CHAPITRE III

A New Labdane Diterpene from the Flowers of Solidago canadensis

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Résumé

Le Solidago canadensis L. est une plante herbacée de la famille des Astéracées. En Amérique du Nord, les fleurs de cette plante étaient fréquemment utilisées en médecine traditionnelle amérindienne. Dans le but de valoriser la biomasse de la forêt boréale par l'étude de l'activité biologique de composés d'origine naturelle, un extrait de fleurs de Solidago canadensis a été étudié. Un nouveau diterpène de type labdane, le 9α,16ξ-dihydroxy-6-oxo-7,13-labdadièn-15,16-olide (solicanolide, 1) et six composés connus : la quercétine (2), l'acide 3-O-caffeoylquinique (3, acide néochlorogénique), l'acide 5-O-caffeoylquinique (4, acide chlorogénique), l'acide 4,5-di-O-caffeoylquinique (5), l'acide 3,5-di-O-caffeoylquinique (6) et l'acide 3,4-di-O-caffeoylquinique (7) ont été isolés. Le composé 7 (acide 3,4-di-O-caffeoylquinique) est rapporté pour la première fois à partir du Solidago canadensis. Cet article décrit l'isolation des composés 1-7 et l'élucidation de la structure du nouveau composé (1). L'évaluation de l'activité anticancéreuse de tous les composés isolés est décrite. Parmi tous les composés isolés, seul le composé 1 possède une activité cytotoxique.

Notes

A New Labdane Diterpene from the Flowers of Solidago canadensis

Marie-Eve Bradette-Hébert, Jean Legault, Serge Lavoie, and André Pichette*

LASEYE, Département des Sciences Fondamentales, Université du Québec à Chicoutimi; 555 Boul. Université, Chicoutimi, Québec, G7H 2B1, Canada. Received June 28, 2007; accepted September 25, 2007

A new labdane diterpene, $9\alpha,16\xi$ -dihydroxy-6-oxo-7,13-labdadien-15,16-olide (solicanolide, 1) and six known compounds identified as quercetin (2), 3-O-caffeoylquinic acid (3, neochlorogenic acid), 5-O-caffeoylquinic acid (4, chlorogenic acid), 4.5-di-O-caffeoylquinic acid (5), 3.5-di-O-caffeoylquinic acid (6) and 3,4-di-O-caffeoylquinic acid (7) were isolated from the flowers of Solidago canadensis. To our knowledge, compound 7 was isolated for the first time in S. canadensis. This work describes the isolation of compounds 1—7 and the structure elucidation of a new compound identified as compound 1. Solicanolide (1) showed cytotoxic activity against A549 (IC₅₀: $13\pm2~\mu\rm M$), DLD-1 (IC₅₀: $26\pm2~\mu\rm M$) and WS1 (IC₅₀: $17\pm1~\mu\rm M$) cell lines.

Key words Solidago canadensis; isolation; labdane; solicanolide; cytotoxicity; caffeoylquinic acid

Solidago canadensis L. is an Asteraceae widely distributed across North America, occurring in almost every state of the U.S.A. and throughout Canada. Numerous interesting secondary metabolites such as: flavonoids, phenolic acids and glucosides, polysaccharides, diterpenes, triterpenoid saponosides, tannins and essential oils¹⁾ were reported for the genus Solidago. Previous phytochemical studies of S. canadensis have led to the isolation of flavonoids,^{2,3)} phenolic acids,⁴⁾ sesquiterpenes,⁵⁾ diterpenes^{6,7)} and saponins.⁷⁾ The flowers of S. canadensis were used in traditional Amerindian medicine as an analgesic,⁸⁾ burns and ulcers treatment,⁹⁾ febrifuge,¹⁰⁾ gastrointestinal^{11,12)} and liver¹¹⁾ aids. In spite of the widespread use of S. canadensis, few investigations were carried out on its bioactive secondary metabolites.

Fractionation of the flower extracts of Solidago canadensis resulted in the isolation of a new diterpene, 9α , 16ξ -dihydroxy-6-oxo-7,13-labdadien-15,16-olide (solicanolide, 1) and six known compounds identified as quercetin (2), 3-O-caffeoylquinic acid (neochlorogenic acid) (3), 5-O-caffeoylquinic acid (chlorogenic acid) (4), 4,5-di-O-caffeoylquinic acid (5), 3,5-di-O-caffeoylquinic acid (6) and 3,4-di-O-caffeoylquinic acid (7). To our knowledge, compound O-caffeoylquinic acid (7). To our knowledge, compound the solitation of compounds O-caffeoylquinic acid (7). To our knowledge, compound the solitation of a new compound identified as compound 1. The cytotoxicity of solicanolide (1) was also investigated in this paper.

Experimental

General Experimental Procedures NMR spectra were recorded in methanol-d4 on a Bruker Avance 400 spectrometer (5 mm QNP with Z-gradient probe) operating at 400.13 MHz (1H) or 100.61 MHz (13C). Chemical shifts were referenced relative to the corresponding residual solvent signals $(\delta_{\mathrm{HC}} \ 3.31/49.0 \, \mathrm{ppm}, \, \mathrm{respectively}).$ The accurate mass determination was carried out with an Applied Biosystems QSTAR XL Hybrid LC/MS/MS system. Optical rotation was obtained on a Jasco DIP-360 digital polarimeter. Analytical HPLC-DAD-MS analysis were performed on an Agilent 1100 series HPLC-DAD-MS system. A Zorbax ODS C18 column (5 µm, 150×4.6 mm) maintained at 25 °C was utilised. The flow rate was 1 ml/min. Agilent G1315B diode array detector was used for UV detection. The UV spectra were recorded from 190 to 400 nm. An Agilent mass selective detector (VL model) equipped with an atmospheric chemical ionisation source (APCI) was employed for MS detection. All mass spectra were acquired in the negative ion mode. The full scan mass spectrum was recorded over the range of m/z 100-1000. Temperature of the drying gas (N2) was 350 °C with a gas flow rate of 10 l/min and a nebulizing pressure of 40 psi. The ionisation voltage was 4000 V and the corona current was 15 mA. All HPLC separations were performed on a preparative Agilent 1100 series (Agilent Technologies Canada Inc.) with a ZORBAX ODS column C18 (2.1×25 cm; 7µm) at a flow rate of 16 ml/min. Compounds were detected by UV absorption at 254 nm. For all HPLC procedures, solvent A was H₂O+0.1% HCOOH, solvent B was MeOH+0.1% HCOOH and solvent C was ACN+0.1% HCOOH. Silica gel 60 (230—400 mesh, Silicycle), Diaion HP-20 (Supelco) and C18 silica gel (230—400 mesh, EMD) were used for column chromatography. The solvents were purchased from EMD. Thin-layer chromatography (TLC) was performed on silica gel 60 F₂₅₄ aluminium sheets (Silicycle) using EtOAc-MeOH-H₂O (100:16.5:13.5) or CHCl₃-MeOH (10:1) as solvent systems. Detection of the phenolic compounds was carried out by spraying natural products with NP/PEG reagent (1%

 $R^1 = caffeoyl, R^2 = R^3 = H$ $R^1 = R^2 = H, R^2 = caffeoyl$

 $R^{\dagger}=R^{2}=caff coyl,\,R^{3}\equiv H$

 $R^1 = R^3 = \text{caffwyl}, R^2 = H$

7 $R^1 = H$, $R^2 = R^3 = caffeoyl$

Fig. 1. Structures of Compounds 1---7

OH H₃C H₃ O CH₃

Fig. 2. ¹H-¹H NOESY Correlations of Compound 1

diphenylboric acid 2-aminoethylester in methanol and 5% polyethylene glycol in ethanol) and observing under UV light before and after revelation (254, 365 nm). Quercetin and chlorogenic acid standards are from Sigma-Aldrich

Plant Material The flowers of Solidago canadensis L. were collected in September 2004 in St-Félicien (48°40'N, 72°27'W), Québec, Canada. The specimens were identified by Mr. Patrick Nadeau (Département des Sciences Fondamentales, Université du Québec à Chicoutimi, Québec, Canada). A voucher specimen was deposited in the Louis-Marie Herbarium of Université Laval, Québec, Canada (QFA-492115).

Extraction and Isolation The air-dried flowers of Solidago canudensis (285 g) were extracted with hot methanol (3×2.51) and then with hot methanol 80% (3×2.51) for 2h. The combined extracts were evaporated under a reduced pressure, yielding a residue (103.81 g), which was suspended in water (400 ml) and successively extracted with dichloromethane $(3\times200 \text{ ml})$ and ethyl acetate $(6\times400 \text{ ml})$, yielding dichloromethane, ethyl acetate and aqueous fractions.

The ethyl acetate fraction (15.6 g) was separated into two fractions (A, B) by chromatography over Diaion (3.2×40.5 cm) eluring with MeOH-H₂O (from 50 to 100%). Fraction B (3.0 g) was chromatographed on a silica gel column (3.6×120.0 cm) with CHCl₃-MeOH (10:1) to obtain six fractions (B1-B6). From B1 (91.4 mg), compound 1 (5.4 mg) was purified by preparative HPLC (40% solvent A, 60% solvent B). Fractions B5 and B6 were combined and then fractionated on a silica gel column (3.0×45.0 cm) with CHCl₃-MeOH (10:1) to give compound 2 (53.6 mg).

The aqueous fraction (66.13 g) was separated into three fractions (C, D, E) by chromatography on a silica gel column (7.0×60.0 cm) using an isocratic solvent system of EtOAc-MeOH-H₂O (100:16.5:13.5). Fraction D (6.70 g), which contained the major constituents, was divided into six fractions (D1—D6) using Diaion (3.2×40.5 cm) by elution with MeOH 20%. Fraction D2 (1.03 g) was separated into nine fractions (D2A—D2I) by chromatography on a C18 silica gel column (1.6×46.0 cm) and eluted with MeOH 10%. From D2A (214.2 mg), compounds 3 (3.8 mg) and 4 (9.3 mg) were purified by preparative HPLC (94% solvent A, 6% solvent C). Fractions D2E (135.1 mg) and D2F (64.9 mg) were combined and then, fractionated by preparative HPLC (84% solvent A, 16% solvent C) to yield compounds 5 (9.1 mg), 6 (7.0 mg) and 7 (11.4 mg).

Solicanolide (1): Yellow oil: $(\alpha)_{15}^{15}$ – 30.1° (c=0.1. MeOH); UV λ_{max} (MeOH) 216 nm; HR-ES1-MS m/z 371.1834 [M+Na]⁺ (Calcd for $C_{20}H_{28}O_{5}$ Na, 371.1842): ¹H- and ¹³C-NMR: see Table 1.

Cell Culture The human lung carcinoma A549 (#CCL-185), colorectal adenocarcinoma DLD-1 (#CCl-221) and skin fibroblast WS1 (#CRL-1502) cell lines were all obtained from the American Type Culture Collection (ATCC, Manassas, U.S.A.). Cells lines were grown in minimum essential medium containing Earle's salts (Mediatoch Cellgro*, Herndon, U.S.A.), supplemented with 10% fetal calf serum (Hyclone, Logan, U.S.A.), 1×solution of vitamins, 1×sodium pyruvate, 1×non-essential amino acids, 1001U of penicillin and 100 µg/ml of streptomycin (Mediatoch Cellgro*). Cells were cultured at 37 °C in a humidified atmosphere containing 5% COs.

Cytotoxic Assay Exponentially growing cells were plated at a density of 5×10³ cells per well in 96-well microplates (BD Falcon) in 100 µl of culture medium and were allowed to adhere for 16 h before treatment. The cells were then incubated for 48 h in the presence or absence of 100 µl of increasing concentrations of compounds 1—7 dissolved in culture medium and an appropriate solvent. The final concentration of solvent in the culture medium was maintained at 0.25% (v/v) to avoid toxicity. Cytotoxicity was assessed using the resazurin reduction test. ¹³ Fluorescence was measured on an automated 96-well Fluoroskan Ascent FlTM plate reader (Labsystems) using an excitation wavelength of 530 mm and an emission wavelength of 590 nm. Cytotoxicity was expressed as the concentration of drug inhibiting cell growth by 50% (ICcs.).

Results and Discussion

Compound 1 was obtained as an optical active yellow oil $([\alpha]_D^{25}-30.1^\circ)$. The molecular formula of compound 1 was established as $C_{20}H_{28}O_5$ on the basis of HR-ESI-MS (Found at m/z 371.1834 [M+Na]⁺, Calcd for $C_{20}H_{28}O_5Na$, 371.1842). 1H -, ^{13}C -NMR and DEPT spectra revealed four methyls, five methylenes, four methines including two olefinics and seven quaternary carbons including a carbonyl and a quaternary alcohol (Table 1). The NMR spectra sug-

Table 1. NMR Assignments of Compound 1

Position	13Ca)	Connected ¹ H ³)	H-H COSY ³	HMBC _q)	NOESY ²⁰
1	32.6 (t)	1.92 (m), 1.57 (m)		H5, 20	
2	19.0 (t)	1.66 (m), 1.53 (m)	H3		
3	43.9 (1)	1.35 (m), 1.19 (m)	H2	H18, 19	
4	33.3 (s)	70000		H5, 18, 19	
5	56.8 (d)	2.88 (s)	H20	H18, 19, 20	H18
6	202.9 (s)			H5	
7	129.6 (d)	5.66 (s)	H17	H17	H17
8	159.1 (s)	******		Н17	
9	76.7 (s)			H5, 7, 17, 20	
10	48.1 (s)	marks.		H5, 20	
11	31.0 (n)	2.11 (m), 1.94 (m)			H14, 19, 20
12	25.4 (t)	2.65 (m)			
13	172.8 (s)				1114, 16
14	117.6 (d)	5.95 (s)		H16	H12, 14
15	173.7 (s)			H14, 16	
16	101.1 (d)	6.06 (s)		H14	H12
17	20.5 (q)	2.02 (s)	H7	H7	H7
18	34,3 (q)	1.14(s)		H19	H5
19	22.2 (q)	1.18 (s)		143, 5, 18	H11, 20
20	18.8 (q)	1.02 (s)	H5	H5	H11, 19

a) Multiplicities were determined by DEPT.
b) Connections were determined by gRSQC.
c) Determined by gCOSY.
d) Correlation from C to the indicated protons.
e) unambiguous NOESY cross peaks.

gests that compound 1 is a labdane diterpene. (4,15)

HMBC correlations between H-17 ($\delta_{\rm H}$ 2.02) and C-7, C-8, C-9 ($\delta_{\rm C}$ 129.6, 159.1, 76.7 respectively) showed that the methyl group was branched β on an α, β -insaturated ketone and that the quaternary alcohol group was on C-9. The position of the ketone was further confirmed by the HMBC correlation between H-5 ($\delta_{\rm H}$ 2.88) and C-6 ($\delta_{\rm C}$ 202.9). The side chain was identified as a γ -hydroxy- α, β -unsaturated- γ -lactone by the presence of typical 16 1H ($\delta_{\rm H}$ 5.95, 6.06) and 13 C-NMR signals ($\delta_{\rm C}$ 101.1, 117.6, 172.8, 173.7).

The relative stereochemistry of compound 1 was partially determined from the 2D NOESY spectrum. Cross-peaks between H-11 ($\delta_{\rm H}$ 1.94, 2.11), H-19 ($\delta_{\rm H}$ 1.18) and H-20 ($\delta_{\rm H}$ 1.02) showed that all these groups are positioned on the same side. Unfortunately, NOESY correlations between H-16 ($\delta_{\rm H}$ 6.06) and both H-11 ($\delta_{\rm H}$ 1.94, 2.11) were observed with the same intensity which prevents the determination of the stereochemistry of the hydroxyl function at C-16. However, while some authors observed both epimers at H-16 of related compounds, ^{17,18}) its single signal in the ¹H- and ¹³C-NMR spectra suggests that only one epimer was isolated.

The determination of the absolute stereochemistry by the Mosher ester method was precluded by the limited amount of this natural product. However, the chemotaxonomic information indicated that all labdanes from *Solidago* genus belong to the normal series^{5,19,20)} and by analogy, compound 1 could be assigned to this series. Therefore, the structure of compound 1 was identified as $9\alpha,16\xi$ -dihydroxy-6-oxo-7,13-labdadien-15,16-olide.

The other compounds isolated were identified as quercetin (2), 3-O-caffeoylquinic acid (3, neochlorogenic acid), 5-O-caffeoylquinic acid (4, chlorogenic acid), 4.5-di-O-caffeoylquinic acid (5), 3.5-di-O-caffeoylquinic acid (6) and 3,4-di-O-caffeoylquinic acid (7). Identification of these compounds was established by NMR (¹H, ¹³C, DEPT, COSY, HSQC and HMBC) and HR-ESI-MS analyses. The data ob-

Communicati	IC ₅₀ (μ _M)			
Compound -	A549	DLD-1	WSI	
Etoposide	1.1±0.1	4.8±0.8	n.đ.	
5-Fluorouracil	4.8 ± 0.6	11±2	n.đ.	
1	13±2	26±2	17±1	
2	>199	>199	>199	
3	>57.5	>57.5	>57.5	
4	>56	>56	>56	
5	>109	>109	>109	
ó	>107	>107	>107	
7	>36.8	>36.8	>36.8	

Data are expressed as means±standard deviations of three determinations; n.d.=not determined.

tained were compared with those reported in the literature^{21,22)} and standards. To our knowledge, this is the first report of the isolation of compound 7 from *Solidago canadensis*.

All isolated compounds were evaluated for their *in vitro* cytotoxic activity against human lung cancer A549, colon cancer DLD-1 and normal fibroblasts, WS1 cell lines. Etoposide and 5-fluorouracil was used as positive controls. As presented in Table 2, only compound 1 was found to be active against A549 (IC₅₀: $13\pm2~\mu\text{M}$), DLD-1 (IC₅₀: $26\pm2~\mu\text{M}$) and WS1 (IC₅₀: $17\pm1~\mu\text{M}$) cell lines.

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CHAPITRE IV

Antioxidant and anti-inflammatory activities of methanol extracts from Solidago canadensis

Article en préparation pour Journal of Ethnopharmacology.

Résumé

Le Solidago canadensis L. est une plante herbacée de la famille des Astéracées. En Amérique du Nord, cette plante était utilisée en médecine traditionnelle amérindienne pour traiter la douleur, les brûlures, les ulcères, la fièvre et les troubles du foie et de l'intestin. Dans le but de valoriser la biomasse de la forêt boréale par l'étude de l'activité biologique de composés d'origine naturelle, des extraits de fleurs, de feuilles et de tiges de Solidago canadensis ont été étudiés. Les extraits méthanoliques de fleurs, de feuilles et de tiges ont montré des propriétés antioxydantes avec des valeurs ORAC de 2.7 ± 0.9 μmol TE/mg pour les fleurs, $10.0 \pm 0.2 \,\mu\text{mol}$ TE/mg pour les feuilles et $3.6 \pm 0.8 \,\mu\text{mol}$ TE/mg pour les tiges. L'extrait méthanolique de fleurs a aussi montré une activité anti-inflammatoire en inhibant la production d'oxyde nitrique de macrophages RAW 264.7 activés avec du LPS (IC₅₀: 8 ± 2 μg/mL). Cependant, les extraits de feuilles et de tiges sont inactifs. Le fractionnement de l'extrait de fleurs a mené à l'isolation de cinq dérivés de l'acide caffeoylquinique connus. L'acide 3,4-di-O-caffeoylquinique est rapporté pour la première fois dans le Solidago canadensis. L'activité anti-inflammatoire des composés isolés a été évaluée, mais ces composés sont inactifs.