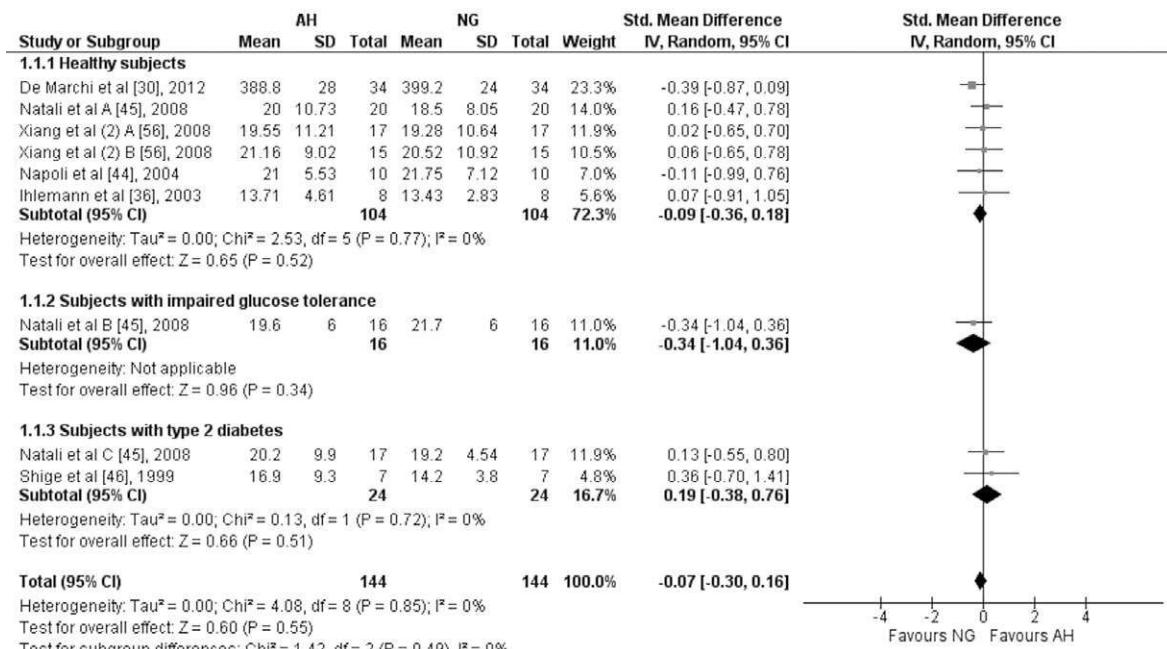


Figure 4. Forest plot of standardized mean difference (SMD) in vascular smooth muscle (VSM) function between acute hyperglycemic and normoglycemic states for all health groups included. Squares represent the SMD in VSM function of each study. The diamond represents the pooled SMD in VSM function by health group and overall. Some studies presented multiple subgroups according to health status; thus, they were evaluated as individual studies (distinguished by A, B, or C). Authors who published multiple studies in a single year had studies distinguished by numeric values (1 and 2). AH indicates acute hyperglycemia; CI, confidence interval; IV, inverse variance; and NG, normoglycemia.

relationship between macrovascular endothelial function and several traditional cardiovascular risk factors demonstrates the degree to which acute hyperglycemia mediates macrovascular endothelial dysfunction and correlates with increases in age, blood pressure, or low-density lipoprotein cholesterol levels. This is consistent with previous research revealing that elderly, hypertensive and subjects with dyslipidemia all exhibited significantly decreased endothelium-dependent vasodilation at rest compared with healthy populations,^{65–67} indicating that any existing macrovascular endothelial dysfunction may therefore be compounded by an acute hyperglycemic stress. The fact that microvascular endothelial dysfunction during acute hyperglycemia was not detected contradicts previously published data that associates decreased microvascular function with incident type 2 diabetes mellitus and suggests a role for microvascular dysfunction in the pathogenesis of type 2 diabetes mellitus.⁶⁸ Although in this meta-analysis, macrovascular endothelial function was affected across healthy and cardiometabolic health groups, VSM function remained preserved during acute hyperglycemia when compared with normoglycemia. In contradiction to previous findings in animal and *in vitro* studies, which found VSM dysfunction mediated by VSM cell proliferation may occur in as little as 6 hours.^{20,52}

Macrovascular endothelial dysfunction observed during acute hyperglycemia by methods assessing endothelium-dependent vasodilation primarily implicate decreased NO bioavailability as a central mechanism of endothelial dysfunction in healthy and cardiometabolic diseased populations.⁶⁹ This may be attributed to acute hyperglycemia increasing oxidative stress and its role in disrupting pathways of NO synthesis.¹² The fact that even healthy people

exhibited decreased macrovascular endothelial function during acute hyperglycemia demonstrates how acutely NO bioavailability may be affected by excess sugar consumption. The magnitude of macrovascular endothelial dysfunction induced by acute hyperglycemia may be compounded when cardiovascular risk factors, such as increased age, blood pressure, or low-density lipoprotein cholesterol, are present. This may be partly due to the fact that health groups exhibiting these clinical markers demonstrate decreased NO bioavailability and therefore impaired endothelial function, even at rest.^{65–67} Although NO is the predominant vasodilator in macrocirculation, it has been demonstrated to have significantly less influence in the microcirculatory system.⁷⁰ The increased influence of other chemical mediators of vasodilation, such as endothelial-derived hyperpolarizing factor and prostaglandin I₂,⁷¹ may, however, explain why microvascular endothelial function remained preserved during acute hyperglycemia. Consideration should also be given to the spatial variability associated with the techniques used in several of the studies using skin microcirculation as a model of assessing microvascular function.⁷² The large spatial variability in single-point laser Doppler flowmetry, for example, may have limited findings.⁷³ Despite this, final conclusions on the effect of acute hyperglycemia on microvascular function should not be made because of the limited availability of microcirculation data. Furthermore, it must be acknowledged that shear stress was not considered when performing analyses of flow-mediated dilation data in many studies. Shear stress, which is responsible for inducing the NO release that causes flow-mediated dilation, is dependent on variability of the hyperemic blood flow response in the microcirculation.⁷⁴

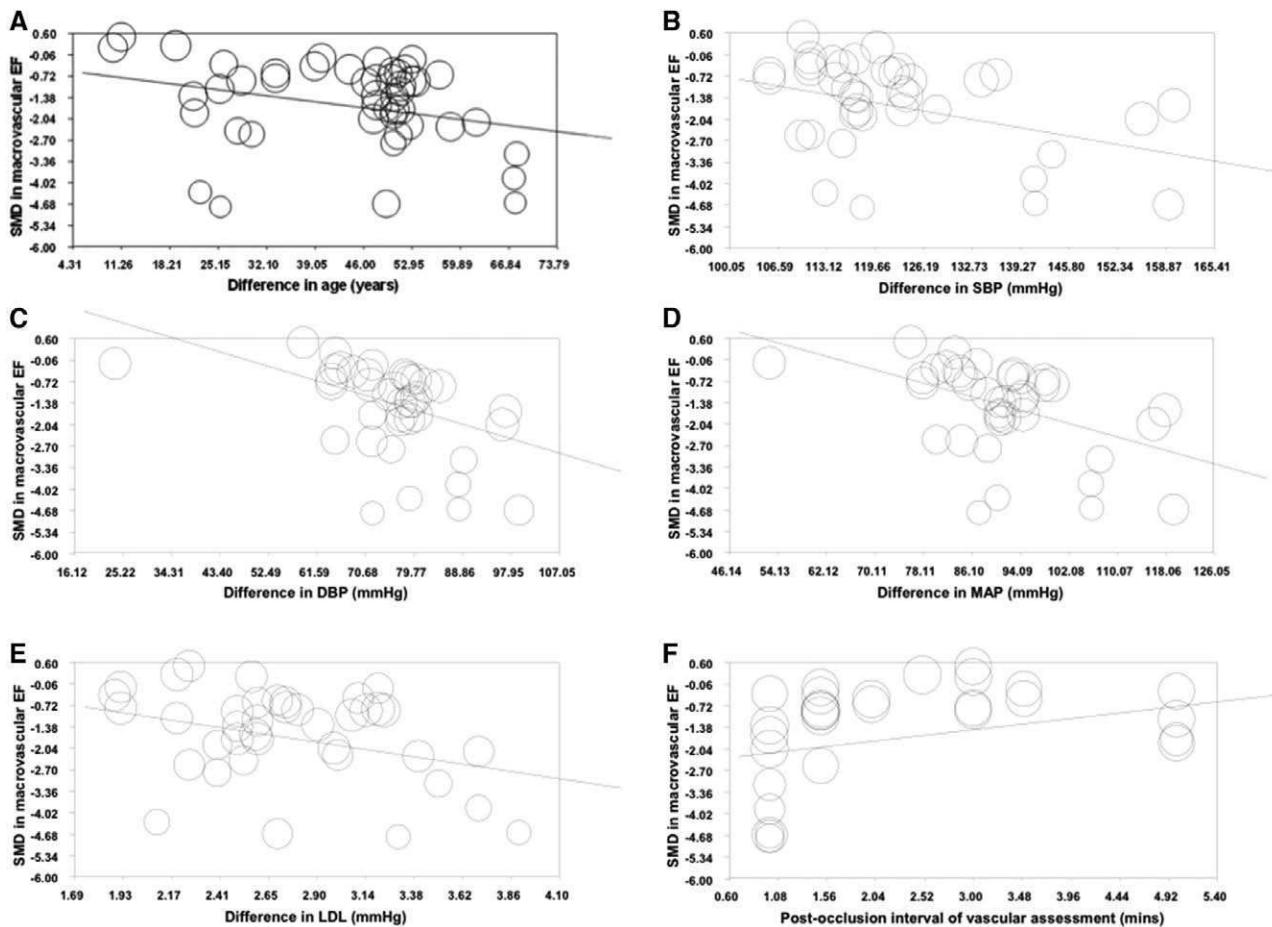


Figure 5. Metaregression plots of standardized mean difference (SMD) in macrovascular endothelial function (EF) according to the difference in (A) age ($\beta=-0.03$; $P=0.005$), (B) systolic blood pressure (SBP; $\beta=-0.04$; $P=0.0004$), (C) diastolic blood pressure (DBP; $\beta=-0.05$; $P<0.00001$), (D) mean arterial pressure (MAP; $\beta=-0.05$; $P=0.00001$), (E) low-density lipoprotein cholesterol (LDL; $\beta=-0.93$; $P=0.005$), and (F) postocclusion interval of vascular assessment ($\beta=0.36$; $P=0.01$). The size of each circle is proportional to the study's weight.

Therefore, macrovascular endothelial dysfunction observed during acute hyperglycemia may partially be mediated by a decrease in shear stress stimulus reflecting microvascular dysfunction.⁷⁴ The fact that VSM function was preserved during acute hyperglycemia indicates that endothelial dysfunction precedes VSM dysfunction, further supporting its role as a primary mechanism of CVD pathogenesis. Although disruptions in NO bioavailability mediated by acute hyperglycemia and the resulting endothelial dysfunction may initially be transient, if repeated often enough, they may lead to cumulative adverse outcomes, including proinflammatory responses and VSM cell proliferation.^{12,75} It has been suggested that acute hyperglycemia may induce VSM cell proliferation by disrupting VSM cell apoptosis, which is a key mechanism to prevent increased neointimal formation and stenosis.¹⁹ Moreover, decreased NO delivery by the endothelium to the VSM may contribute to VSM cell proliferation through increased periods of higher vasoconstrictive tone.⁷⁵ Ultimately, VSM cell proliferation may signal the beginning of a detectable and significant VSM dysfunction, representing a critical event in vascular remodeling and the development of CVD.⁷⁶

Given that CVD is the single leading cause of death, accounting for 30% of the annual global mortality rate,⁷⁷ and

that even healthy populations are subject to vascular dysfunction during acute hyperglycemia, there is a clear need to further investigate the effects of acute hyperglycemia on the underlying mechanisms of vascular function *in vivo*. Because of a surge in added sugar consumption in recent decades, predominantly in the form of sugar-sweetened beverages,² humans are more often in a state of acute hyperglycemia and therefore are more frequently inducing endothelial dysfunction. Considering this, future research should quantify what frequency and dosage of sugar consumption mediate atherosclerotic vascular changes in healthy and cardiometabolic diseased populations. Previously, certain ethnicities have demonstrated decreased vascular function at rest compared with white subjects.⁷⁸ Whether this exacerbates any vascular dysfunction mediated by acute hyperglycemia is still unknown and requires further research. To provide more comprehensive conclusions on how macrovascular and microvascular functions are affected by acute hyperglycemia, future research should consider shear stress as a covariate of conduit artery flow-mediated dilation data during statistical analyses.^{79,80} Furthermore, noting that vascular function is not entirely mediated by NO,¹² future research may also investigate the effect of sugar-sweetened beverage consumption on numerous mechanisms of vasodilation (eg, NO, endothelium-derived

hyperpolarizing factor, and prostaglandin I₂) and vasoconstriction (eg, endothelin-1), the influence of which varies from microcirculation to macrocirculation.

There are many inherent limitations to our analyses that require comment. As previously discussed, significant heterogeneity was observed among studies that assessed endothelial function. Studies published in languages other than English were not included, and the quality of evidence for outcomes assessed in this meta-analysis was low-to-moderate. Subanalyses of microcirculatory and VSM data were limited because of the low number of studies assessing microvascular and VSM function in normoglycemic and acute hyperglycemic states. Furthermore, some studies used methods of assessing microcirculation that can be easily influenced by spatial variability and thus may limit results when assessing microvascular function. The risk of publication or other biases was detected when assessing the SMD in endothelial function. However, the quality of studies was evaluated by specific tools for the quality assessment of observational research,^{81,82} revealing a predominantly low-bias risk. It must be acknowledged that the ethnicity of the populations was poorly reported by studies included in this meta-analysis, and thus, conclusions on the effect of ethnicity cannot be drawn from these data. Finally, many studies using flow-mediated dilation as a method of assessing macrovascular endothelial function did not report shear stress. Therefore, it was not possible to comprehensively conclude whether macrovascular endothelial dysfunction found during acute hyperglycemia is because of intrinsic abnormalities of macrovascular endothelial function or if it is partially attributable to microvascular dysfunction and decreased stimulus for conduit artery dilation.⁷⁹

In conclusion, based on studies included in this meta-analysis, current evidence suggests that acute hyperglycemia decreases macrovascular endothelial function with no changes in microvascular endothelial function and systemic VSM function across healthy and cardiometabolic populations. This further supports endothelial dysfunction mediated by decreased NO availability as a primary mechanism in the pathogenesis of CVD, which may begin long before vascular remodeling is detectable or the onset of cardiometabolic diseases. Noting that microvascular data were limited, the microcirculatory system should therefore not be dismissed as a possible site of vascular dysfunction. Considering this, future studies should investigate the effects of sugar-sweetened beverage consumption on the underlying mechanisms of human vascular function at microvascular and macrovascular levels. These studies will provide a better understanding of how acute hyperglycemia induces vascular dysfunction and how it contributes to the pathogenesis of CVD from healthy to cardiometabolic populations.

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Disclosures

None.

References

- World Health Organisation. Diabetes fact sheet. <http://www.who.int/mediacentre/factsheets/fs312/en/>. November 22, 2014.
- Brown CM, Dulloo AG, Montani JP. Sugary drinks in the pathogenesis of obesity and cardiovascular diseases. *Int J Obes (Lond)*. 2008;32(suppl 6):S28–S34. doi: 10.1038/ijo.2008.204.
- Johnson RK, Appel LJ, Brands M, Howard BV, Lefevre M, Lustig RH, Sacks F, Steffen LM, Wylie-Rosett J; American Heart Association Nutrition Committee of the Council on Nutrition, Physical Activity, and Metabolism and the Council on Epidemiology and Prevention. Dietary sugars intake and cardiovascular health: a scientific statement from the American Heart Association. *Circulation*. 2009;120:1011–1020. doi: 10.1161/CIRCULATIONAHA.109.192627.
- Mathers CD, Loncar D. Projections of global mortality and burden of disease from 2002 to 2030. *PLoS Med*. 2006;3:e442. doi: 10.1371/journal.pmed.0030442.
- Malik AH, Akram Y, Shetty S, Malik SS, Yanchou Njike V. Impact of sugar-sweetened beverages on blood pressure. *Am J Cardiol*. 2014;113:1574–1580. doi: 10.1016/j.amjcard.2014.01.437.
- Malik VS, Pan A, Willett WC, Hu FB. Sugar-sweetened beverages and weight gain in children and adults: a systematic review and meta-analysis. *Am J Clin Nutr*. 2013;98:1084–1102. doi: 10.3945/ajcn.113.058362.
- Malik VS, Popkin BM, Bray GA, Després JP, Willett WC, Hu FB. Sugar-sweetened beverages and risk of metabolic syndrome and type 2 diabetes: a meta-analysis. *Diabetes Care*. 2010;33:2477–2483. doi: 10.2337/dc10-1079.
- Romaguera D, Norat T, Wark PA, et al. Consumption of sweet beverages and type 2 diabetes incidence in European adults: results from EPIC-INTERACT. *Diabetologia*. 2013;56:1520–1530.
- International Diabetes Federation. IDF worldwide definition of the metabolic syndrome. <http://www.idf.org/metabolic-syndrome>. December 10, 2014.
- World Health Organisation. Obesity and overweight fact sheet. <http://www.who.int/mediacentre/factsheets/fs311/en/>. December 10, 2014.
- Tominaga M, Eguchi H, Manaka H, Igarashi K, Kato T, Sekikawa A. Impaired glucose tolerance is a risk factor for cardiovascular disease, but not impaired fasting glucose. The Funagata Diabetes Study. *Diabetes Care*. 1999;22:920–924.
- Mah E, Bruno RS. Postprandial hyperglycemia on vascular endothelial function: mechanisms and consequences. *Nutr Res*. 2012;32:727–740. doi: 10.1016/j.nutres.2012.08.002.
- Lefèbvre PJ, Scheen AJ. The postprandial state and risk of cardiovascular disease. *Diabet Med*. 1998;15(suppl 4):S63–S68. doi: 10.1002/(SICI)1096-9136(1998120)15:4+<S63::AID-DIA737>3.0.CO;2-7.
- Souza EG, De Lorenzo A, Huguenin G, Oliveira GM, Tibiriçá E. Impairment of systemic microvascular endothelial and smooth muscle function in individuals with early-onset coronary artery disease: studies with laser speckle contrast imaging. *Coron Artery Dis*. 2014;25:23–28. doi: 10.1097/MCA.0000000000000055.
- Buchwald IB, Cacanyiova S, Neumann J, Samoilova VE, Boecker W, Kristek F. The role of arterial smooth muscle in vasorelaxation. *Biochem Biophys Res Commun*. 2008;377:504–507. doi: 10.1016/j.bbrc.2008.10.019.
- Karaca Ü, Schram MT, Houben AJ, Muris DM, Stehouwer CD. Microvascular dysfunction as a link between obesity, insulin resistance and hypertension. *Diabetes Res Clin Pract*. 2014;103:382–387. doi: 10.1016/j.diabres.2013.12.012.
- Celermajer DS. Endothelial dysfunction: does it matter? is it reversible? *J Am Coll Cardiol*. 1997;30:325–333.

18. Coutinho M, Gerstein HC, Wang Y, Yusuf S. The relationship between glucose and incident cardiovascular events. A metaregression analysis of published data from 20 studies of 95,783 individuals followed for 12.4 years. *Diabetes Care*. 1999;22:233–240.
19. Davis C, Fischer J, Ley K, Sarembock JJ. The role of inflammation in vascular injury and repair. *J Thromb Haemost*. 2003;1:1699–1709.
20. Sun J, Xu Y, Dai Z, Sun Y. Intermittent high glucose enhances proliferation of vascular smooth muscle cells by upregulating osteopontin. *Mol Cell Endocrinol*. 2009;313:64–69. doi: 10.1016/j.mce.2009.08.019.
21. Wang L, Zhu LH, Jiang H, Tang QZ, Yan L, Wang D, Liu C, Bian ZY, Li H. Grape seed proanthocyanidins attenuate vascular smooth muscle cell proliferation via blocking phosphatidylinositol 3-kinase-dependent signaling pathways. *J Cell Physiol*. 2010;223:713–726. doi: 10.1002/jcp.22080.
22. Arora S, Lidor A, Abularrage CJ, Weiswasser JM, Nylen E, Kellicut D, Sidawy AN. Thiamine (vitamin B1) improves endothelium-dependent vasodilatation in the presence of hyperglycemia. *Ann Vasc Surg*. 2006;20:653–658. doi: 10.1007/s10016-006-9055-6.
23. Bagg W, Whalley GA, Sathu A, Gamble G, Sharpe N, Braatvedt GD. The effect of acute hyperglycaemia on brachial artery flow mediated dilatation in normal volunteers. *Aust N Z J Med*. 2000;30:344–350.
24. Ballard KD, Mah E, Guo Y, Pei R, Volek JS, Bruno RS. Low-fat milk ingestion prevents postprandial hyperglycemia-mediated impairments in vascular endothelial function in obese individuals with metabolic syndrome. *J Nutr*. 2013;143:1602–1610. doi: 10.3945/jn.113.179465.
25. Baynard T, Carhart RL Jr, Weinstock RS, Ploutz-Snyder LL, Kanaley JA. Short-term exercise training improves aerobic capacity with no change in arterial function in obesity. *Eur J Appl Physiol*. 2009;107:299–308. doi: 10.1007/s00421-009-1126-2.
26. Ceriello A, Esposito K, Piconi L, Ihnat M, Thorpe J, Testa R, Bonfigli AR, Giugliano D. Glucose “peak” and glucose “spike”: impact on endothelial function and oxidative stress. *Diabetes Res Clin Pract*. 2008;82:262–267. doi: 10.1016/j.diabres.2008.07.015.
27. Ceriello A, Esposito K, Piconi L, Ihnat MA, Thorpe JE, Testa R, Boemi M, Giugliano D. Oscillating glucose is more deleterious to endothelial function and oxidative stress than mean glucose in normal and type 2 diabetic patients. *Diabetes*. 2008;57:1349–1354. doi: 10.2337/db08-0063.
28. Ceriello A, Esposito K, Testa R, Bonfigli AR, Marra M, Giugliano D. The possible protective role of glucagon-like peptide 1 on endothelium during the meal and evidence for an “endothelial resistance” to glucagon-like peptide 1 in diabetes. *Diabetes Care*. 2011;34:697–702. doi: 10.2337/dc10-1949.
29. Chittari MV, McTernan P, Bawazeer N, Constantinides K, Ciotola M, O’Hare JP, Kumar S, Ceriello A. Impact of acute hyperglycaemia on endothelial function and retinal vascular reactivity in patients with Type 2 diabetes. *Diabet Med*. 2011;28:450–454. doi: 10.1111/j.1464-5491.2010.03223.x.
30. De Marchi S, Prior M, Rigoni A, Zecchetto S, Rulfo F, Arosio E. Ascorbic acid prevents vascular dysfunction induced by oral glucose load in healthy subjects. *Eur J Intern Med*. 2012;23:54–57. doi: 10.1016/j.ejim.2011.07.019.
31. Dengel DR, Kelly AS, Steinberger J, Sinaiko AR. Effect of oral glucose loading on endothelial function in normal-weight and overweight children. *Clin Sci (Lond)*. 2007;112:493–498. doi: 10.1042/CS20060305.
32. Dye AS, Huang H, Bauer JA, Hoffman RP. Hyperglycemia increases muscle blood flow and alters endothelial function in adolescents with type 1 diabetes. *Exp Diabetes Res*. 2012;2012:170380. doi: 10.1155/2012/170380.
33. Fujimoto K, Hozumi T, Watanabe H, Tokai K, Shimada K, Yoshiyama M, Homma S, Yoshikawa J. Acute hyperglycemia induced by oral glucose loading suppresses coronary microcirculation on transthoracic Doppler echocardiography in healthy young adults. *Echocardiography*. 2006;23:829–834. doi: 10.1111/j.1540-8175.2006.00325.x.
34. Grasser EK, Yepuri G, Dulloo AG, Montani JP. Cardio- and cerebrovascular responses to the energy drink Red Bull in young adults: a randomized cross-over study. *Eur J Nutr*. 2014;53:1561–1571. doi: 10.1007/s00394-014-0661-8.
35. Grassi D, Desideri G, Necozione S, Ruggieri F, Blumberg JB, Stornello M, Ferri C. Protective effects of flavanol-rich dark chocolate on endothelial function and wave reflection during acute hyperglycemia. *Hypertension*. 2012;60:827–832. doi: 10.1161/HYPERTENSIONAHA.112.193995.
36. Ihlemann N, Rask-Madsen C, Perner A, Dominguez H, Hermann T, Køber L, Torp-Pedersen C. Tetrahydrobiopterin restores endothelial dysfunction induced by an oral glucose challenge in healthy subjects. *Am J Physiol Heart Circ Physiol*. 2003;285:H875–H882. doi: 10.1152/ajpheart.00008.2003.
37. Kato T, Inoue T, Node K. Postprandial endothelial dysfunction in subjects with new-onset type 2 diabetes: an acarbose and nateglinide comparative study. *Cardiovasc Diabetol*. 2010;9:12. doi: 10.1186/1475-2840-9-12.
38. Kawano H, Motoyama T, Hirashima O, Hirai N, Miyao Y, Sakamoto T, Kugiyama K, Ogawa H, Yasue H. Hyperglycemia rapidly suppresses flow-mediated endothelium-dependent vasodilation of brachial artery. *J Am Coll Cardiol*. 1999;34:146–154.
39. Kim SH, Park KW, Kim YS, Oh S, Chae IH, Kim HS, Kim CH. Effects of acute hyperglycemia on endothelium-dependent vasodilation in patients with diabetes mellitus or impaired glucose metabolism. *Endothelium*. 2003;10:65–70.
40. Lavi T, Karasik A, Koren-Morag N, Kanety H, Feinberg MS, Shechter M. The acute effect of various glycemic index dietary carbohydrates on endothelial function in nondiabetic overweight and obese subjects. *J Am Coll Cardiol*. 2009;53:2283–2287. doi: 10.1016/j.jacc.2009.03.025.
41. Mah E, Noh SK, Ballard KD, Matos ME, Volek JS, Bruno RS. Postprandial hyperglycemia impairs vascular endothelial function in healthy men by inducing lipid peroxidation and increasing asymmetric dimethylarginine:arginine. *J Nutr*. 2011;141:1961–1968. doi: 10.3945/jn.111.144592.
42. Mah E, Noh SK, Ballard KD, Park HJ, Volek JS, Bruno RS. Supplementation of a γ-tocopherol-rich mixture of tocopherols in healthy men protects against vascular endothelial dysfunction induced by postprandial hyperglycemia. *J Nutr Biochem*. 2013;24:196–203. doi: 10.1016/j.jnutbio.2012.04.015.
43. Nakayama H, Tsuge N, Sawada H, Higashi Y. Chronic intake of onion extract containing quercetin improved postprandial endothelial dysfunction in healthy men. *J Am Coll Nutr*. 2013;32:160–164. doi: 10.1080/07315724.2013.797858.
44. Napoli R, Guardasole V, Angelini V, Capasso AM, Zarra E, Cittadini A, Matarazzo M, Saccà L. Food and red wine do not exert acute effects on vascular reactivity. *Metabolism*. 2004;53:1081–1086.
45. Natali A, Baldi S, Vittone F, Muscelli E, Casolaro A, Morgantini C, Palombo C, Ferrannini E. Effects of glucose tolerance on the changes provoked by glucose ingestion in microvascular function. *Diabetologia*. 2008;51:862–871. doi: 10.1007/s00125-008-0971-6.
46. Shige H, Ishikawa T, Suzukawa M, Ito T, Nakajima K, Higashi K, Ayaori M, Tabata S, Ohsuzu F, Nakamura H. Endothelium-dependent flow-mediated vasodilation in the postprandial state in type 2 diabetes mellitus. *Am J Cardiol*. 1999;84:1272–4, A9.
47. Siafarikas A, Watts K, Beye P, Jones TW, Davis EA, Green DJ. Lack of effect of oral glucose loading on conduit vessel endothelial function in healthy subjects. *Clin Sci (Lond)*. 2004;107:191–196. doi: 10.1042/CS20040004.
48. Stirban A, Negrean M, Stratmann B, Gawlowski T, Horstmann T, Götting C, Kleesiek K, Mueller-Roesel M, Koschinsky T, Uribarri J, Vlassara H, Tschoepe D. Benfotiamine prevents macro- and microvascular endothelial dysfunction and oxidative stress following a meal rich in advanced glycation end products in individuals with type 2 diabetes. *Diabetes Care*. 2006;29:2064–2071. doi: 10.2337/dc06-0531.
49. Suzuki K, Watanabe K, Futami-Suda S, Yano H, Motoyama M, Matsumura N, Igari Y, Suzuki T, Nakano H, Oba K. The effects of postprandial glucose and insulin levels on postprandial endothelial function in subjects with normal glucose tolerance. *Cardiovasc Diabetol*. 2012;11:98. doi: 10.1186/1475-2840-11-98.
50. Title LM, Cummings PM, Giddens K, Nassar BA. Oral glucose loading acutely attenuates endothelium-dependent vasodilation in healthy adults without diabetes: an effect prevented by vitamins C and E. *J Am Coll Cardiol*. 2000;36:2185–2191.
51. Tushuizen ME, Nieuwland R, Scheffer PG, Sturk A, Heine RJ, Diamant M. Two consecutive high-fat meals affect endothelial-dependent vasodilation, oxidative stress and cellular microparticles in healthy men. *J Thromb Haemost*. 2006;4:1003–1010. doi: 10.1111/j.1538-7836.2006.01914.x.
52. Wang L, Guo L, Zhang L, Zhou Y, He Q, Zhang Z, Wang M. Effects of glucose load and nateglinide intervention on endothelial function and oxidative stress. *J Diabetes Res*. 2013;2013:849295. doi: 10.1155/2013/849295.
53. Watanabe K, Oba K, Suzuki T, Ouchi M, Suzuki K, Futami-Suda S, Sekimizu K, Yamamoto N, Nakano H. Oral glucose loading attenuates endothelial function in normal individual. *Eur J Clin Invest*. 2011;41:465–473. doi: 10.1111/j.1365-2362.2010.02424.x.
54. Weiss EP, Arif H, Villareal DT, Marzetti E, Holloszy JO. Endothelial function after high-sugar-food ingestion improves with endurance exercise performed on the previous day. *Am J Clin Nutr*. 2008;88:51–57.
55. Williams SB, Goldfine AB, Timimi FK, Ting HH, Roddy MA, Simonson DC, Creager MA. Acute hyperglycemia attenuates endothelium-dependent vasodilation in humans *in vivo*. *Circulation*. 1998;97:1695–1701.
56. Xiang GD, Sun HL, Hou J, Yue L, Xu L. Acute hyperglycemia rapidly suppresses endothelium-dependent arterial dilation in first-degree relatives of

- type 2 diabetic patients. *Exp Clin Endocrinol Diabetes*. 2008;116:112–117. doi: 10.1055/s-2007-984478.
57. Xiang GD, Sun HL, Zhao LS, Hou J, Yue L, Xu L. The antioxidant alpha-lipoic acid improves endothelial dysfunction induced by acute hyperglycaemia during OGTT in impaired glucose tolerance. *Clin Endocrinol (Oxf)*. 2008;68:716–723. doi: 10.1111/j.1365-2265.2007.03099.x.
 58. Zhang TX, Peng F, Chai DJ, Lin JX. Effects of combined glucose and fat load on endothelium-dependent brachial artery vasodilatation in hypertensive patients. *Am J Med Sci*. 2012;344:447–451. doi: 10.1097/MAJ.0b013e31824a0164.
 59. Zhang TX, Xu JX, Peng F, Chai DJ, Lin JX. Metformin reduces vascular endothelial dysfunction caused by an acute glucose load in patients with hypertension. *Blood Press*. 2013;22:106–113. doi: 10.3109/08037051.2012.732761.
 60. Zhu W, Zhong C, Yu Y, Li K. Acute effects of hyperglycaemia with and without exercise on endothelial function in healthy young men. *Eur J Appl Physiol*. 2007;99:585–591. doi: 10.1007/s00421-006-0378-3.
 61. Lee IK, Kim HS, Bae JH. Endothelial dysfunction: Its relationship with acute hyperglycaemia and hyperlipidemia. *Int J Clin Pract Suppl*. 2002;129:59–64.
 62. Bülow J, Astrup A, Christensen NJ, Kastrup J. Blood flow in skin, subcutaneous adipose tissue and skeletal muscle in the forearm of normal man during an oral glucose load. *Acta Physiol Scand*. 1987;130:657–661. doi: 10.1111/j.1748-1716.1987.tb08189.x.
 63. Forst T, Kunt T, Pohlmann T, Goitom K, Löbig M, Engelbach M, Beyer J, Pfützner A. Microvascular skin blood flow following the ingestion of 75 g glucose in healthy individuals. *Exp Clin Endocrinol Diabetes*. 1998;106:454–459. doi: 10.1055/s-0029-1212015.
 64. Riley RD, Higgins JP, Deeks JJ. Interpretation of random effects meta-analyses. *BMJ*. 2011;342:d549.
 65. Ma L, Zhao S, Li J, Zhou Q, Gao M. Interaction of hypertension and diabetes on impairment of endothelial function. *Chin Med J (Engl)*. 2001;114:563–567.
 66. Rosendorff C. Effects of LDL cholesterol on vascular function. *J Hum Hypertens*. 2002;16(suppl 1):S26–S28. doi: 10.1038/sj.jhh.1001337.
 67. Trinity JD, Groot HJ, Layec G, Rossman MJ, Ives SJ, Morgan DE, Gmelch BS, Bledsoe AD, Richardson RS. Passive leg movement and nitric oxide-mediated vascular function: The impact of age. *Am J Physiol Heart Circ Physiol*. 2015;308(6):H672–H679. doi: 10.1152/ajpheart.00806.2014.
 68. Muris DM, Houben AJ, Schram MT, Stehouwer CD. Microvascular dysfunction is associated with a higher incidence of type 2 diabetes mellitus: a systematic review and meta-analysis. *Arterioscler Thromb Vasc Biol*. 2012;32:3082–3094. doi: 10.1161/ATVBAHA.112.300291.
 69. Green DJ, Dawson EA, Groenewoud HM, Jones H, Thijsen DH. Is flow-mediated dilation nitric oxide mediated?: a meta-analysis. *Hypertension*. 2014;63:376–382. doi: 10.1161/HYPERTENSIONAHA.113.02044.
 70. Sandoo A, Carroll D, Metsios GS, Kitas GD, Veldhuijzen van Zanten JJ. The association between microvascular and macrovascular endothelial function in patients with rheumatoid arthritis: a cross-sectional study. *Arthritis Res Ther*. 2011;13:R99. doi: 10.1186/ar3374.
 71. Féleto M, Vanhoutte PM. Endothelium-derived hyperpolarizing factor: where are we now? *Arterioscler Thromb Vasc Biol*. 2006;26:1215–1225. doi: 10.1161/01.ATV.0000217611.81085.c5.
 72. Rousseau P, Mahé G, Haj-Yassin F, Durand S, Humeau A, Leftheriotis G, Abraham P. Increasing the “region of interest” and “time of interest”, both reduce the variability of blood flow measurements using laser speckle contrast imaging. *Microvasc Res*. 2011;82:88–91. doi: 10.1016/j.mvr.2011.03.009.
 73. Tew GA, Klonizakis M, Crank H, Briers JD, Hodges GJ. Comparison of laser speckle contrast imaging with laser Doppler for assessing microvascular function. *Microvasc Res*. 2011;82:326–332. doi: 10.1016/j.mvr.2011.07.007.
 74. Mitchell GF. Effects of central arterial aging on the structure and function of the peripheral vasculature: implications for end-organ damage. *J Appl Physiol* (1985). 2008;105:1652–1660. doi: 10.1152/japplphysiol.90549.2008.
 75. Montero D, Walther G, Pérez-Martin A, Vicente-Salar N, Roche E, Vinet A. Vascular smooth muscle function in type 2 diabetes mellitus: a systematic review and meta-analysis. *Diabetologia*. 2013;56:2122–2133. doi: 10.1007/s00125-013-2974-1.
 76. Akamatsu D, Sato A, Goto H, et al. Nitroglycerin-mediated vasodilatation of the brachial artery may predict long-term cardiovascular events irrespective of the presence of atherosclerotic disease. *J Atheroscler Thromb*. 2010;17:1266–1274.
 77. World Health Organisation. Cardiovascular diseases fact sheet. <http://www.who.int/mediacentre/factsheets/fs317/en/>. December 11, 2014.
 78. Ozkor MA, Rahman AM, Murrow JR, Kavtaradze N, Lin J, Manatunga A, Hayek S, Quyyumi AA. Differences in vascular nitric oxide and endothelium-derived hyperpolarizing factor bioavailability in blacks and whites. *Arterioscler Thromb Vasc Biol*. 2014;34:1320–1327. doi: 10.1161/ATVBAHA.113.303136.
 79. Mitchell GF, Parise H, Vita JA, Larson MG, Warner E, Keaney JF Jr, Keyes MJ, Levy D, Vasan RS, Benjamin EJ. Local shear stress and brachial artery flow-mediated dilation: the Framingham Heart Study. *Hypertension*. 2004;44:134–139. doi: 10.1161/01.HYP.0000137305.77635.68.
 80. Thijsen DH, Black MA, Pyke KE, Padilla J, Atkinson G, Harris RA, Parker B, Widlansky ME, Tschakovsky ME, Green DJ. Assessment of flow-mediated dilation in humans: a methodological and physiological guideline. *Am J Physiol Heart Circ Physiol*. 2011;300:H2–12. doi: 10.1152/ajpheart.00471.2010.
 81. Ross LE, Grigoriadis S, Mamiashvili L, Koren G, Steiner M, Dennis CL, Cheung A, Mousmanis P. Quality assessment of observational studies in psychiatry: an example from perinatal psychiatric research. *Int J Methods Psychiatr Res*. 2011;20:224–234. doi: 10.1002/mpr.356.
 82. Sprung VS, Atkinson G, Cuthbertson DJ, Pugh CJ, Aziz N, Green DJ, Cable NT, Jones H. Endothelial function measured using flow-mediated dilation in polycystic ovary syndrome: a meta-analysis of the observational studies. *Clin Endocrinol (Oxf)*. 2013;78:438–446. doi: 10.1111/j.1365-2265.2012.04490.x.

Significance

Acute hyperglycemia has previously been proposed to contribute to vascular dysfunction, which represents one of the main precursors to CVD. However, the effect of acute hyperglycemia on mechanisms of vascular function in humans is unclear because of discrepant results. Given this, we conducted the first systematic review and meta-analysis of its kind comparing endothelial function alone or in combination with VSM function during acute hyperglycemia in healthy and cardiometabolic diseased individuals. We demonstrate that acute hyperglycemia transiently impairs macrovascular endothelial function, which may represent, among others, a primary mechanism in the pathogenesis of CVD. This is significant when considering the surge in added sugar consumption in recent decades; humans are more often in a state of acute hyperglycemia and therefore are more frequently inducing endothelial dysfunction. This study provides the foundation for future research that will investigate the effect of acute hyperglycemia on underlying mechanisms of vascular function.

Acute hyperglycemia impairs flow-mediated dilatation through an increase in vascular oxidative stress: winter is coming for excess sugar consumption

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In two editorials recently published in the *Journal of Thoracic Disease*, Robert P. Hoffman and John David Horowitz *et al.* separately reviewed the interaction between acute hyperglycemia and vascular function (1,2) with a focus on the results of our meta-analysis published in *Arteriosclerosis, Thrombosis and Vascular Biology* (3). We thank the authors for their interest in our research and for their contribution to further understanding the effects of acute hyperglycemia on cardiovascular health.

Hoffman importantly highlighted that although our meta-analysis defined the methods used to assess vascular function, the primary outcome of each study wasn't clearly described. Indeed, it can be confirmed that for all studies included in the meta-analysis, the percentage increase from baseline measurement in response to a specific test of vascular reactivity was the primary outcome for both microvascular data (e.g., acetylcholine and sodium nitroprusside iontophoresis) and macrovascular data (e.g., flow- and nitrate-mediated dilation); and was used to determine standardized mean difference between vascular function in the acute hyperglycemic and normoglycemic states. In agreement with Hoffman, if there is a ceiling effect to the maximal vasodilatory capability of a blood vessel, then variations in baseline measurements due to the potential vasodilating effects of increased blood glucose or blood insulin concentrations during acute hyperglycemia would limit interpretation of the results when expressing vascular data solely as the percentage increase from baseline (1). Given

that only a few studies in the meta-analysis provided absolute values for baseline measurements of microcirculatory blood perfusion or brachial artery diameter, comparisons to detect differences in baseline data between the acute hyperglycemic and normoglycemic states were not possible. Such research deficiencies emphasize the need for future studies to clearly report absolute values of vascular function.

Further to this, Hoffman continued to address the potential confounding effects of the hyperinsulinemia that accompanies acute hyperglycemia. Indeed, insulin is a recognised vasodilator that contributes to vascular smooth muscle relaxation in an endothelium-dependent manner by stimulating the synthesis of nitric oxide via the PI3K/Akt pathway and the subsequent activation of endothelial nitric oxide synthase (eNOS) by phosphorylation at serine 1177 (4). Given that shear stress induces vasodilation through the same endothelium-dependent mechanism (5), it may be hypothesized that the impairment of the PI3K/Akt pathway that may be responsible for the acute hyperglycemia-mediated decrease in flow-mediated dilation may also cause a reduction in the vasodilatory action of insulin. Furthermore, it must be acknowledged that whilst blood glucose concentration increases rapidly following sugar consumption, increases in blood insulin concentration and its vasodilatory action are significantly delayed (6,7). Therefore, it is likely that the deleterious vascular effects of acute hyperglycemia may occur and be measured prior to any significant vasodilatory influence of insulin; moreover,

suggesting the redundancy of insulin's implication in potentially mediating heterogeneity between acute hyperglycemic and normoglycemic baseline measurements in vascular assessments performed soon after sugar consumption.

Considering that our meta-analysis highlighted the role of decreased nitric oxide bioavailability in acute hyperglycemia-mediated endothelial dysfunction, Horowitz *et al.* presented mechanisms that may contribute to impaired nitric oxide release (2). Nitric oxide synthesis is catalyzed by eNOS, which oxidizes L-arginine at its N-terminal oxygenase domain. However, L-arginine can also be converted to asymmetric N^G, N^G -dimethylarginine (ADMA) by protein arginine N methyltransferase (PRMT) (8) and arginase (9). The authors argue that elevated production of reactive oxygen species (ROS) during acute hyperglycemia may increase PRMT and arginase activity resulting in decreased bioavailability of L-arginine and increased ADMA. In addition to limiting substrate availability required for nitric oxide synthesis, ADMA directly competes with arginine for eNOS binding sites, thereby decreasing nitric oxide bioavailability. An increase in ROS (oxidative stress) during acute hyperglycemia may also impair eNOS activity by oxidizing its essential co-factor, tetrahydrobiopterin (BH_4) to dihydrobiopterin (BH_2) (10). Such elevations in BH_2 concentration decrease the binding of BH_4 to the active site of eNOS, compounding the superoxide generation (11) that reduces nitric oxide bioavailability and subsequently impairs endothelial function.

Given that it is now clearly established that acute hyperglycemia induces transient oxidative stress that is responsible for endothelial dysfunction (12), there is a great interest in approaches that increase antioxidant defenses that can prevent endothelial dysfunction. Physical activity is one such method that is known to stimulate antioxidant mechanisms, which may enhance eNOS coupling and eNOS activation by phosphorylation at serine 1177 (13). Nevertheless, further experimental and clinical studies are needed to explore the ability of exercise training to prevent oxidative stress and the eNOS uncoupling phenomenon occurring during acute hyperglycemia.

In conclusion, our meta-analysis provided evidence that acute hyperglycemia induces endothelial dysfunction. Due to limited availability of microcirculatory studies, this effect was contained to the macrocirculation. However, further research is needed to clearly establish that acute hyperglycemia-mediated endothelial dysfunction might also occur in the microcirculation. Given that added

sugar consumption has increased dramatically in recent decades, especially in children, highlights the importance of conducting such research that will inform public health policy on the role of excess sugar consumption in the pathogenesis of cardiovascular disease.

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Footnote

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Hoffman RP. Hyperglycemic endothelial dysfunction: does it happen and does it matter? *J Thorac Dis* 2015;7:1693-5.

References

1. Hoffman RP. Hyperglycemic endothelial dysfunction: does it happen and does it matter? *J Thorac Dis* 2015;7:1693-5.
2. Horowitz JD, Chong CR, Ngo DT, *et al.* Effects of acute hyperglycaemia on cardiovascular homeostasis: does a spoonful of sugar make the flow-mediated dilatation go down? *J Thorac Dis* 2015;7:E607-11.
3. Loader J, Montero D, Lorenzen C, *et al.* Acute Hyperglycemia Impairs Vascular Function in Healthy and Cardiometabolic Diseased Subjects: Systematic Review and Meta-Analysis. *Arterioscler Thromb Vasc Biol* 2015;35:2060-72.
4. Muniyappa R, Quon MJ. Insulin action and insulin resistance in vascular endothelium. *Curr Opin Clin Nutr Metab Care* 2007;10:523-30.
5. Fleming I, Fisslthaler B, Dixit M, *et al.* Role of PECAM-1 in the shear-stress-induced activation of Akt and the endothelial nitric oxide synthase (eNOS) in endothelial cells. *J Cell Sci* 2005;118:4103-11.
6. Fugmann A, Lind L, Andersson PE, *et al.* The effect of euglucaemic hyperinsulinaemia on forearm blood flow

- and glucose uptake in the human forearm. *Acta Diabetol* 1998;35:203-6.
7. Tack CJ, Schefman AE, Willems JL, et al. Direct vasodilator effects of physiological hyperinsulin-aemia in human skeletal muscle. *Eur J Clin Invest* 1996;26:772-8.
 8. Wu G, Morris SM Jr. Arginine metabolism: nitric oxide and beyond. *Biochem J* 1998;336:1-17.
 9. Pope AJ, Karuppiah K, Cardounel AJ. Role of the PRMT-DDAH-ADMA axis in the regulation of endothelial nitric oxide production. *Pharmacol Res* 2009;60:461-5.
 10. Crabtree MJ, Smith CL, Lam G, et al. Ratio of 5,6,7,8-tetrahydrobiopterin to 7,8-dihydrobiopterin in endothelial cells determines glucose-elicited changes in NO vs. superoxide production by eNOS. *Am J Physiol Heart Circ Physiol* 2008;294:H1530-40.
 11. Vásquez-Vivar J, Martásek P, Whitsett J, et al. The ratio between tetrahydrobiopterin and oxidized tetrahydrobiopterin analogues controls superoxide release from endothelial nitric oxide synthase: an EPR spin trapping study. *Biochem J* 2002;362:733-9.
 12. Monnier L, Mas E, Ginet C, et al. Activation of oxidative stress by acute glucose fluctuations compared with sustained chronic hyperglycemia in patients with type 2 diabetes. *JAMA* 2006;295:1681-7.
 13. Farah C, Kleindienst A, Bolea G, et al. Exercise-induced cardioprotection: a role for eNOS uncoupling and NO metabolites. *Basic Res Cardiol* 2013;108:389.

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**Hyperglycaemia and perivascular adipose tissue, two triggers in vascular dysfunction:
Impact of oxidative stress-eNOS pathway and effect of exercise training.**

The globalization of the western diet has mediated prevalence in cardiovascular disease related mortality, the single leading cause of death worldwide. Considering this, it is imperative that the underlying mechanisms of cardiovascular dysfunctions are continually investigated to establish a greater understanding of its pathogenesis from a healthy state to the presence of cardiometabolic diseases; and to improve upon current treatment and preventative strategies. Therefore, the **first aim** of this research was to identify vascular impact of acute hyperglycaemic stress induced by sweet sugar beverage consumption, with a translational approach. The results of this study demonstrated that consumption of a single commercially available sugar-sweetened beverage (SSB) induced transient micro- and macrovascular endothelial dysfunction, even in a healthy population. Further exploration into the underlying mechanisms of SSB-mediated endothelial dysfunction indicated that an increase in oxidative stress disrupts normal function of the nitric oxide pathway. Although disturbances in cardiovascular function may initially be transient, repetitive acute metabolic stress may translate to chronic cardiometabolic disease. Therefore, the **second aim** of this research was to assess the impact of a chronic metabolic disorder, metabolic syndrome (MetS), on vascular function in a rat model. Despite increasing sympathetic activity, the MetS rats didn't present elevated arterial pressure. Such findings may be explained by a compensatory adaptation of endothelial function that increases production of nitric oxide in response to α -adrenergic agonist and, thus, regulates arterial pressure despite sympathetic hyperactivity. Considering this, the **third aim** of this research evaluated the impact of perivascular adipose tissue (PVAT) on vascular function in MetS rats; demonstrating that MetS altered the adiponectin-endothelial nitric oxide synthase pathway in PVAT, in an oxidative stress-dependant manner. Exercise training is well recognized as a non-pharmacological strategy that has a beneficial impact on both metabolic and cardiovascular disorders via an improvement in function of the nitric oxide pathway. Considering this, research also assessed the efficacy of this approach to prevent vascular injury induced by acute hyperglycaemia in a healthy population and by PVAT in those with MetS. It was demonstrated that exercise attenuated acute hyperglycemia-mediated endothelial dysfunction; and restored endothelium-dependent vascular reactivity in rats with MetS, due to an improvement in the biocommunication between PVAT and arterial tissue and a notable enhancement of the adiponectine-endothelial nitric oxide synthase pathway.

Hyperglycémie et tissu adipeux, deux acteurs de la dysfonction vasculaire : Implication du couple stress oxydant – eNOS et modulation par l'exercice physique.

Les troubles métaboliques caractéristiques d'une alimentation de type « Western diet », sont à l'origine de pathologies cardiovasculaires, première cause de mortalité dans le monde. Il apparaît nécessaire d'améliorer la compréhension des mécanismes impliqués dans l'installation des dysfonctions cardiovasculaires afin de pouvoir proposer des stratégies thérapeutiques ou préventives adaptées. Ainsi, **le premier objectif de la thèse** a été d'évaluer les effets d'une boisson sucrée sur la fonction vasculaire macro- et microcirculatoire chez des sujets sains, par une approche translationnelle allant de la clinique humaine à un modèle expérimental de rongeur. Nos résultats montrent une altération de la fonction endothéliale en réponse à une prise de boisson sucrée, dans l'ensemble des lits vasculaires. L'exploration des mécanismes sous-jacents ces altérations nous a permis d'identifier l'implication du couple stress-oxydant/voie du NO. **Un second objectif de thèse**, a été d'étudier l'impact d'un stress métabolique chronique sur la fonction vasculaire et son incidence sur la régulation de la pression artérielle. Comme observé chez certains sujets souffrant de syndrome métabolique, notre modèle de rat ne présentait pas d'hypertension artérielle, malgré une hyperactivité du système sympathique. Ceci semble être expliqué par une compensation endothéliale eNOS-dépendant, qui permet de garantir le maintien d'une pression artérielle normale en dépit de l'effet vasopresseur adrénnergique élevé. Le **troisième objectif de thèse** a porté sur un nouvel élément participant au maintien de l'homéostasie vasculaire et impacté par les situations pathologiques : le tissu adipeux périvasculaire (PVAT). Nos travaux démontrent dans le contexte du SMet, une altération de la voie adiponectine/eNOS dans le PVAT, en parallèle d'une augmentation de la production d'espèces oxygénées réactives. La pratique régulière d'un exercice physique est aujourd'hui reconnue comme une stratégie non-pharmacologique permettant d'impacter à la fois les désordres métaboliques et cardiovasculaires, notamment via une amélioration de la voie du NO. Nos résultats démontrent une limitation de l'apparition des dysfonctions endothéliales causée par une hyperglycémie aigüe lorsqu'un protocole d'exercice physique chronique est réalisé. Enfin, l'exercice physique permet également de prévenir les modifications des propriétés vasoactives du PVAT dans un modèle de rat SMet. Ce phénomène pourrait être expliqué par une amélioration du statut oxydant de la paroi artérielle, et à une potentialisation de la voie adiponectine/eNOS par l'exercice physique.